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(54) Title: MORPHOGEN-INDUCED MODULATION OF INFLAMMATORY RESPONSE

#### (57) Abstract

The present invention is directed to methods and compositions for alleviating tissue destructive effects associated with the inflammatory response to tissue injury in a mammal. The methods and compositions include administering a therapeutically effective concentration of a morphogen or morphogen-stimulating agent sufficient to alleviate immune cell-mediated tissue destruction.

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# MORPHOGEN-INDUCED MODULATION OF INFLAMMATORY RESPONSE

#### Field of the Invention

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The present invention relates generally to a method for modulating the inflammatory response induced in a mammal following tissue injury. More particularly, this invention relates to a method for alleviating immune-cell mediated tissue destruction associated with the inflammatory response.

#### Background of the Invention

The body's inflammatory response to tissue injury 20 can cause significant tissue destruction, leading to loss of tissue function. Damage to cells resulting from the effects of inflammatory response e.g., by immune-cell mediated tissue destruction, has been implicated as the cause of reduced tissue function or loss of tissue function in diseases of the joints 25 (e.g., rheumatoid and osteo-arthritis) and of many organs, including the kidney, pancreas, skin, lung and heart. For example, glomular nephritis, diabetes, inflammatory bowel disease, vascular diseases such as 30 atheroclerosis and vasculitis, and skin diseases such as psoriasis and dermatitis are believed to result in large part from unwanted acute inflammatory reaction A number of these diseases, including and fibrosis. arthritis, psoriasis and inflammatory bowel disease are 35 considered to be chronic inflammatory diseases.

damaged tissue also often is replaced by fibrotic tissue, e.g., scar tissue, which further reduces tissue function. Graft and transplanted organ rejection also is believed to be primarily due to the action of the body's immune/inflammatory response system.

The immune-cell mediated tissue destruction often follows an initial tissue injury or insult. The secondary damage, resulting from the inflammatory 10 response, often is the source of significant tissue damage. Among the factors thought to mediate these damaging effects are those associated with modulating the body's inflammatory response following tissue injury, e.g., cytokines such as interleukin-1 (IL-1) 15 and tumor necrosis factor (TNF), and oxygen-derived free radicals such as superoxide anions. These humoral agents are produced by adhering neutrophilic leukocytes or by endothelial cells and have been identified at ischemic sites upon reperfusion. Moreover, TNF 20 concentrations are increased in humans after myocardial infarction.

A variety of lung diseases are characterized by airway inflammation, including chronic bronchitis, emphysema, idiopathic pulmonary fibrosis and asthma. Another type of lung-related inflammation disorders are inflammatory diseases characterized by a generalized, wide-spread acute inflammatory response such as adult respiratory distress syndrome. Another dysfunction associated with the inflammatory response is that mounted in response to injury caused by hyperoxia, e.g., prolonged exposure to lethally high concentrations of 02 (95-100% 02). Similarly, reduced

blood flow to a tissue (and, therefore reduced or lack of oxygen to tissues), as described below, also can induce a primary tissue injury that stimulates the inflammatory response.

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It is well known that damage occurs to cells in mammals which have been deprived of oxygen. the interruption of blood flow, whether partial (hypoxia) or complete (ischemia) and the ensuing 10 inflammatory responses may be the most important cause of coagulative necrosis or cell death in human disease. The complications of atherosclerosis, for example, are generally the result of ischemic cell injury in the brain, heart, small intestines, kidneys, and lower 15 extremities. Highly differentiated cells, such as the proximal tubular cells of the kidney, cardiac myocytes, and the neurons of the central nervous system, all depend on aerobic respiration to produce ATP, the energy necessary to carry out their specialized 20 functions. When ischemia limits the oxygen supply and ATP is depleted, the affected cells may become irreversibly injured. The ensuing inflammatory responses to this initial injury provide additional insult to the affected tissue. Examples of such 25 hypoxia or ischemia are the partial or total loss of blood supply to the body as a whole, an organ within the body, or a region within an organ, such as occurs in cardiac arrest, pulmonary embolus, renal artery occlusion, coronary occlusion or occlusive stroke.

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The tissue damage associated with ischemiareperfusion injury is believed to comprise both the
initial cell damage induced by the deprivation of
oxygen to the cell and its subsequent recirculation, as
well as the damage caused by the body's response to

this initial damage. It is thought that reperfusion injury may result in dysfunction to the endothelium of the vasculature as well as injury to the surrounding tissue. In idiopathic pulmonary fibrosis, for example, scar tissue accumulates on the lung tissue lining, inhibiting the tissue's elasticity. The tissue damage associated with hyperoxia injury is believed to follow a similar mechanism, where the initial damage is mediated primarily through the presence of toxic oxygen metabolites followed by an inflammatory response to this initial injury.

Similarly, tissues and organs for transplantation also are subject to the tissue destructive effects

15 associated with the recipient host body's inflammatory response following transplantation. It is currently believed that the initial destructive response is due in large part to reperfusion injury to the transplanted organ after it has been transplanted to the organ recipient.

Accordingly, the success of organ or tissue transplantation depends greatly on the preservation of the tissue activity (e.g., tissue or organ viability)

25 at the harvest of the organ, during storage of the harvested organ, and at transplantation. To date, preservation of organs such as lungs, pancreas, heart and liver remains a significant stumbling block to the successful transplantation of these organs. U.S.

30 Patent No. 4,952,409 describes a superoxide dismutase-containing liposome to inhibit reperfusion injury. U.S. Patent No. 5,002,965 describes the use of ginkolides, known platelet activating factor antagonists, to inhibit reperfusion injury. Both of these factors are described working primarily by

inhibiting the release of and/or inhibiting the damaging effects of free oxygen radicals. A number of patents also have issued on the use of immunosuppressants for inhibiting graft rejection. A representative listing includes U.S. Patent Nos. 5,104,858, 5,008,246 and 5,068,323. A significant problem with many immunosuppressants is their low therapeutic index, requiring the administration of high doses that can have significant toxic side effects.

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Rheumatoid and osteoarthritis are prevalent diseases characterized by chronic inflammation of the synovial membrane lining the afflicted joint. A major consequence of chronic inflammatory joint disease (e.g., rheumatoid arthritis) and degenerative arthritis 15 (e.g., osteoarthritis) is loss of function of those affected joints. This loss of function is due primarily to destruction of the major structural components of the joint, cartilage and bone, and subsequent loss of the proper joint anatomy. As a consequence of chronic disease, joint destruction ensues and can lead to irreversible and permanent damage to the joint and loss of function. Current treatment methods for severe cases of rheumatoid 25 arthritis typically include the removal of the synovial membrane, e.g., synovectomy. Surgical synovectomy has many limitations, including the risk of the surgical procedure itself, and the fact that a surgeon often cannot remove all of the diseased membrane. 30 diseased tissue remaining typically regenerates, causing the same symptoms which the surgery was meant to alleviate.

Psoriasis is a chronic, recurrent, scaling skin disease of unknown etiology characterized by chronic inflammation of the skin. Erythematous eruptions, often in papules or plaques, and usually having a white silvery scale, can affect any part of the skin, but most commonly affect the scalp, elbows, knees and lower back. The disease usually occurs in adults, but children may also be affected. Patients with psoriasis have a much greater incidence of arthritis (psoraitic arthritis), and generalized exfoliation and even death can threaten afflicted individuals.

Current therapeutic regimens include topical or intralesional application of corticosteroids, topical administration of keratolytics, and use of tar and UV light on affected areas. No single therapy is ideal, and it is rare for a patient not to be treated with several alternatives during the relapsing and remitting course of the disease. Whereas systematic treatment can induce prompt resolution of psoriatic lesions, suppression often requires ever-increasing doses, sometimes with toxic side effect, and tapering of therapy may result in rebound phenomena with extensions of lesions, possibly to exfoliation.

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Inflammatory bowel disease (IBD) describes a class of clinical disorders of the gastrointestinal mucosa characterized by chronic inflammation and severe ulceration of the mucosa. The two major diseases in this classification are ulcerative colitis and regional enteritis (Crohn's Disease). Like oral mucositis, the diseases classified as IBD are associated with severe mucosal ulceration (frequently penetrating the wall of the bowel and forming strictures and fistulas), severe mucosal and submucosal inflammation and edema, and

fibrosis (e.g., scar tissue formation which interferes with the acid protective function of the gastrointestinal lining.) Other forms of IBD include regional ileitis and proctitis. Clinically, patients with fulminant IBD can be severely ill with massive diarrhea, blood loss, dehydration, weight loss and fever. The prognosis of the disease is not good and frequently requires resection of the diseased tissue.

Therefore, an object of the present invention is to 10 provide a method for protecting mammalian tissue, particularly human tissue, from the damage associated with the inflammatory response following a tissue injury. The inflammatory reaction may be in response 15 to an initial tissue injury or insult. The original injury may be chemically, mechanically, biologically or immunologically related. Another object is to provide methods and compositions for protecting tissue from the tissue destructive effects associated with chronic 20 inflammatory diseases, including arthritis (e.g., reheumatoid or osteoarthritis), psoriatic arthritis, psoriasis and dermatitis, inflammatory bowel disease and other autoimmune diseases. Yet another object is to provide methods and compositions for enhancing the 25 viability of mammalian tissues and organs to be transplanted, including protecting the transplanted organs from immune cell-mediated tissue destruction, such as the tissue damage associated with ischemiareperfusion injury. This tissue damage may occur 30 during donor tissue or organ harvesting and transport, as well as following initiation of blood flow after transplantation of the organ or tissue in the recipient host.

Another object of the invention is to provide a method for alleviating tissue damage associated with ischemic-reperfusion injury in a mammal following a deprivation of oxygen to a tissue in the mammal. Other objects of the present invention include providing a method for alleviating tissue damage associated with ischemic-reperfusion injury in a human which has suffered from hypoxia or ischemia following cardiac arrest, pulmonary embolus, renal artery occlusion, coronary occlusion or occlusive stroke. A further object is to provide a method for alleviating tissue damage associated with hyperoxia-induced tissue injury, e.g., lethally high oxygen concentrations.

- Still another object of the invention is to provide a method for modulating inflammatory responses in general, particularly those induced in a human following tissue injury.
- These and other objects and features of the invention will be apparent from the description, drawings and claims which follow.

#### Summary of the Invention

The present invention provides a method for alleviating the tissue destructive effects associated with activation of the inflammatory response following tissue injury. The method comprises the step of providing to the affected tissue a therapeutically effective concentration of a morphogenic protein ("morphogen", as defined herein) upon tissue injury or in anticipation of tissue injury, sufficient to substantially inhibit or reduce the tissue destructive effects of the inflammatory response.

In one aspect, the invention features compositions
and therapeutic treatment methods that comprise the
step of administering to a mammal a therapeutically
effective amount of a morphogenic protein
("morphogen"), as defined herein, upon injury to a
tissue, or in anticipation of such injury, for a time
and at a concentration sufficient to inhibit the tissue
destructive effects associated with the body's
inflammatory response, including repairing damaged
tissue, and/or inhibiting additional damage thereto.

25 In another aspect, the invention features compositions and therapeutic treatment methods for protecting tissues and organs from the tissue destructive effects of the inflammatory response which include administering to the mammal, upon injury to a tissue or in anticipation of such injury, a compound that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen within the body of the mammal sufficient to protect the tissue from the tissue destructive effects associated with the inflammatory response, including repairing damaged

tissue and/or inhibiting additional damage thereto.

These compounds are referred to herein as morphogenstimulating agents, and are understood to include
substances which, when administered to a mammal, act on
cells of tissue(s) or organ(s) that normally are
responsible for, or capable of, producing a morphogen
and/or secreting a morphogen, and which cause the
endogenous level of the morphogen to be altered. The
agent may act, for example, by stimulating expression
and/or secretion of an endogenous morphogen.

As embodied herein, the term "ischemic-reperfusion injury" refers to the initial damage associated with oxygen deprivation of a cell and the subsequent damage 15 associated with the inflammatory response when the cell is resupplied with oxygen. As embodied herein, the term "hyperoxia-induced injury" refers to the tissue damage associated with prolonged exposure to lethally high doses of oxygen, e.g., greater than 95%  $0_2$ , including the tissue damage associated with the inflammatory response to the toxically high oxygen dose. Accordingly, as used herein, "toxic oxygen concentrations" refers to the tissue damage associated withthe injury induced by both lethally low oxygen concentrations of oxygen (including a complete lack of oxygen), and by lethally high oxygen concentrations. The expression "alleviating" means the protection from, reduction of and/or elimination of undesired tissue destruction, particularly immune cell-mediated tissue destruction. The tissue destruction may be in response to an initial tissue injury, which may be mechanical, chemical or immunological in origin. The expression "enhance the viability of" living tissues or organs, as used herein, means protection from, reduction of and/or elimination of reduced or lost tissue or organ function as a result of tissue death, particularly immune cellmediated tissue death. "Transplanted" living tissue
encompasses both tissue transplants (e.g., as in the
case of bone marrow transplants) and tissue grafts.

5 Finally, a "free oxygen radical inhibiting agent" means
a molecule capable of inhibiting the release of and/or
inhibiting tissue damaging effects of free oxygen
radicals.

In one embodiment of the invention, the invention 10 provides methods and compositions for alleviating the ischemic-reperfusion injury in mammalian tissue resulting from a deprivation of, and subsequent reperfusion of, oxygen to the tissue. In another 15 embodiment, the invention provides a method for alleviating the tissue-destructive effects associated with hyperoxia. In still another embodiment of the invention, the invention provides methods and compositions for maintaining the viability of tissues 20 and organs, particularly living tissues and organs to be transplanted, including protecting them from ischemia-reperfusion injury. In still another embodiment, the invention provides methods for protecting tissues and organs from the tissue destructive effects of chronic inflammatory diseases, such as arthritis, psoriasis, dermatitis, including contact dermatitis, IBD and other chronic inflammatory diseases of the gastrointestinal tract, as well as the tissue destructive effects associated with other, known 30 autoimmune diseases, such as diabetes, multiple sclerosis, amyotrophic lateral sclerosis (ALS), and other autoimmune neurodegenerative diseases.

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In one aspect of the invention, the morphogen is provided to the damaged tissue following an initial injury to the tissue. The morphogen may be provided directly to the tissue, as by injection to the damaged tissue site or by topical administration, or may be provided indirectly, e.g., systemically by oral or parenteral means. Alternatively, as described above, an agent capable of stimulating endogenous morphogen expression and/or secretion may be administered to the mammal. Preferably, the agent can stimulate an endogenous morphogen in cells associated with the damaged tissue. Alternatively, morphogen expression and/or secretion may be stimulated in a distant tissue and the morphogen transported to the damaged tissue by the circulatory system.

In another aspect of the invention, the morphogen is provided to tissue at risk of damage due to immune cell-mediated tissue destruction. Examples of such tissues include tissue grafts and tissue or organ transplants, as well as any tissue or organ about to undergo a surgical procedure or other clinical procedure likely to either inhibit blood flow to the tissue or otherwise induce an inflammatory response.

Here the morphogen or morphogen-stimulating agent preferably is provided to the patient prior to induction of the injury, e.g., as a prophylactic, to provide a cyto-protective effect to the tissue at risk.

organ to be transplanted, the tissue or organ to be transplanted preferably is exposed to a morphogen prior to transplantation. Most preferably, the tissue or organ is exposed to the morphogen prior to its removal from the donor, by providing the donor with a

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composition comprising a morphogen or morphogenstimulating agent. Alternatively or, in addition, once
removed from the donor, the organ or tissue is placed
in a preservation solution containing a morphogen or
morphogen-stimulating agent. In addition, the
recipient also preferably is provided with a morphogen
or morphogen-stimulating agent just prior to, or
concommitant with, transplantation. In all cases, the
morphogen or morphogen-stimulating agent may be
administered directly to the tissue at risk, as by
injection or topical administration to the tissue, or
it may be provided systemically, either by oral or
parenteral administration.

15 The morphogens described herein are envisioned to be useful in enhancing viability of any organ or living tissue to be transplanted. The morphogens may be used to particular advantage in lung, heart, liver, kidney or pancreas transplants, as well as in transplantation and/or grafting of bone marrow, skin, gastrointestinal mucosa, and other living tissues.

Where the patient suffers from a chronic inflammatory disease, such as diabetes, arthritis, psoriasis, IBD, and the like, the morphogen or morphogen-stimulating agent preferably is administered at regular intervals as a prophylactic, to prevent and/or inhibit the tissue damage normally associated with the disease during flare periods. As above, the morphogen or morphogen-stimulating agent may be provided directly to the tissue at risk, for example by injection or by topical administration, or indirectly, as by systemic e.g., oral or parenteral administration.

Among the morphogens useful in this invention are proteins originally identified as osteogenic proteins, such as the OP-1, OP-2 and CBMP2 proteins, as well as amino acid sequence-related proteins such as DPP (from 5 Drosophila), Vgl (from Xenopus), Vgr-1 (from mouse, see U.S. 5,011,691 to Oppermann et al.), GDF-1 (from mouse, see Lee (1991) PNAS 88:4250-4254), all of which are presented in Table II and Seq. ID Nos.5-14), and the recently identified 60A protein (from Drosophila, Seq. 10 ID No. 24, see Wharton et al. (1991) PNAS 88:9214-9218.) The members of this family, which include members of the TGF-β super-family of proteins, share substantial amino acid sequence homology in their The proteins are translated as a C-terminal regions. 15 precursor, having an N-terminal signal peptide sequence, typically less than about 30 residues, followed by a "pro" domain that is cleaved to yield the mature sequence. The signal peptide is cleaved rapidly upon translation, at a cleavage site that can be predicted in a given sequence using the method of Von Heijne ((1986) Nucleic Acids Research 14:4683-4691.) Table I, below, describes the various morphogens identified to date, including their nomenclature as used herein, their Seq. ID references, and publication sources for the amino acid sequences for the full 25 length proteins not included in the Seq. Listing. disclosure of these publications is incorporated herein by reference.

#### TABLE I

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"OP-1" Refers generically to the group of morphogenically active proteins expressed from part or all of a DNA sequence encoding OP-1 protein, including allelic and species variants thereof, e.g., human

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OP-1 ("hOP-1", Seq. ID No. 5, mature protein amino acid sequence), or mouse OP-1 ("mOP-1", Seq. ID No. 6, mature protein amino acid sequence.) conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID The cDNA sequences and the Nos. 5 and 6. amino acids encoding the full length proteins are provided in Seq. Id Nos. 16 and 17 (hOP1) and Seq. ID Nos. 18 and 19 (mOP1.) The mature proteins are defined by residues 293-431 (hOP1) and 292-430 (mOP1). The "pro" regions of the proteins, cleaved to yield the mature, morphogenically active proteins are defined essentially by residues 30-292 (hOP1) and residues 30-291 (mOP1).

"OP-2"

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refers generically to the group of active proteins expressed from part or all of a DNA sequence encoding OP-2 protein, including allelic and species variants thereof, e.g., human OP-2 ("hOP-2", Seq. ID No. 7, mature protein amino acid sequence) or mouse OP-2 ("mOP-2", Seq. ID No. 8, mature protein amino acid sequence). The conserved seven cysteine skeleton is defined by residues 38 to 139 of Seq. ID Nos. 7 and 8. The cDNA sequences and the amino acids encoding the full length proteins are provided in Seq. ID Nos. 20 and 21 (hOP2) and Seq. ID Nos. 22 and 23 (mOP2.) The mature proteins are defined essentially by residues 264-402 (hOP2) and 261-399 (mOP2). The "pro"

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"CBMP2"

regions of the proteins, cleaved to yield the mature, morphogenically active proteins likely are defined essentially by residues 18-263 (hOP2) and residues 18-260 (mOP2). (Another cleavage site also occurs 21 residues upstream for both OP-2 proteins.) refers generically to the morphogenically active proteins expressed from a DNA sequence encoding the CBMP2 proteins, including allelic and species variants thereof, e.g., human CBMP2A ("CBMP2A(fx)", Seq ID No. 9) or human CBMP2B DNA ("CBMP2B(fx)", Seq. ID No. 10). The amino acid sequence for the full length proteins, referred to in the literature as BMP2A and BMP2B, or BMP2 and BMP4, appear in Wozney, et al. (1988) Science 242:1528-

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"DPP(fx)"

293-408.

refers to protein sequences encoded by the Drosophila DPP gene and defining the conserved seven cysteine skeleton (Seq. ID No. 11). The amino acid sequence for the full length protein appears in Padgett, et

The pro domain for BMP2 (BMP2A)

likely includes residues 25-248 or 25-282; the mature protein, residues 249-396 or 283-396. The pro domain for BMP4 (BMP2B) likely includes residues 25-256 or 25-292; the mature protein, residues 257-408 or al (1987) <u>Nature 325</u>: 81-84. The prodomain likely extends from the signal peptide cleavage site to residue 456; the mature protein likely is defined by residues 457-588.

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"Vgl(fx)" refers to protein sequences encoded by the Xenopus Vgl gene and defining the conserved seven cysteine skeleton (Seq. ID No. 12). The amino acid sequence for the full length protein appears in Weeks (1987) Cell 51: 861-867. The prodomain likely extends from the signal peptide cleavage site to residue 246; the mature protein likely is defined by residues 247-360.

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"Vgr-1(fx)" refers to protein sequences encoded by the murine Vgr-1 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 13). The amino acid sequence for the full length protein appears in Lyons, et al, (1989) PNAS 86: 4554-4558. The prodomain likely extends from the signal peptide cleavage site to residue 299; the mature protein likely is defined by residues 300-438.

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"GDF-1(fx)" refers to protein sequences encoded by the human GDF-1 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 14). The cDNA and encoded amino sequence for the full length protein is

5		provided in Seq. ID. No. 32. The prodomain likely extends from the signal peptide clavage site to residue 214; the mature protein likely is defined by residues 215-372.
	"60A"	refers generically to the morphogenically active proteins expressed from part or all of a DNA sequence (from the Drosophila 60A
10		gene) encoding the 60A proteins (see Seq. ID No. 24 wherein the cDNA and encoded amino acid sequence for the full length protein is provided). "60A(fx)" refers to the protein sequences defining the
15		conserved seven cysteine skeleton (residues 354 to 455 of Seq. ID No. 24.) The prodomain likely extends from the signal peptide cleavage site to residue 324; the mature protein likely is defined
20		by residues 325-455.
25	"BMP3(fx)"	refers to protein sequences encoded by the human BMP3 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 26). The amino acid sequence for the full
25		length protein appears in Wozney et al.  (1988) <u>Science</u> 242: 1528-1534. The pro domain likely extends from the signal peptide cleavage site to residue 290; the
30		mature protein likely is defined by

residues 291-472.

"BMP5(fx)" refers to protein sequences encoded by the human BMP5 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 27).

The amino acid sequence for the full length protein appears in Celeste, et al.

(1991) PNAS 87: 9843-9847. The pro domain likely extends from the signal peptide cleavage site to residue 316; the mature protein likely is defined by residues

317-454.

"BMP6(fx)" refers to protein sequences encoded by the human BMP6 gene and defining the conserved seven cysteine skeleton (Seq. ID No. 28).

The amino acid sequence for the full length protein appears in Celeste, et al. (1990) PNAS 87: 9843-5847. The pro domain likely includes extends from the signal peptide cleavage site to residue 374; the mature sequence likely includes residues 375-513.

The OP-2 proteins have an additional cysteine

residue in this region (e.g., see residue 41 of Seq. ID

Nos. 7 and 8), in addition to the conserved cysteine
skeleton in common with the other proteins in this
family. The GDF-1 protein has a four amino acid insert
within the conserved skeleton (residues 44-47 of Seq.

ID No. 14) but this insert likely does not interfere
with the relationship of the cysteines in the folded
structure. In addition, the CBMP2 proteins are missing
one amino acid residue within the cysteine skeleton.

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The morphogens are inactive when reduced, but are active as oxidized homodimers and when oxidized in combination with other morphogens of this invention Thus, as defined herein, a (e.g., as heterodimers). 5 morphogen is a dimeric protein comprising a pair of polypeptide chains, wherein each polypeptide chain comprises at least the C-terminal six cysteine skeleton defined by residues 43-139 of Seq. ID No. 5, including functionally equivalent arrangements of these cysteines 10 (e.g., amino acid insertions or deletions which alter the linear arrangement of the cysteines in the sequence but not their relationship in the folded structure), such that, when the polypeptide chains are folded, the dimeric protein species comprising the pair of 15 polypeptide chains has the appropriate threedimensional structure, including the appropriate intraor inter-chain disulfide bonds such that the protein is capable of acting as a morphogen as defined herein. Specifically, the morphogens generally are capable of all of the following biological functions in a 20 morphogenically permissive environment: stimulating proliferation of progenitor cells; stimulating the differentiation of progenitor cells; stimulating the proliferation of differentiated cells; and supporting 25 the growth and maintenance of differentiated cells, including the "redifferentiation" of transformed cells. In addition, it is also anticipated that these morphogens are capable of inducing redifferentiation of committed cells under appropriate environmental 30 conditions.

In one preferred aspect, the morphogens of this invention comprise one of two species of generic amino acid sequences: Generic Sequence 1 (Seq. ID No. 1) or 35 Generic Sequence 2 (Seq. ID No. 2); where each Xaa

indicates one of the 20 naturally-occurring L-isomer,
 α-amino acids or a derivative thereof. Generic
 Sequence 1 comprises the conserved six cysteine
 skeleton and Generic Sequence 2 comprises the conserved
 six cysteine skeleton plus the additional cysteine
 identified in OP-2 (see residue 36, Seq. ID No. 2). In
 another preferred aspect, these sequences further
 comprise the following additional sequence at their N terminus:

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Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)

Preferred amino acid sequences within the 15 foregoing generic sequences include: Generic Sequence 3 (Seq. ID No. 3), Generic Sequence 4 (Seq. ID No. 4), Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31), listed below. Generic Sequences accommodate the homologies shared among the various preferred members of this morphogen 20 family identified in Table II, as well as the amino acid sequence variation among them. Specifically, Generic Sequences 3 and 4 are composite amino acid sequences of the following proteins presented in Table II and identified in Seq. ID Nos. 5-14: human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP (from Drosophila, Seq. ID 30 No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14.) The generic sequences include both the amino acid identity shared by the sequences in Table II, as well as alternative residues for the variable positions within the sequence. Note that

these generic sequences allow for an additional cysteine at position 41 or 46 in Generic Sequences 3 or 4, respectively, providing an appropriate cysteine skeleton where inter- or intramolecular disulfide bonds can form, and contain certain critical amino acids which influence the tertiary structure of the proteins.

## Generic Sequence 3

Leu Tyr Val Xaa Phe

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Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

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Xaa Ala Pro Xaa Gly Xaa Xaa Ala

15 20

15 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa

25 30

Xaa Pro Xaa Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Asn His Ala Xaa Xaa

20 40 45

Xaa Xaa Leu Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Xaa Xaa Xaa Cys

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25 Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa

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Xaa Xaa Xaa Leu Xaa Xaa Xaa

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Xaa Xaa Xaa Xaa Val Xaa Leu Xaa

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5 Xaa Xaa Xaa Xaa Met Xaa Val Xaa

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Xaa Cys Gly Cys Xaa

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wherein each Xaa is independently selected from a group 10 of one or more specified amino acids defined as follows: "Res." means "residue" and Xaa at res.4 = (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser or Lys); Xaa at res.7 = (Asp or Glu); Xaa at res.8 = (Leu or Val); Xaa at res.11 = (Gln, Leu, Asp, His or Asn); 15 Xaa at res.12 = (Asp, Arg or Asn); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.18 |= (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Leu or Gln); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at 20 res.26 = (Glu, His, Tyr, Asp or Gln); Xaa at res.28 = (Glu, Lys, Asp or Gln); Xaa at res.30 = (Ala, Ser, Pro or Gln); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu or Val); Xaa at res.34 = (Asn, Asp, Ala or Thr); Xaa at res.35 = (Ser, Asp, Glu, Leu or Ala); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn or Ser); Xaa at res.39 = (Ala, Ser or Gly); Xaa at res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile or Val); Xaa at res.45 = (Val or Leu); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.49 = (Val or Met); Xaa at res.50 = (His or Asn); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa

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at res.52 = (Ile, Met, Asn, Ala or Val); Xaa at res.53 = (Asn, Lys, Ala or Glu); Xaa at res.54 = (Pro or Ser); Xaa at res.55 = (Glu, Asp, Asn, or Gly); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at 5 res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys or Leu); Xaa at res.60 = (Pro or Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr or Ala); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser or Asp); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr or Val); Xaa at res.71 = (Ser or Ala); Xaa at res.72 = (Val or Met); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 = (Phe, Tyr or Leu); Xaa at res.76 = (Asp or Asn); Xaa at res.77 = 15 (Asp, Glu, Asn or Ser); Xaa at res.78 = (Ser, Gln, Asn or Tyr); Xaa at res.79 = (Ser, Asn, Asp or Glu); Xaa at res.80 = (Asn, Thr or Lys); Xaa at res.82 = (Ile or Val); Xaa at res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln or His); Xaa at res.86 = (Tyr or His); 20 Xaa at res.87 = (Arg, Gln or Glu); Xaa at res.88 = (Asn, Glu or Asp); Xaa at res.90 = (Val, Thr or Ala); Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at res.93 = (Ala, Gly or Glu); and Xaa at res.97 = (His or Arg);

Generic Sequence 4

 Cys Xaa Xaa Xaa Xaa Leu Tyr Val Xaa Phe

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 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa
 15

 Xaa Ala Pro Xaa Gly Xaa Xaa Ala
 20
 25

 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa
 35

Xaa Pro Xaa Xaa Xaa Xaa Xaa 40

Xaa Xaa Xaa Asn His Ala Xaa Xaa

5 Xaa Xaa Leu Xaa Xaa Xaa Xaa Xaa 55

Xaa Xaa Xaa Xaa Xaa Xaa Cys 60 65

Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa

10 70

Xaa Xaa Xaa Leu Xaa Xaa Xaa 75 80

Xaa Xaa Xaa Xaa Val Xaa Leu Xaa 85

15 Xaa Xaa Xaa Xaa Met Xaa Val Xaa 90 95

> Xaa Cys Gly Cys Xaa 100

wherein each Xaa is independently selected from a group 20 of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys or Arg); Xaa at res.3 = (Lys or Arg); Xaa at res.4 = (His or Arg); Xaa at res.5 = (Glu, Ser, His, Gly, Arg or Pro); Xaa at res.9 = (Ser, Asp or Glu); Xaa at 25 res.11 = (Arg, Gln, Ser or Lys); Xaa at res.12 = (Asp or Glu); Xaa at res.13 = (Leu or Val); Xaa at res.16 = (Gln, Leu, Asp, His or Asn); Xaa at res.17 = (Asp, Arg, or Asn); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro 30 or Arg); Xaa at res.25 = (Tyr or Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Leu, or Gln); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp or Gln); Xaa at res.33 = Glu, Lys, Asp or Gln); Xaa at res.35 = (Ala, Ser or Pro); Xaa at res.36 = (Phe, Leu or Tyr); Xaa at res.38 = (Leu or Val); Xaa at res.39 =

(Asn, Asp, Ala or Thr); Xaa at res.40 = (Ser, Asp, Glu, Leu or Ala); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.44 = (Ala, Ser or Gly); Xaa at res.45 = (Thr, Leu 5 or Ser); Xaa at res.49 = (Ile or Val); Xaa at res.50 = (Val or Leu); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.54 = (Val or Met); Xaa at res.55 = (His or Asn); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, 10 Met, Asn, Ala or Val); Xaa at res.58 = (Asn, Lys, Ala or Glu); Xaa at res.59 = (Pro or Ser); Xaa at res.60 = (Glu, Asp, or Gly); Xaa at res.61 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser or Ala); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = 15 (Lys or Leu); Xaa at res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr or Ala); Xaa at res.71 = (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser or Asp); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = 20 (Ile, Thr or Val); Xaa at res.76 = (Ser or Ala); Xaa at res.77 = (Val or Met); Xaa at res.79 = (Tyr or Phe); Xaa at res.80 = (Phe, Tyr or Leu); Xaa at res.81 = (Asp or Asn); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn or Tyr); Xaa at res.84 = (Ser, 25 Asn, Asp or Glu); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile or Val); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln or His); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln or Glu); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 30 = (Val, Thr or Ala); Xaa at res.97 = (Arg, Lys, Val, Asp or Glu); Xaa at res.98 = (Ala, Gly or Glu); and Xaa at res.102 = (His or Arg).

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Similarly, Generic Sequence 5 (Seq. ID No. 30) and Generic Sequence 6 (Seq. ID No. 31) accommodate the homologies shared among all the morphogen protein family members identified in Table II. Specifically, 5 Generic Sequences 5 and 6 are composite amino acid sequences of human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 (Seq. ID Nos. 7, 8, and 20-22), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), DPP 10 (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), and GDF-1 (from mouse, Seq. ID No. 14), human BMP3 (Seq. ID No. 26), human BMP5 (Seq. ID No. 27), human BMP6 (Seq. ID No. 28) and 60(A) (from Drosophila, Seq. ID Nos. 24-25). The generic sequences include both the amino acid identity shared by these sequences in the C-terminal domain, defined by the six and seven cysteine skeletons (Generic Sequences 5 and 6, respectively), as well as alternative residues for the 20 variable positions within the sequence. As for Generic Sequences 3 and 4, Generic Sequences 5 and 6 allow for an additional cysteine at position 41 (Generic Sequence 5) or position 46 (Generic Sequence 6), providing an appropriate cysteine skeleton where inter- or 25 intramolecular disulfide bonds can form, and containing certain critical amino acids which influence the tertiary structure of the proteins.

#### Generic Sequence 5

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Leu Xaa Xaa Xaa Phe

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Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa

Xaa Xaa Pro Xaa Xaa Xaa Xaa Ala 20 15 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 30 25 Xaa Pro Xaa Xaa Xaa Xaa Xaa 5 35 Xaa Xaa Xaa Asn His Ala Xaa Xaa 45 40 Xaa Xaa Xaa Xaa Xaa Xaa Xaa 50 10 Xaa Xaa Xaa Xaa Xaa Xaa Cys 60 55 Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa 65 Xaa Xaa Xaa Leu Xaa Xaa Xaa 15 75 70 Xaa Xaa Xaa Xaa Val Xaa Leu Xaa 80 Xaa Xaa Xaa Met Xaa Val Xaa 90 85 20 Xaa Cys Xaa Cys Xaa

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wherein each Xaa is independently selected from a group of one or more specified amino acids defined as

25 follows: "Res." means "residue" and Xaa at res.2 =

(Tyr or Lys); Xaa at res.3 = Val or Ile); Xaa at res.4

= (Ser, Asp or Glu); Xaa at res.6 = (Arg, Gln, Ser, Lys or Ala); Xaa at res.7 = (Asp, Glu or Lys); Xaa at res.8

= (Leu, Val or Ile); Xaa at res.11 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.12 = (Asp, Arg, Asn or Glu); Xaa at res.14 = (Ile or Val); Xaa at res.15 = (Ile or Val); Xaa at res.16 (Ala or Ser); Xaa at res.18 5 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.19 = (Gly or Ser); Xaa at res.20 = (Tyr or Phe); Xaa at res.21 = (Ala, Ser, Asp, Met, His, Gln, Leu or Gly); Xaa at res.23 = (Tyr, Asn or Phe); Xaa at res.26 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.28 = (Glu, 10 Lys, Asp, Gln or Ala); Xaa at res.30 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.31 = (Phe, Leu or Tyr); Xaa at res.33 = (Leu, Val or Met); Xaa at res.34 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.35 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.36 = (Tyr, Cys, His, Ser or Ile); Xaa at res.37 = (Met, Phe, Gly or Leu); Xaa at res.38 = (Asn, Ser or Lys); Xaa at res.39 = (Ala, Ser, Gly or Pro); Xaa at res.40 = (Thr, Leu or Ser); Xaa at res.44 = (Ile, Val or Thr); Xaa at res.45 = (Val, Leu or Ile); Xaa at res.46 = (Gln or Arg); Xaa at res.47 = (Thr, Ala or Ser); Xaa at res.48 = (Leu or Ile); Xaa at 20 res.49 = (Val or Met); Xaa at res.50 = (His, Asn or Arg); Xaa at res.51 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.52 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.53 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at res.54 = (Pro, Ser or Val); Xaa at res.55 = (Glu, Asp, 25 Asn, Gly, Val or Lys); Xaa at res.56 = (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.57 = (Val, Ala or Ile); Xaa at res.58 = (Pro or Asp); Xaa at res.59 = (Lys, Leu or Glu); Xaa at res.60 = (Pro or 30 Ala); Xaa at res.63 = (Ala or Val); Xaa at res.65 = (Thr, Ala or Glu); Xaa at res.66 = (Gln, Lys, Arg or Glu); Xaa at res.67 = (Leu, Met or Val); Xaa at res.68 = (Asn, Ser, Asp or Gly); Xaa at res.69 = (Ala, Pro or Ser); Xaa at res.70 = (Ile, Thr, Val or Leu); Xaa at res.71 = (Ser, Ala or Pro); Xaa at res.72 = (Val, Met 35

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or Ile); Xaa at res.74 = (Tyr or Phe); Xaa at res.75 =
(Phe, Tyr, Leu or His); Xaa at res.76 = (Asp, Asn or
Leu); Xaa at res.77 = (Asp, Glu, Asn or Ser); Xaa at
res.78 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.79 =

5 (Ser, Asn, Asp, Glu or Lys); Xaa at res.80 = (Asn, Thr
or Lys); Xaa at res.82 = (Ile, Val or Asn); Xaa at
res.84 = (Lys or Arg); Xaa at res.85 = (Lys, Asn, Gln,
His or Val); Xaa at res.86 = (Tyr or His); Xaa at
res.87 = (Arg, Gln, Glu or Pro); Xaa at res.88 = (Asn,

10 Glu or Asp); Xaa at res.90 = (Val, Thr, Ala or Ile);
Xaa at res.92 = (Arg, Lys, Val, Asp or Glu); Xaa at
res.93 = (Ala, Gly, Glu or Ser); Xaa at res.95 = (Gly
or Ala) and Xaa at res.97 = (His or Arg).

### Generic Sequence 6

Cys Xaa Xaa Xaa Leu Xaa Xaa Phe 10 5 1 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa 15 Xaa Xaa Pro Xaa Xaa Xaa Xaa Ala 25 20 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 30 Xaa Pro Xaa Xaa Xaa Xaa Xaa 40 Xaa Xaa Xaa Asn His Ala Xaa Xaa 50 45 Xaa Cys

Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa 70

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Xaa Xaa Xaa Leu Xaa Xaa Xaa 75 80
Xaa Xaa Xaa Xaa Val Xaa Leu Xaa 85

5 Xaa Xaa Xaa Xaa Met Xaa Val Xaa 90 95

Xaa Cys Xaa Cys Xaa 100

10 wherein each Xaa is independently selected from a group of one or more specified amino acids as defined by the following: "Res." means "residue" and Xaa at res.2 = (Lys, Arg, Ala or Gln); Xaa at res.3 = (Lys, Arg or Met); Xaa at res.4 = (His, Arg or Gln); Xaa at res.5 = 15 (Glu, Ser, His, Gly, Arg, Pro, Thr, or Tyr); Xaa at res.7 = (Tyr or Lys); Xaa at res.8 = (Val or Ile); Xaa at res.9 = (Ser, Asp or Glu); Xaa at res.11 = (Arg, Gln, Ser, Lys or Ala); Xaa at res.12 = (Asp, Glu, or Lys); Xaa at res.13 = (Leu, Val or Ile); Xaa at res.16 20 = (Gln, Leu, Asp, His, Asn or Ser); Xaa at res.17 = (Asp, Arg, Asn or Glu); Xaa at res.19 = (Ile or Val); Xaa at res.20 = (Ile or Val); Xaa at res.21 = (Ala or Ser); Xaa at res.23 = (Glu, Gln, Leu, Lys, Pro or Arg); Xaa at res.24 = (Gly or Ser); Xaa at res.25 = (Tyr or 25 Phe); Xaa at res.26 = (Ala, Ser, Asp, Met, His, Gln, Leu, or Gly); Xaa at res.28 = (Tyr, Asn or Phe); Xaa at res.31 = (Glu, His, Tyr, Asp, Gln or Ser); Xaa at res.33 = Glu, Lys, Asp, Gln or Ala); Xaa at res.35 = (Ala, Ser, Pro, Gln or Asn); Xaa at res.36 = (Phe, Leu 30 or Tyr); Xaa at res.38 = (Leu, Val or Met); Xaa at res.39 = (Asn, Asp, Ala, Thr or Pro); Xaa at res.40 = (Ser, Asp, Glu, Leu, Ala or Lys); Xaa at res.41 = (Tyr, Cys, His, Ser or Ile); Xaa at res.42 = (Met, Phe, Gly or Leu); Xaa at res.43 = (Asn, Ser or Lys); Xaa at 35 res.44 = (Ala, Ser, Gly or Pro); Xaa at res.45 = (Thr,

Leu or Ser); Xaa at res.49 = (Ile, Val or Thr); Xaa at res.50 = (Val, Leu or Ile); Xaa at res.51 = (Gln or Arg); Xaa at res.52 = (Thr, Ala or Ser); Xaa at res.53 = (Leu or Ile); Xaa at res.54 = (Val or Met); Xaa at 5 res.55 = (His, Asn or Arg); Xaa at res.56 = (Phe, Leu, Asn, Ser, Ala or Val); Xaa at res.57 = (Ile, Met, Asn, Ala, Val or Leu); Xaa at res.58 = (Asn, Lys, Ala, Glu, Gly or Phe); Xaa at res.59 = (Pro, Ser or Val); Xaa at res.60 = (Glu, Asp, Gly, Val or Lys); Xaa at res.61 = 10 (Thr, Ala, Val, Lys, Asp, Tyr, Ser, Ala, Pro or His); Xaa at res.62 = (Val, Ala or Ile); Xaa at res.63 = (Pro or Asp); Xaa at res.64 = (Lys, Leu or Glu); Xaa at res.65 = (Pro or Ala); Xaa at res.68 = (Ala or Val); Xaa at res.70 = (Thr, Ala or Glu); Xaa at res.71 = 15 (Gln, Lys, Arg or Glu); Xaa at res.72 = (Leu, Met or Val); Xaa at res.73 = (Asn, Ser, Asp or Gly); Xaa at res.74 = (Ala, Pro or Ser); Xaa at res.75 = (Ile, Thr, Val or Leu); Xaa at res.76 = (Ser, Ala or Pro); Xaa at res.77 = (Val, Met or Ile); Xaa at res.79 = (Tyr or 20 Phe); Xaa at res.80 = (Phe, Tyr, Leu or His); Xaa at res.81 = (Asp, Asn or Leu); Xaa at res.82 = (Asp, Glu, Asn or Ser); Xaa at res.83 = (Ser, Gln, Asn, Tyr or Asp); Xaa at res.84 = (Ser, Asn, Asp, Glu or Lys); Xaa at res.85 = (Asn, Thr or Lys); Xaa at res.87 = (Ile,25 Val or Asn); Xaa at res.89 = (Lys or Arg); Xaa at res.90 = (Lys, Asn, Gln, His or Val); Xaa at res.91 = (Tyr or His); Xaa at res.92 = (Arg, Gln, Glu or Pro); Xaa at res.93 = (Asn, Glu or Asp); Xaa at res.95 = (Val, Thr, Ala or Ile); Xaa at res.97 = (Arg, Lys, Val, Asp or Glu); Xaa at res.98 = (Ala, Gly, Glu or Ser); Xaa at res.100 = (Gly or Ala); and Xaa at res.102 = (His or Arg).

Particularly useful sequences for use as morphogens in this invention include the C-terminal domains, e.g., the C-terminal 96-102 amino acid residues of Vgl, Vgr-1, DPP, OP-1, OP-2, CBMP-2A, CBMP-2B, GDF-1 (see 5 Table II, below, and Seq. ID Nos. 5-14), as well as proteins comprising the C-terminal domains of 60A, BMP3, BMP5 and BMP6 (see Seq. ID Nos. 24-28), all of which include at least the conserved six or seven cysteine skeleton. In addition, biosynthetic 10 constructs designed from the generic sequences, such as COP-1, 3-5, 7, 16, disclosed in U.S. Pat. No. 5,011,691, also are useful. Other sequences include the inhibins/activin proteins (see, for example, U.S. Pat. Nos. 4,968,590 and 5,011,691). Accordingly, other 15 useful sequences are those sharing at least 70% amino acid sequence homology or "similarity", and preferably 80% homology or similarity with any of the sequences These are anticipated to include allelic and species variants and mutants, and biosynthetic muteins, 20 as well as novel members of this morphogenic family of proteins. Particularly envisioned in the family of related proteins are those proteins exhibiting morphogenic activity and wherein the amino acid changes from the preferred sequences include conservative 25 changes, e.g., those as defined by Dayoff et al., Atlas of Protein Sequence and Structure; vol. 5, Suppl. 3, pp. 345-362, (M.O. Dayoff, ed., Nat'l BioMed. Research Fdn., Washington, D.C. 1979). As used herein, potentially useful sequences are aligned with a known 30 morphogen sequence using the method of Needleman et al. ((1970) J.Mol.Biol. 48:443-453) and identities calculated by the Align program (DNAstar, Inc.). "Homology" or "similarity" as used herein includes allowed conservative changes as defined by Dayoff et 35 al.

The currently most preferred protein sequences useful as morphogens in this invention include those having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the Drosophila 60A protein. Accordingly, in another preferred aspect of the invention, useful morphogens include active proteins comprising species of polypeptide chains having the generic amino acid sequence herein referred to as "OPX", which accommodates the homologies between the various identified species of OP1 and OP2 (Seq. ID No. 29).

The morphogens useful in the methods, composition and devices of this invention include proteins comprising any of the polypeptide chains described above, whether isolated from naturally-occurring sources, or produced by recombinant DNA or other synthetic techniques, and includes allelic and species variants of these proteins, naturally-occurring or biosynthetic mutants thereof, as well as various 25 truncated and fusion constructs. Deletion or addition mutants also are envisioned to be active, including those which may alter the conserved C-terminal cysteine skeleton, provided that the alteration does not functionally disrupt the relationship of these 30 cysteines in the folded structure. Accordingly, such active forms are considered the equivalent of the specifically described constructs disclosed herein.

The proteins may include forms having varying glycosylation patterns, varying N-termini, a family of related proteins having regions of amino acid sequence homology, and active truncated or mutated forms of native or biosynthetic proteins, produced by expression of recombinant DNA in host cells.

The morphogenic proteins can be expressed from intact or truncated cDNA or from synthetic DNAs in procaryotic or eucaryotic host cells, and purified, cleaved, refolded, and dimerized to form morphogenically active compositions. Currently preferred host cells include <a href="Ecolic or mammalian cells">E. colic or mammalian cells</a>, such as CHO, COS or BSC cells. A detailed description of the morphogens useful in the methods, compositions and devices of this invention is disclosed in copending US patent application Serial Nos. 752,764, filed August 30, 1991, and 667,274, filed March 11, 1991, the disclosure of which are incorporated herein by reference.

Thus, in view of this disclosure, skilled genetic engineers can isolate genes from cDNA or genomic libraries of various different species which encode appropriate amino acid sequences, or construct DNAs from oligonucleotides, and then can express them in various types of host cells, including both procaryotes and eucaryotes, to produce large quantities of active proteins capable of protecting tissues and organs from immune cell-mediated tissue destruction, including substantially inhibiting such damage and/or regenerating the damaged tissue in a variety of mammals, including humans.

The foregoing and other objects, features and advantages of the present invention will be made more apparent from the following detailed description of the invention.

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### Brief Description of the Drawings

- shows the cardioprotective effects of morphogen (hOP1) in a rat myocardial ischemia-reperfusion model, as evidenced by the smaller loss of myocardial creatine kinase in hOP1-treated rats;
- FIG 2 shows the effects of 20 μg of morphogen (hOP1 given 24 hours prior to isolation of rat heart on endothelial-dependent vasorelaxation to acetycholine following induced ischemiareperfusion injury;
- 15 FIG 3 shows the effect of morphogen (hOP1) on neutrophil adherence to LTB<sub>4</sub>-stimulated mesenteric artery endothelium in neutrophilactivated rats;
- 20 FIG 4 (A and B) are schematic representations of morphogen inhibition of early mononuclear phagocytic multinuclearization in vivo;
- FIG 5 graphs the effect of a morphogen (e.g., OP-1)
  and a placebo control on mucositic lesion
  formation; and
- FIG 6 (A-D) graphs the effects of a morphogen (eg., OP-1, Figs. 6A and 6C) and TGF-β (Fig. 6B and 6D) on collagen (6A and 6B) and hyaluronic acid (6C and 6D) production in primary fibroblast cultures.

### Detailed Description of the Invention

It now has been surprisingly discovered that the morphogens defined herein are effective agents in alleviating the tissue destructive effects associated with the body's inflammatory response to tissue injury. In particular, as disclosed herein, the morphogens are capable of alleviating the necrotic tissue effects associated with the ensuing inflammatory responses that occur following an initial tissue injury.

When tissue injury occurs, whether caused by bacteria, trauma, chemicals, heat, or any other phenomenon, the body's inflammatory response is stimulated. In response to signals released from the damaged cells (e.g., cytokines), extravascularization of immune effector cells is induced. Under ordinary circumstances these invading immune effector cells kill the infectious agent and/or infected or damaged cells 20 (through the release of killing substances such as superoxides, perforins, and other antimicrobial agents stored in granules), remove the dead tissues and organisms (through phagocytosis), release various biological response modifiers that promote rapid 25 healing and covering of the wound (quite often resulting in the formation of fibrotic scar tissue), and then, after the area is successfully healed, exit from the site of the initial insult. Once the site is perceived to be normal, the local release of inflammatory cytokines ceases and the display of adhesion molecules on the vessel endothelium returns to basal levels. In some cases, however, the zeal of these interacting signals and cellular systems, which are designed to capture and contain very rapidly multiplying infectious agents, act to the detriment of

the body, killing additional, otherwise healthy, surrounding tissue. This additional unnecessary tissue death further compromises organ function and sometimes results in death of the individual. In addition, the resulting scar tissue that often forms can interfere with normal tissue function as occurs, for example, in idiopathic pulmonary fibrosis, IBD and organ cirrhosis.

The vascular endothelium constitutes the first 10 barrier between circulating immune effector cells and extravascular tissues. Extravasation of these circulating cells requires that they bind to the vascular endothelial cells, cross the basement membrane, and enter insulted tissues e.g. by 15 phagocytosis or protease-mediated extracellular matrix degradation. Without being limited to a particular theory, it is believed that the morphogens of this invention may modulate the inflammatory response in part by modulating the attachment of immune effector 20 cells to the luminal side of the endothelium of blood vessels at or near sites of tissue damage and/or inflammatory lesions. Because the method reduces or prevents the attachment of immune effector cells at these sites, it also prevents the subsequent release of 25 tissue destructive agents by these same immune effector cells at sites of tissue damage and/or inflammatory lesions. Because attachment of immune effector cells to the endothelium must precede their extravascularization, the method also prevents the 30 initial or continued entry of these cells into extravascular sites of tissue destruction or ongoing inflammatory lesions. Therefore, the invention not only relates to a method to reduce or prevent the immune cell-mediated cellular destruction at 35 extravascular sites of recent tissue destruction, but

also relates to a method to prevent or reduce the continued entry of immune effector cells into extravascular sites of ongoing inflammatory cascades. As will be appreciated by those skilled in the art, the morphogens of this invention also may be contemplated in mechanisms for disrupting the functional interaction of immune effector cells with endothelium where the adhesion molecules are induced by means other than in response to tissue injury.

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One source of tissue injury is induced by cell exposure to toxic oxygen concentrations, such as ischemic-reperfusion tissue injury (oxygen deprivation), and following hyperoxia injury (lethally high oxygen concentrations). Accordingly, the process of the present invention provides a method for alleviating the tissue damage induced by ischemic-reperfusion injury or hyperoxia-induced injury comprising the step of administering to the afflicted individual a therapeutic amount of a morphogen prior to, during, or after damage to the affected tissue. Where the toxic oxygen concentrations may be deliberately induced, as by a surgical or clinical procedure, the morphogen preferably is administered prior to induction.

In addition, the morphogens described herein, in contrast to fibrogenic growth factors such as TGF-β, stimulate tissue morphogenesis and do not stimulate fibrosis or scar tissue formation (see Example 9, below.) Accordingly, in addition to inhibiting the tissue destructive effects associated with the inflammatory response, the morphogens further enhance the viability of damaged tissue and/or organs by stimulating the regeneration of the damaged tissue and preventing fibrogenesis.

The morphogens described herein also can inhibit epithelial cell proliferation (see Example 10, below.) This activity of the morphogens also may be particularly useful in the treatment of psoriasis and other inflammatory diseases that involve epithelial cell populations.

Provided below are detailed descriptions of suitable morphogens useful in the methods and compositions of this invention, as well as methods for their administration and application, and numerous, nonlimiting examples which 1) illustrate the suitability of the morphogens and morphogen-stimulating agents described herein as therapeutic agents for protecting tissue from the tissue destructive effects associated with the body's inflammatory response; and 2) provide assays with which to test candidate morphogens and morphogen-stimulating agents for their efficacy.

20

#### I. Useful Morphogens

As defined herein a protein is morphogenic if it is

25 capable of inducing the developmental cascade of
cellular and molecular events that culminate in the
formation of new, organ-specific tissue and comprises
at least the conserved C-terminal six cysteine skeleton
or its functional equivalent (see supra).

30 Specifically, the morphogens generally are capable of
all of the following biological functions in a
morphogenically permissive environment: stimulating
proliferation of progenitor cells; stimulating the
differentiation of progenitor cells; stimulating the
proliferation of differentiated cells; and supporting

the growth and maintenance of differentiated cells, including the "redifferentiation" of transformed cells. Details of how the morphogens useful in the method of this invention first were identified, as well as a description on how to make, use and test them for morphogenic activity are disclosed in USSN 667,274, filed March 11, 1991 and USSN 752,764, filed August 30, 1991, the disclosures of which are hereinabove incorporated by reference. As disclosed therein, the morphogens may be purified from naturally-sourced material or recombinantly produced from procaryotic or eucaryotic host cells, using the genetic sequences disclosed therein. Alternatively, novel morphogenic sequences may be identified following the procedures disclosed therein.

Particularly useful proteins include those which comprise the naturally derived sequences disclosed in Table II. Other useful sequences include biosynthetic constructs such as those disclosed in U.S. Pat. 5,011,691, the disclosure of which is incorporated herein by reference (e.g., COP-1, COP-3, COP-4, COP-5, COP-7, and COP-16).

Accordingly, the morphogens useful in the methods and compositions of this invention also may be described by morphogenically active proteins having amino acid sequences sharing 70% or, preferably, 80% homology (similarity) with any of the sequences described above, where "homology" is as defined herein above.

The morphogens useful in the method of this invention also can be described by any of the 6 generic sequences described herein (Generic Sequences 1, 2, 3, 4, 5 and 6). Generic sequences 1 and 2 also may include, at their N-terminus, the sequence

Cys Xaa Xaa Xaa Xaa (Seq. ID No. 15)

Table II, set forth below, compares the amino acid 10 sequences of the active regions of native proteins that have been identified as morphogens, including human OP-1 (hOP-1, Seq. ID Nos. 5 and 16-17), mouse OP-1 (mOP-1, Seq. ID Nos. 6 and 18-19), human and mouse OP-2 15 (Seq. ID Nos. 7, 8, and 20-23), CBMP2A (Seq. ID No. 9), CBMP2B (Seq. ID No. 10), BMP3 (Seq. ID No. 26), DPP (from Drosophila, Seq. ID No. 11), Vgl, (from Xenopus, Seq. ID No. 12), Vgr-1 (from mouse, Seq. ID No. 13), GDF-1 (from mouse, Seq. ID Nos. 14, 32 and 33), 60A 20 protein (from Drosophila, Seq. ID Nos. 24 and 25), BMP5 (Seq. ID No. 27) and BMP6 (Seq. ID No. 28). sequences are aligned essentially following the method of Needleman et al. (1970) J. Mol. Biol., 48:443-453, calculated using the Align Program (DNAstar, Inc.) 25 the table, three dots indicates that the amino acid in that position is the same as the amino acid in hOP-1. Three dashes indicates that no amino acid is present in that position, and are included for purposes of illustrating homologies. For example, amino acid 30 residue 60 of CBMP-2A and CBMP-2B is "missing". Of course, both these amino acid sequences in this region comprise Asn-Ser (residues 58, 59), with CBMP-2A then comprising Lys and Ile, whereas CBMP-2B comprises Ser and Ile.

### TABLE II

	hOP-1	Cys	Lys	Lys	His	Glu	Leu	Tyr	Val			ક
	mOP-1	•••	•••	•••	* * * *	•••	•••	•••	• • •			
5	hOP-2	• • •	Arg	Arg	•••	•••	• • •	• • •	•••			<u>3</u>
	mOP-2	• • •	Arg	Arg	•••	•••	•,••	• • •	•••			
	DPP	•••	Arg	Arg	•••	Ser	•••	• • •	•••			
	Vgl	•••	• • •	Lys	Arg	His	•••	•••	• • •	1		
	Vgr-1	•••	• • •	• • •	•••	Gly	•••	•••	•••			
10	CBMP-2A	• • •	• • •	Arg	•••	Pro	• • •	• • •	•••			
	CBMP-2B	•••	Arg	Arg		Ser		•••	•••			
	BMP3	•••	Ala	Arg	Arg	Tyr	•••	Lys	• • •			-
	GDF-1	•••	Arg	Ala	Arg	Arg	•••	• • •	• • •			
	60A	•••	Gln	Het	Glu	Thr	• • •	• • •	•••			
15	BMP5	•••	• • •	• • •	• • •	•••	• • •	•••	• • •			
	вир6	•••	Arg	• • •	•••	• • •	•. • •	•••	• • •	1		
		1 -				5				1		
			,									
20	hOP-1	Ser	Phe	Arg	Asp	Leu	Gly	Trp	Gln	Asp	. •	
	mOP-1	•••	•••	• • •	• • •	• • •	•••	•••	•••	• • •		
	hOP-2	•••	• • •	Gln	•••	•••	•••	• •••	Leu	•••		
	mOP-2	Ser	• • •	• • •	• • •	• • •	•••	• • •	Leu	• • •		
	DPP	Asp		Ser	•••	Val	•••	• • •	Asp			
25	Vgl	Glu	•••	Lys	• • •	Val	•••	• • •	•••	Asn		
	Vgr-1	•••	•••	Gln	•••	Val	• • •		•••	• • •		
	CBMP-2A	Asp	•••	Ser	• • •	. Val	•••	• • •	Asn	•••		
	CBMP-2B	Asp	• • •	Ser		Val	• • •	• • •	Asn	•••		
	вир3	Asp	• • •	Ala	•••	Ile	•••	•••	Ser	Glu		
30	GDF-1	• • •	•••		Glu	Val	• • •	• • •	His	Arg		
	60A	Asp	• • •	Lys	• • •	•••	• • •	•••	His	•••		, s
	BMP5	•••		• • •	•••	•••	•••	•••	•••	• • •		
	BMP6	•••	•••	Gln	•••	•••	•••	•••		•••		نِ
	<del></del>		10					15				

	h0P-1	Trp	Ile	Ile	Ala	Pro	Glu	Gly	Tyr	Ala
•	mOP-1	•••	•••	•••	•••	• • •	• • •	•••	• • •	• • •
	hOP-2	•••	Val	• • •	•••	• • •	Gln	• • •	• • •	Ser
	mOP-2	• • •	Val	•••	• • •	•••	Gln	•••	•••	Ser
5	DPP	•••	• • •	Val	• • •	• • •	Leu	•••	•••	Asp
	<b>∳</b> gl	•••	Val	•••	•••	• • •	Gln	• • •	•••	Met
	Vgr-1	•••	• • •	•••	•••	• • •	Lys	• • •	• • •	• • •
	CBMP-2A	•••		Val	•••	• • •	Pro	• • •	• • •	His
	CBMP-2B	•••	•••	Val	• • •	• • •	Pro	• • •	•••	Gln
10	BMP3	•••	• • •	•••	Ser	• • •	Lys	Ser	Phe	Asp
	GDF-1	•••	Val	•••	• • •	• • •	Arg	• • •	Phe	Leu
	60A	•••		•••	•••	• • •		• • •	•••	Gly
	BMP5	•••	• • •		• • •	• • •	•••	• • •	• • •	• • •
	BMP6	•••	• • •	•••	•••		Lys	•••	• • •	• • •
15				20			_		25	
	100.1	47 -	<b></b>		C	Glu	Gly	Glu	Cys	Ala
	hOP-1	Ala	Tyr	Tyr	Cys		•		•	
	mOP-1	• • •	• • •	•••	• • •	• • •	• • •	•••	• • •	Ser
20	h0P-2	• • •	•••	•••	•••	• • •	• • •	•••	•••	
	mOP-2	•••	•••	•••	• • •	***	• • •		•••	Pro
	DPP	• • •		• • •	•••	His	• • •	Lys	• • •	
	Vgl	• • •	Asn	•••		Tyr	• • •	•••	•••	Pro
	Vgr-1	•••	Asn	•••	• • •	Asp	• • •	•••	•••	Ser
25	CBMP-2A	• • •	Phe	•••	• • •	His	• • •	Glu	• • •	Pro
	CBMP-2B	• • •	Phe	•••	• • •	His	• • •	Asp	•••	Pro
	BMP3	• • •	•••	•••	• • •	Ser	• • •	Ala	•••	Gln
	GDF-1	•••	Asn	• • •	• • •	Gln	•••	Gln	•••	• • •
	60A	•••	Phe	•••	•••	Ser	•••	•••	•••	Asn
30	BMP5	• • •	Phe	•••	•••	Asp	•••	•••	•••	Ser
	BMP6	•••	Asn	•••	•••	Asp	• • •	• • •	•••	Ser
					30					35
	hOP-1	Phe	Pro	Leu	Asn	Ser	Tyr	Met	Asn	Ala
35	mOP-1	1110	•••		•••	•••	•••		• • •	• • •
J	mor - 1	• • •	• • •		•••					

	hOP-2	•••	• • •	•••	Asp	•••	Cys	•••	• • •	•••
	mOP-2	•••	•••	• • •	Asp	• • •	Cys	•••	• • •	•••
	DPP	• • •	• • •	• • •	Ala	Asp	His	Phe	• • •	Ser
	Vgl	Tyr	•••	•••	Thr	Glu	Ile	Leu	• • •	Gly
5	Vgr-1	• • •		• • •	• • •	Ala	His	•••	•••	• • •
•	CBMP-2A		• • •		Ala	Asp	His	Leu	• • •	Ser
	CBMP-2B	•••	• • •	• • •	Ala	Asp	His	Leu	• • •	Ser
	GDF-1	Leu		Val	Ala	Leu	Ser	Gly	Ser**	• • •
	BMP3	•••		Met	Pro	Lys	Ser	Leu	Lys	Pro
10	60A	•••			•••	Ala	His	• • •	• • •	•••
10	BMP5	• • •	• • • •		•••	Ala	His	Met	•••	• • •
	BMP6	•••		• • •	•••	Ala	His	Met	•••	• • •
	Dill 0					40				
15	hOP-1	Thr	Asn	His	Ala	Ile	Val	Gln	Thr	Leu
1-3	mOP-1	• • •		• • •	•••	•••	•••	• • •	•••	• • •
	hOP-2	• • •		•••	•••	•••	Leu		Ser	• •
	mOP-2	• • •		• • •		•••	Leu	•••	Ser	• •
	DPP	• • •		• • •	•••	Val	• • •	•••	. • • •	• • •
20	Vgl	Ser		•••	•••	• • •	Leu	• • •	• • •	• • •
20	Vgr-1	• • •	•••	•••	•••	•••	• • •	•••	•••	• • •
	CBMP-2A	• • •	•••	• • •		•••	•••	•••	•••	• • •
	CBMP-2B	•••		•••	• • •	·	•••	• • •	• • •	• • •
	BMP3	Ser				Thr	Ile	• • •	Ser	Ile
25	GDF-1	Leu		•••		Val	Leu	Arg	Ala	• • •
23	60A		•••	•••		•••	• • •	• • •	•••	•••
	BMP5	•••	•••		•••	•••	• • •	•••	• • •	• • •
	BMP6	•••	•••	• • •		• • •	•••	• • •	•••	•••
	DILL 0	45			÷		50			
20										
30										
	hOP-1	Val	His	Phe	Ile	Asn	Pro	Glu	Thr	Val
	mOP-1	•••		•••	• • •	• • •	•••	Asp	• • •	•••
		•••	His	Leu	Het	Lys	•••	Asn	Ala	•••
ar	hOP-2	•••	His	Leu	Het	Lys		Asp	Val	•••
35	mOP-2	•••	1123			•				

	DPP	•••	Asn	Asn	Asn	• • •	• • •	Gly	Lys	• • •
	Vgl	• • •	• • •	Ser	• • •	Glu	•••	•••	Asp	Ile
	Vgr-1	• • •	•••	Val	Het	•••	•••	• • •	Tyr	• • •
	CBMP-2A	•••	Asn	Ser	Val	•••	Ser		Lys	Ile
5	CBMP-2B	•••	Asn	Ser	Val	• • •	Ser		Ser	Ile
	BMP3	• • •	Arg	Ala*	Gly	Val	Val	Pro	Gly	Ile
	GDF-1	Met	•••	Ala	Ala	Ala	• • •	Gly	Ala	Ala
	60A	• • •	• • •	Leu	Leu	Glu		Lys	Lys	• • •
	BMP5	• • •	• • •	Leu	Met	Phe	• • •	Asp	His	• • •
10	BMP6	• • •	• • •	Leu	Met		•••	• • •	Tyr	• • •
			55					60		
	hOP-1	Pro	Lys	Pro	Cys	Cys	Ala	Pro	Thr	Gln
15	mOP-1	•••	• • •	• • •	•••	• • •	• • •	•••	•••	•••
	hOP-2	•••	•••	Ala	• • •	• • •	•••	•••	• • •	Lys
	mOP-2	• • •	• • •	Ala	• • •	• • •	•••	• • •	•••	Lys
	DPP	• • •	• • •	Ala	• • •	• • •	Val	•••	• • •	•••
	Vgl	• • •	Leu	• • •	• • •	• • •	Val	• • •	•••	Lys
20	Vgr-1	• • •	. • • •	•••	• • •	• • •	• • •	•••	• • •	Lys
	CBMP-2A	• • •	• • •	Ala	• • •	• • •	Val	• • •	•••	Glu
	CBMP-2B	• • •	• • •	Ala	• • •	• • •	Val	•••	•••	Glu
	BMP3	• • •	Glu	•••	• • •	• • •	Val	•••	Glu	Lys
	GDF-1	Asp	Leu	•••	• • •	• • •	Val	• • •	Ala	Arg
25	A06	• • •	• • •	•••	• • •	• • •	• • •	• • •	• • •	Arg
	BMP5	• • •	• • •	• • •	• • •	• • •	• • •	• • •	• • •	Lys
	BMP6	•••	• • •	• • •	• • •	• • •	•••	• • •	• • •	Lys
				65					70	
30	h0P-1	Leu	Asn	Ala	Ile	Ser	Val	Leu	Tyr	Phe
	mOP-1	• • •	• • •	• • •	•••	• • •	• • •	•••	•••	• • •,
	hOP-2	• • •	Ser	•••	Thr	• • •	• • •	•••	•••	Tyr
	mOP-2	•••	Ser	•••	Thr	• • •	• • •	• • •	•••	Tyr
	Vgl	Met	Ser	Pro	• • •	• • •	Het	•••	Phe	Tyr
35	Vgr-1	Val	• • •	•••	•••	• • •	• • •	• • •	***	•••

										_	
	DPP	• • •	Asp	Ser	Val	Ala	Het	•••	•••	Leu	
	CBMP-2A	• • •	Ser	•••	•••	• • •	Het	• • •	•••	Leu	
	CBMP-2B	•••	Ser	• • •	• • •	• • •	Het	• • •	• • •	Leu	5
	вир3	Het	Ser	Ser	Leu	•••	Ile	• • •	Phe	Tyr	
5	GDF-1	•••	Ser	Pro	•••	• • •	•••	•••	Phe	• • •	3
	60A	•••	Gly	•••	Leu	Pro	•••	• • •	• • •	His	
	виР5	•••	• • •	•••	• • •	• • •	• • •	•••	•••	•••	
	вмР6	•••	•••	•••	•••	•••	• • •	•••	•••	Ť. • •	
					75					80	
10											
	hOP-1	Asp	Asp	Ser	Ser	Asn	Val	Ile	Leu	Lys	
	mOP-1	•••		•••	•••	•••	•••	• • •	• • •	•••	
	hOP-2		Ser	•••	Asn	• •, •	•••	• • •	•••	Arg	
	mOP-2	• • •	Ser	•••	Asn	• • •	•••	•••	•••	Arg	
15	DPP	Asn		Gln	•••	Thr	•••	Val	• • •	•••	
	Vgl		Asn	Asn	Asp	•••	• • •	Val		Arg	
	Vgr-1	•••		Asn	•••		• • •	•••	• • •	• • •	
	CBHP-2A	•••	Glu	Asn	Glu	Lys	• • •	Val	• • •	• • •	
	CBMP-2B	•••	Glu	Tyr	Asp	Lys	• • •	Val	• • •	• • •	
20	BMP3	• • • •	Glu	Asn	Lys	• • •	•••	Val	•••	• •	
20	GDF-1	•••	Asn		Asp	• • •	•••	Val	•••	Arg	
	60A	Leu	Asn	Asp	Glu	• • •	•••	Asn	•••	• • •	
	BHP5		• • •		• • •	• • •	•••	•••	• • •	•.• • •	
	BMP6	•••	• • •	Asn	• • •		• • •	•••	• • •	• • •	
25		٠				85	•				
	h0P-1	Lys	Tyr	Arg	Asn	Met	Val	Val	Arg		
	mOP-1	•••		• • •	•••	•••	• • •	• • •	• • •		
30	hOP-2	• • •	His		•••	•••	•••	• • •	Lys		
50	mOP-2	•••	His		•••	•••	• • •	• • •	Lys		5
	DPP	Asn	•••	Gln	Glu	•••	Thr	•••	Val		
	Vgl	His	• • •	Glu	•••	•••	Ala	•••	Asp		É
	Vgr-1	•••			•••	•••	•••	•••	•••		
35	CBMP-2A	Asn	• • •	Gln	Asp	•••		•••	Glu		
JJ	CDIII - ZII				-			•			

	CBMP-2B	Asn	• • •	Gln	Glu	• • •	• • •	• • •	Glu
	BMP3	Val	• • •	Pro	•••	•••	Thr	•••	Glu
	GDF-1	Gln	• • •	Glu	Asp	•••	• • •	•••	Asp
	60A	• • •	• • •	•••	• • •	•••	Ile	• • •	Lys
5	BMP5	• • •	•••	• • •	•••	• • •		• • •	• • •
	BMP6	• • •	•••	• • •	Trp	• • •		• • •	• • •
		90					95		
10	hOP-1	Ala	Cys	Gly	Cys	His			
	mOP-1	• • •		• • •	• • •	• • •			
	hOP-2	• • •	• • •	•••	• • •	• • •			
	mOP-2	•••	• • •	• • •	• • •	• • •			
	DPP	Gly	• • •	• • •	• • •	Arg			
15	Vgl	Glu	• • •	•••	• • •	Arg			
	Vgr-1	•••	• • •	• • •	• • •	• • •			
	CBMP-2A	Gly	• • •	• • •	• • •	Arg			
	CBMP-2B	Gly	•••	• • •	• • •	Arg			
	BMP3	Ser	•••	Ala	• • •	Arg			
20	GDF-1	Glu	• • •	•••	• • •	Arg			
	60A	Ser	• • •	•••	• • •	•••			
	BMP5	Ser	• • •	• • •	• • •	• • •			
	BMP6	• • •	• • •	•••	• • •	• • •			
				100					
25	**Between	residues	56 an	d 57	of BMP3	is a	Val	residue;	

25 \*\*Between residues 56 and 57 of BMP3 is a Val residue; between residues 43 and 44 of GDF-1 lies the amino acid sequence Gly-Gly-Pro-Pro.

30 As is apparent from the foregoing amino acid sequence comparisons, significant amino acid changes can be made within the generic sequences while retaining the morphogenic activity. For example, while the GDF-1 protein sequence depicted in Table II shares only about 50% amino acid identity with the hOP1

2

sequence described therein, the GDF-1 sequence shares greater than 70% amino acid sequence homology (or "similarity") with the hOP1 sequence, where "homology" or "similarity" includes allowed conservative amino acid changes within the sequence as defined by Dayoff, et al., Atlas of Protein Sequence and Structure vol.5, supp.3, pp.345-362, (M.O. Dayoff, ed., Nat'l BioMed. Res. Fd'n, Washington D.C. 1979.)

The currently most preferred protein sequences 10 useful as morphogens in this invention include those having greater than 60% identity, preferably greater than 65% identity, with the amino acid sequence defining the conserved six cysteine skeleton of hOP1 15 (e.g., residues 43-139 of Seq. ID No. 5). These most preferred sequences include both allelic and species variants of the OP-1 and OP-2 proteins, including the Drosophila 60A protein. Accordingly, in still another preferred aspect, the invention includes morphogens 20 comprising species of polypeptide chains having the generic amino acid sequence referred to herein as "OPX", which defines the seven cysteine skeleton and accommodates the identities between the various identified mouse and human OP1 and OP2 proteins. 25 is presented in Seq. ID No. 29. As described therein, each Xaa at a given position independently is selected from the residues occurring at the corresponding position in the C-terminal sequence of mouse or human OP1 or OP2 (see Seq. ID Nos. 5-8 and/or Seq. ID Nos. 16-23). 30

## II. Formulations and Methods for Administering Therapeutic Agents

The morphogens may be provided to an individual by 5 any suitable means, preferably directly (e.g., locally, as by injection or topical administration to a tissue locus) or systemically (e.g., parenterally or orally). Where the morphogen is to be provided parenterally, such as by intravenous, subcutaneous, intramuscular, 10 intraorbital, ophthalmic, intraventricular, intracranial, intracapsular, intraspinal, intracisternal, intraperitoneal, buccal, rectal, vaginal, intranasal or by aerosol administration, the morphogen preferably comprises part of an aqueous The solution is physiologically acceptable 15 solution. so that in addition to delivery of the desired morphogen to the patient, the solution does not otherwise adversely affect the patient's electrolyte and volume balance. The aqueous medium for the 20 morphogen thus may comprise normal physiologic saline (9.85% NaCl, 0.15M), pH 7-7.4. The aqueous solution containing the morphogen can be made, for example, by dissolving the protein in 50% ethanol containing acetonitrile in 0.1% trifluoroacetic acid (TFA) or 0.1% 25 HCl, or equivalent solvents. One volume of the resultant solution then is added, for example, to ten volumes of phosphate buffered saline (PBS), which further may include 0.1-0.2% human serum albumin (HSA). The resultant solution preferably is vortexed 30 extensively. If desired, a given morphogen may be made more soluble by association with a suitable molecule. For example, association of the mature dimer with the pro domain of the morphogen keeps the morphogen soluble in physiological buffers. In fact, the endogenous 35 protein is thought to be transported in this form.

Another molecule capable of enhancing solubility and particularly useful for oral administrations, is casein. For example, addition of 0.2% casein increases solubility of the mature active form of OP-1 by 80%.

5 Other components found in milk and/or various serum proteins also may be useful.

Useful solutions for parenteral administration may be prepared by any of the methods well known in the 10 pharmaceutical art, described, for example, in Remington's Pharmaceutical Sciences (Gennaro, A., ed.), Mack Pub., 1990. Formulations may include, for example, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, hydrogenated 15 naphthalenes, and the like. Formulations for direct administration, in particular, may include glycerol and other compositions of high viscosity to help maintain the morphogen at the desired locus. Biocompatible, preferably bioresorbable, polymers, including, for example, hyaluronic acid, collagen, tricalcium 20 phosphate, polybutyrate, lactide and glycolide polymers, and lactide/glycolide copolymers, may be useful excipients to control the release of the morphogen in vivo. Other potentially useful parenteral 25 delivery systems for these morphogens include ethylenevinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation administration contain as excipients, for example, lactose, or may be aqueous solutions containing, for example, polyoxyethylene-9lauryl ether, glycocholate and deoxycholate, or oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally.

Formulations for parenteral administration may also include glycocholate for buccal administration, methoxysalicylate for rectal administration, or cutric acid for vaginal administration.

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Suppositories for rectal administration also may be prepared by mixing the morphogen or morphogen-stimulating agent with a non-irritating excipient such as cocoa butter or other compositions which are solid at room temperature and liquid at body temperatures.

Formulations for topical administration to the skin surface may be prepared by dispersing the morphogen or morphogen-stimulating agent with a dermally acceptable carrier such as a lotion, cream, ointment or soap. Particularly useful are carriers capable of forming a film or layer over the skin to localize application and inhibit removal. For topical administration to internal tissue surfaces, the morphogen may be dispersed in a liquid tissue adhesive or other substance known to enhance adsorption to a tissue surface. For example, hydroxypropylcellulose or fibrinogen/thrombin solutions may be used to advantage. Alternatively, tissue-coating solutions, such as pectin-containing formulations, may be used.

Alternatively, the morphogens described herein may be administered orally. Oral administration of proteins as therapeutics generally is not practiced as most proteins are readily degraded by digestive enzymes and acids in the mammalian digestive system before they can be absorbed into the bloodstream. However, the morphogens described herein typically are acid stable and protease-resistant (see, for example, U.S. Pat.No. 4,968,590.) In addition, at least one morphogen, OP-1,

has been identified in mammary gland extract, colostrum and 57-day milk. Moreover, the OP-1 purified from mammary gland extract is morphogenically active. Specifically, this protein induces endochondral bone 5 formation in mammals when implanted subcutaneously in association with a suitable matrix material, using a standard in vivo bone assay, such as is disclosed in U.S. Pat.No. 4,968,590. Moreover, the morphogen also is detected in the bloodstream. Finally, soluble form 10 morphogen, e.g., mature morphogen associated with the pro domain, is morphogenically active. These findings indicate that oral and parenteral administration are viable means for administering morphogens to an individual. In addition, while the mature forms of 15 certain morphogens described herein typically are sparingly soluble, the morphogen form found in milk (and mammary gland extract and colostrum) is readily soluble, probably by association of the mature, morphogenically active form with part or all of the pro 20 domain of the intact sequence and/or by association with one or more milk components. Accordingly, the compounds provided herein also may be associated with molecules capable of enhancing their solubility in vitro or in vivo.

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Where the morphogen or morphogen-stimulating agent comprises part of a tissue or organ preservation solution, any commercially available preservation solution may be used to advantage. For example, useful solutions known in the art include Collins solution, Wisconsin solution, Belzer solution, Eurocollins solution and lactated Ringer's solution. Generally, an organ preservation solution usually possesses one or more of the following properties: (a) an osmotic pressure substantially equal to that of the inside of a

mammalian cell, (solutions typically are hyperosmolar and have K+ and/or Mg++ ions present in an amount sufficient to produce an osmotic pressure slightly higher than the inside of a mammalian cell; (b) the 5 solution typically is capable of maintaining substantially normal ATP levels in the cells; and (c) the solution usually allows optimum maintenance of qlucose metabolism in the cells. Organ preservation solutions also may contain anticoagulants, energy 10 sources such as glucose, fructose and other sugars, metabolites, heavy metal chelators, glycerol and other materials of high viscosity to enhance survival at low temperatures, free oxygen radical inhibiting agents and A detailed description of a pH indicator. 15 preservation solutions and useful components may be found, for example, in US Patent No. 5,002,965, the disclosure of which is incorporated herein by reference.

The compounds provided herein also may be associated with molecules capable of targeting the morphogen or morphogen-stimulating agent to the desired tissue. For example, an antibody, antibody fragment, or other binding protein that interacts specifically with a surface molecule on cells of the desired tissue, may be used. Useful targeting molecules may be designed, for example, using the single chain binding site technology disclosed, for example, in U.S. Pat. No. 5,091,513.

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As described above, the morphogens provided herein share significant sequence homology in the C-terminal active domains. By contrast, the sequences typically diverge significantly in the sequences which define the pro domain. Accordingly, the pro domain is thought to

be morphogen-specific. As described above, it is also known that the various morphogens identified to date are differentially expressed in the different tissues. Accordingly, without being limited to any given theory, 5 it is likely that, under natural conditions in the body, selected morphogens typically act on a given tissue. Accordingly, part or all of the pro domains which have been identified associated with the active form of the morphogen in solution, may serve as 10 targeting molecules for the morphogens described herein. For example, the pro domains may interact specifically with one or more molecules at the target tissue to direct the morphogen associated with the pro domain to that tissue. Accordingly, another useful 15 targeting molecule for targeting morphogen to a tissue of interest is part or all of a morphogen pro domain. For example, part or all of the pro domain of GDF-1 may be used to target a morphogen to nerve tissue. Alternatively, part or all of the pro domain of OP-1 or 20 CBMP2 may be used to target a morphogen to bone tissue, both of which proteins are found naturally associated with bone tissue.

The morphogens described herein are useful for

25 providing neuroprotective effects to alleviate neural
pathway damage associated with the body's
immune/inflammatory response to an initial injury to
nerve tissue. As used herein, a "neural pathway"
describes a nerve circuit for the passage of electric
30 signals from a source to a target cell site and
includes both the central nervous system (CNS) and
peripheral nervous system (PNS). The pathway includes
the neurons through which the electric impulse is
transported, including groups of interconnecting
35 neurons, the nerve fibers formed by bundled neuronal

axons, and the glial cells surrounding and associated with the neurons. An inflammatory response to nerve tissue injury may follow trauma to nerve tissue, caused, for example, by an autoimmune (including autoantibody) dysfunction, neoplastic lesion, infection, chemical or mechanical trauma, or other disease. An exemplary nerve-related inflammatory disease is multiple sclerosis. Neural pathway damage also can result from a reduction or interruption, e.g., 10 occlusion, of a neural blood supply, as in an embolic stroke, (e.g, ischemia or hypoxia-induced injury), or by other trauma to the nerve or surrounding material. In addition, at least part of the damage associated with a number of primary brain tumors also appears to 15 be immunologically related. Application of the morphogen directly to the cells to be treated, or providing the morphogen to the mammal systemically, for example, intravenously or indirectly by oral administration, may be used to alleviate and/or inhibit 20 the immunologically related response to a neural injury. Alternatively, administration of an agent capable of stimulating morphogen expression and/or secretion in vivo, preferably at the site of injury, also may be used. Where the injury is to be induced, 25 as during surgery or other aggressive clinical treatment, the morphogen or agent may be provided prior to induction of the injury to provide a neuroprotective effect to the nerve tissue at risk.

Where the morphogen is intended for use as a therapeutic to alleviate tissue damage associated with an immune/inflammatory condition of the CNS, an additional problem must be addressed: overcoming the so-called "blood-brain barrier", the brain capillary wall structure that effectively screens out all but

selected categories of molecules present in the blood, preventing their passage into the brain. The blood-brain barrier may be bypassed effectively by direct infusion of the morphogen or morphogen-5 stimulating agent into the brain. Alternatively, the morphogen or morphogen-stimulating agent may be modified to enhance its transport across the blood-brain barrier. For example, truncated forms of the morphogen or a morphogen-stimulating agent may be 10 most successful. Alternatively, the morphogen or morphogen-stimulating agent may be modified to render it more lipophilic, or it may be conjugated to another molecule which is naturally transported across the barrier, using standard means known to those skilled in 15 the art, as, for example, described in Pardridge, Endocrine Reviews 7:314-330 (1986) and U.S. Pat. No. 4,801,575.

agents provided herein may be administered alone or in combination with other molecules known to be beneficial in the treatment compositions and methods described herein, including, but not limited to anticoagulants, free oxygen radical inhibiting agents, salicylic acid, vitamin D, and other antiinflammatory agents. Psoriais treatments also may include ultra-violet light treatment, zinc oxide and retinoids.

The compounds provided herein can be formulated

into pharmaceutical compositions by admixture with

pharmaceutically acceptable nontoxic excipients and

carriers. As noted above, such compositions may be

prepared for parenteral administration, particularly in

the form of liquid solutions or suspensions; for oral administration, particularly in the form of tablets or capsules; or intranasally, particularly in the form of powders, nasal drops, or aerosols.

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The compositions can be formulated for parenteral or oral administration to humans or other mammals in therapeutically effective amounts, e.g., amounts which provide appropriate concentrations for a time sufficient to alleivate the tissue destructive effects associated with the inflammatory response, including protecting tissue in anticipation of tissue damage.

As will be appreciated by those skilled in the art, 15 the concentration of the compounds described in a therapeutic composition will vary depending upon a number of factors, including the dosage of the drug to be administered, the chemical characteristics (e.g., hydrophobicity) of the compounds employed, and the 20 route of administration. The preferred dosage of drug to be administered also is likely to depend on such variables as the type and extent of progression of the tissue damage, the overall health status of the particular patient, the relative biological efficacy of 25 the compound selected, the formulation of the compound excipients, and its route of administration. In general terms, the compounds of this invention may be provided in an aqueous physiological buffer solution containing about 0.001% to 10% w/v compound for parenteral 30 administration. Typical dose ranges are from about 10 ng/kg to about 1 g/kg of body weight per day; a preferred dose range is from about 0.1  $\mu$ g/kg to 100 mg/kg of body weight per day. Optimally, the morphogen dosage given is between 0.1-100  $\mu g$  of protein 35 per kilogram weight of the patient. No obvious

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morphogen induced pathological lesions are induced when mature morphogen (e.g., OP-1, 20 μg) is administered daily to normal growing rats for 21 consecutive days.
 Moreover, 10 μg systemic injections of morphogen (e.g., OP-1) injected daily for 10 days into normal newborn mice does not produce any gross abnormalities.

In administering morphogens systemically in the methods of the present invention, preferably a large volume loading dose is used at the start of the treatment. The treatment then is continued with a maintenance dose. Further administration then can be determined by monitoring at intervals the levels of the morphogen in the blood.

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Where tissue injury is induced deliberately as part of, for example, a surgical procedure, the morphogen preferably is provided just prior to, or concomitant with induction of the trauma. Preferably, the morphogen is administered prophylactically in a surgical setting.

Alternatively, an effective amount of an agent capable of stimulating endogenous morphogen levels may be administered by any of the routes described above. For example, an agent capable of stimulating morphogen production and/or secretion from cells of affected tissue and/or transplant tissue may be provided to a mammal, e.g., by direct administration of the agent to the tissue to be treated. A method for identifying and testing agents capable of modulating the levels of endogenous morphogens in a given tissue is described generally herein in Example 15, and in detail in copending USSN 752,859, filed August 30, 1991, the disclosure of which is incorporated herein by reference. Briefly, candidate compounds can be

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identified and tested by incubating the compound <u>in</u>

<u>vitro</u> with a test tissue or cells thereof, for a time
sufficient to allow the compound to affect the
production, i.e., the expression and/or secretion, of a
morphogen produced by the cells of that tissue.

For purposes of the present invention, the abovedescribed morphogens effective in alleviating tissue damage associated with ischemic-reperfusion injury (or 10 the agents that stimulate them, referred to collectively herein as "therapeutic agent") are administered prior to or during the restoration of oxygen (e.g., restoration of blood flow, reperfusion.) Where treatment is to follow an existing injury, the 15 therapeutic agent preferably is administered as an intravenous infusion provided acutely after the hypoxic or ischemic condition occurs. For example, the therapeutic agent can be administered by intravenous infusion immediately after a cerebral infarction, a 20 myocardial infarction, asphyxia, or a cardiopulmonary arrest. Where ischemia or hypoxia injury is deliberately and/or unavoidably induced as part of, for example, a surgical procedure where circulation to an organ or organ system is deliberately and/or 25 transiently interrupted, e.g., in carotid enterectomy, coronary artery bypass, grafting, organ transplanting, fibrinolytic therapy, etc., the therapeutic agent preferably is provided just prior to, or concomitant with, reduction of oxygen to the tissue. Preferably, 30 the morphogen is administered prophylactically in a surgical setting.

Similarly, where hyperoxia-induced injury already has occurred, the morphogen is administered upon diagnosis. Where hyperoxia injury may be induced as,

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for example, during treatment of prematurely newborn babies, or patients suffering from pulmonary diseases such as emphysema, the therapeutic agent preferably is administered prior to administration of oxygen e.g., prophylactically.

### III. Examples

# 10 Example 1. <u>Identification of Morphogen-Expressing</u> <u>Tissue</u>

Determining the tissue distribution of morphogens may be used to identify different morphogens expressed 15 in a given tissue, as well as to identify new, related Tissue distribution also may be used to identify useful morphogen-producing tissue for use in screening and identifying candidate morphogenstimulating agents. The morphogens (or their mRNA 20 transcripts) readily are identified in different tissues using standard methodologies and minor modifications thereof in tissues where expression may be low. For example, protein distribution may be determined using standard Western blot analysis or immunofluorescent techniques, and antibodies specific to the morphogen or morphogens of interest. Similarly, the distribution of morphogen transcripts may be determined using standard Northern hybridization protocols and transcript-specific probes.

Any probe capable of hybridizing specifically to a transcript, and distinguishing the transcript of interest from other, related transcripts may be used. Because the morphogens described herein share such high sequence homology in their active, C-terminal domains,

the tissue distribution of a specific morphogen transcript may best be determined using a probe specific for the pro region of the immature protein and/or the N-terminal region of the mature protein. 5 Another useful sequence is the 3' non-coding region flanking and immediately following the stop codon. These portions of the sequence vary substantially among the morphogens of this invention, and accordingly, are specific for each protein. For example, a particularly 10 useful Vgr-1-specific probe sequence is the PvuII-SacI fragment, a 265 bp fragment encoding both a portion of the untranslated pro region and the N-terminus of the mature sequence (see Lyons et al. (1989) PNAS 86:4554-4558 for a description of the cDNA sequence). Similarly, particularly useful mOP-1-specific probe sequences are the BstX1-BglI fragment, a 0.68 Kb sequence that covers approximately two-thirds of the mOP-1 pro region; a StuI-StuI fragment, a 0.2 Kb sequence immediately upstream of the 7-cysteine domain; and the Earl-Pstl fragment, an 0.3 Kb fragment 20 containing a portion of the 3'untranslated sequence (See Seq. ID No. 18, where the pro region is defined essentially by residues 30-291.) Similar approaches may be used, for example, with hOP-1 (Seq. ID No. 16) 25 or human or mouse OP-2 (Seq. ID Nos. 20 and 22.)

Using these morphogen-specific probes, which may be synthetically engineered or obtained from cloned sequences, morphogen transcripts can be identified in 30 mammalian tissue, using standard methodologies well known to those having ordinary skill in the art. Briefly, total RNA is prepared from various adult murine tissues (e.g., liver, kidney, testis, heart, brain, thymus and stomach) by a standard methodology such as by the method of Chomczyaski et al. ((1987)

Anal. Biochem 162:156-159) and described below. Poly (A)+ RNA is prepared by using oligo (dT)-cellulose chromatography (e.g., Type 7, from Pharmacia LKB Biotechnology, Inc.). Poly (A)+ RNA (generally 15  $\mu$ g) 5 from each tissue is fractionated on a 1% agarose/formaldehyde gel and transferred onto a Nytran membrane (Schleicher & Schuell). Following the transfer, the membrane is baked at 80°C and the RNA is cross-linked under UV light (generally 30 seconds at 1 10  $mW/cm^2$ ). Prior to hybridization, the appropriate probe is denatured by heating. The hybridization is carried out in a lucite cylinder rotating in a roller bottle apparatus at approximately 1 rev/min for approximately. 15 hours at 37°C using a hybridization mix of 40% 15 formamide, 5 x Denhardts, 5 x SSPE, and 0.1% SDS. Following hybridization, the non-specific counts are washed off the filters in 0.1 x SSPE, 0.1% SDS at 50°C.

Examples demonstrating the tissue distribution of 20 various morphogens, including Vgr-1, OP-1, BMP2, BMP3, BMP4, BMP5, GDF-1, and OP-2 in developing and adult tissue are disclosed in co-pending USSN 752,764, and in Ozkaynak, et al., (1991) Biochem. Biophys. Res. Commn. 179:116-123, and Ozkaynak, et al. (1992) (JBC, in 25 press), the disclosures of which are incorporated herein by reference. Using the general probing methodology described herein, northern blot hybridizations using probes specific for these morphogens to probe brain, spleen, lung, heart, liver 30 and kidney tissue indicate that kidney-related tissue appears to be the primary expression source for OP-1, with brain, heart and lung tissues being secondary sources. OP-1 mRNA also was identified in salivary glands, specifically rat parotid glands, using this probing methodology. Lung tissue appears to be the

primary tissue expression source for Vgr-1, BMP5, BMP4 and BMP3. Lower levels of Vgr-1 also are seen in kidney and heart tissue, while the liver appears to be a secondary expression source for BMP5, and the spleen appears to be a secondary expression source for BMP4. GDF-1 appears to be expressed primarily in brain tissue. To date, OP-2 appears to be expressed primarily in early embryonic tissue. Specifically, northern blots of murine embryos and 6-day post-natal animals shows abundant OP2 expression in 8-day embryos. Expression is reduced significantly in 17-day embryos and is not detected in post-natal animals.

### Example 2. Active Morphogens in Body Fluids

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OP-1 expression has been identified in saliva (specifically, the rat parotid gland, see Example 1); human blood serum, and various milk forms, including mammary gland extract, colostrum, and 57-day bovine 20 milk. Moreover, and as described in USSN 923,780, the disclosure of which is incorporated herein by reference, the body fluid-extracted protein is morphogenically active. The discovery that the morphogen naturally is present in milk and saliva, 25 together with the known observation that mature, active OP-1 is acid-stable and protease-resistant, indicate that oral administration is a useful route for therapeutic administration of morphogen to a mammal. Oral administration typically is the preferred mode of 30 delivery for extended or prophylactic therapies. addition, the identification of morphogen in all milk forms, including colostrum, suggests that the protein may play a significant role in tissue development, including skeletal development, of juveniles.

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### 2.1 Morphogen Detection in Milk

oP-1 was partially purified from rat mammary gland extract and bovine colostrum and 57 day milk by passing these fluids over a series of chromatography columns:

(e.g., cation-exchange, affinity and reverse phase). At each step the eluant was collected in fractions and these were tested for the presence of OP-1 by standard immunoblot. Immunoreactive fractions then were combined and purified further. The final, partially purified product then was examined for the presence of OP-1 by Western blot analysis using OP-1-specific antisera, and tested for in vivo and in vitro activity.

OP-1 purified from the different milk sources were characterized by Western blotting using antibodies raised against OP-1 and BMP2. Antibodies were prepared using standard immunology protocols well known in the art, and as described generally in Example 15, below, using full-length <a href="E.coli-produced OP-1">E.coli-produced OP-1</a> and BMP2 as the immunogens. In all cases, the purified OP-1 reacted only with the anti-OP-1 antibody, and not with anti-BMP2 antibody.

The morphogenic activity of OP-1 purified from mammary gland extract was evaluated in vivo essentially following the rat model assay described in U.S. Pat. No. 4,968,590, hereby incorporated by reference. Briefly, a sample was prepared from each OP-1 immunoreactive fraction of the mammary gland extract-derived OP-1 final product by lyophilizing a portion (33%) of the fraction and resuspending the protein in 220µl of 50% acetonitrile/0.1% TFA. After vortexing, 25 mg of collagen matrix was added. The samples were lyophilized overnight, and implanted in

Long Evans rats (Charles River Laboratories, Wilmington, MA, 28-35 days old). Each fraction was implanted in duplicate. For details of the collagen matrix implantation procedure, see, for example, U.S. Pat. No. 4,968,590, hereby incorporated by reference. After 12 days, the implants were removed and evaluated for new bone formation by histological observation as described in U.S. Patent No. 4,968,590. In all cases, the immunoreactive fractions were osteogenically active.

### 2.2 Morphogen Detection in Serum

Morphogen may be detected in serum using morphogen-15 specific antibodies. The assay may be performed using any standard immunoassay, such as Western blot (immunoblot) and the like. Preferably, the assay is performed using an affinity column to which the morphogen-specific antibody is bound and through which 20 the sample serum then is poured, to selectively extract the morphogen of interest. The morphogen then is A suitable elution buffer may be determined eluted. empirically by determining appropriate binding and elution conditions first with a control (e.g., 25 purified, recombinantly-produced morphogen.) Fractions then are tested for the presence of the morphogen by standard immunoblot, and the results confirmed by N-terminal sequencing. Preferably, the affinity column is prepared using monoclonal antibodies. Morphogen 30 concentrations in serum or other fluid samples then may be determined using standard protein quantification techniques, including by spectrophotometric absorbance or by quantitation of conjugated antibody.

Presented below is a sample protocol for identifying OP-1 in serum. Following this general methodology other morphogens may be detected in body fluids, including serum. The identification of 5 morphogen in serum further indicates that systemic administration is a suitable means for providing therapeutic concentrations of a morphogen to an individual, and that morphogens likely behave systemically as endocrine-like factors. Finally, using 10 this protocol, fluctuations in endogenous morphogen levels can be detected, and these altered levels may be used as an indicator of tissue dysfunction. Alternatively, fluctuations in morphogen levels may be assessed by monitoring morphogen transcription levels, 15 either by standard northern blot analysis as described in Example 1, or by in situ hybridization, using a labelled probe capable of hybridizing specifically to morphogen mRNA, and standard RNA hybridization protocols well described in the art and described 20 generally in Example 1.

OP-1 was detected in human serum using the following assay. A monoclonal antibody raised against mammalian, recombinantly produced OP-1 using standard immunology techniques well described in the art and described generally in Example 15, was immobilized by passing the antibody over an agarose-activated gel (e.g., Affi-Gel<sup>TM</sup>, from Bio-Rad Laboratories, Richmond, CA, prepared following manufacturer's instructions) and used to purify OP-1 from serum. Human serum then was passed over the column and eluted with 3M K-thiocyanate. K-thiocyanante fractions then were dialyzed in 6M urea, 20mM PO<sub>4</sub>, pH 7.0, applied to a C8 HPLC column, and eluted with a 20 minute, 25-50% acetonitrile/0.1% TFA gradient. Mature, recombinantly

produced OP-1 homodimers elute between 20-22 minutes. Fractions then were collected and tested for the presence of OP-1 by standard immunoblot using an OP-1 specific antibody as for Example 2.A.

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Administered or endogenous morphogen levels may be monitored in the therapies described herein by comparing the quantity of morphogen present in a body fluid sample with a predetermined reference value, for example, to evaluate the efficiency of a therapeutic protocol, and the like. In addition, fluctuations in the level of endogenous morphogen antibodies may be detected by this method, most likely in serum, using an antibody or other binding protein capable of interacting specifically with the endogenous morphogen antibody. Detected fluctuations in the levels of the morphogen or endogenous antibody may be used, for example, as indicators of a change in tissue status. For example, as damaged tissue is regenerated and the 20 tissue or organ's function returns to "normal" and, in the absence of additional tissue damage, lower doses of morphogen may be required, and a higher level of circulating morphogen antibody may be measured.

# 25 Example 3. Effect of Morphogen after the Onset of the Ischemic Process

The cardioprotective effect of morphogens following ischemic-reperfusion injury in a mammal can readily be assessed in a rat model. In this example, morphogen (e.g., OP-1) is administered just prior to the onset of the ischemic process in experimentally-induced myocardial infracted rats, essentially following the method of Lefer, et al. (1990) Science 249:61-64 and (1992) J. Mol. Cell. Cardiol. 24: 385-393, the

disclosures of which are hereby incorporated by reference. Briefly, loss of myocardial tissue function following ischemia and reperfusion is assayed by measuring loss of myocardial creatine kinease activity (CK) and loss of endothelium-dependent vasorelaxation function (see Example 4, below).

In a first group of ether-anesthetized rats, the
left coronary artery was occluded just proximal to the
first main branch with a silk ligature to induce a
myocardial infarction (MI). The ligature was removed
10 minutes after occlusion to allow for coronary
reperfusion. This first group is referred to herein as
the "myocardial infarcted" (MI) group. A second group
of rats underwent the same procedure except that the
coronary artery was not occluded, and thus no
myocardial infarction occurred. The second group of
rats is referred to herein as the "sham myocardial
infarcted group" (SHAM MI).

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The first group of rats, the MI group of rats, further was divided into three sup-groups. 2μg of morphogen (OP-1) were injected intravenously into the first sub-group of MI rats 10 minutes after ligature, immediately before reperfusion; into the second sub-group of MI rats 20 μg of OP-1 were injected intravenously 10 minutes after ligature and immediately before reperfusion; and into the third sub-group of MI rats (control) was injected vehicle only, e.g., 0.9%
30 NaCl, as for the OP-1 treated rats.

Twenty-four hours later, the hearts were removed from all of the rats and the levels of creatine kinase (CK) from the left ventricle (the infarcted region) and from the interventricular septum (the control

nonischemic region) were determined by standard means. By comparing the difference in CK activities in both regions, the amount of CK activity lost from the infarcted region was used as an index of cardiac cellular injury to the infarcted region.

As shown in Figure 1, the data indicate that morphogens (e.g., OP-1) can provide significant cardioprotective effect when provided to ischemic tissue. In the figure, CK loss is graphed as the difference in specific CK activity between the interventricular septum and the left ventricle.

The loss of CK activity by the subgroup of MI rats which received 2 µg of OP-1 just before reperfusion showed some protection as compared with the control MI rats which received injections of vehicle alone, when the levels from both subgroups are measured against, and compared to, the levels obtained for the SHAM MI control. Significant cardioprotection was observed in the subgroup of MI rats which received 20 µg of OP-1 immediately before reperfusion as compared with the control MI rats which received injections of vehicle alone, when the levels from both subgroups are measured against, and compared to, the levels contained within the SHAM MI control.

These data indicate that OP-1 offers significant cardiac protection when administered after ischemia and 30 before reperfusion.

A variation of this example also may be performed providing morphogen to the animal prior to induction of ischemia. The experiments may be performed both in normal and immune-compromised rats to assess the cardioprotective effects of morphogen administered prior to ischemia.

# Example 4. <u>Vasodilation of Myocardial Infarcted</u> Cardiac Tissue Treated with Morphogen

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Certain vasodilators like acetylcholine (ACh) and adenosine diphosphate (ADP, an immune mediator) exert their vasodilation activity only in the presence of intact endothelium, which is stimulated to release a substance termed endothelium-derived relaxing factor (EDRF). If the endothelium is injured so that EDRF is not released, no vasodilation occurs in response to these endothelium-dependent agents. In contrast, several other vasodilators including nitroglycerine (NTG) and nitroprusside, are endothelium-independent dilators, as they dilate blood vessels directly.

The present example demonstrates the ability of OP
1 to prevent the loss of cardioendothelium-dependent

25 relaxation (EDR) activity in the coronary
microvasculature following reperfusion of ischemic
myocardium, and their ability to reduce myocardial
injury 24 hours after morphogen treatment. Briefly, 2
or 24 hours after morphogen treatment ischemia
30 reperfusion injury is induced in isolated rat hearts,
the reperfused hearts are are vasodilated with either
ACh or NTG. In the absence of morphogen treatment,
injured tissue should inhibit ACh-induced vasodilation,
but not NTG-induced vasodilation. Morphogen treatment

35 in expected to enhance ACh-induced vasodilation in the
reperfused hearts.

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Accordingly, 48 adult male Sprague-Dawley rats (250-330 g) were divided into eight groups of 6 rats Twelve rats were subjected to sham myocardial infarcts (SHAM MI) as described in Example 3. The 5 hearts of the remaining 36 rats were isolated as follows: one set of twelve rats was injected intravenously with OP-1 24 hours prior to isolation of the heart; another set of rats was injected intravenously with  $20\mu g$  of OP-1 2 hours prior to 10 isolation of the heart; the final group of rats was injected with vehicle only (e.g., 0.9% NaCl.). The rats then were anesthetized with pentobarbital sodium (35 mg/kg, intraperitonial); their hearts were isolated and perfused by the Langendorff method at a constant 15 flow (15 ml/min) with oxygenated Krebs-Henseleit solution (Aoki et al. (1988) J. Pharmacol. 95:35).

Each group of rats then were divided into two subgroups of six rats each. Twenty minutes before reperfusion, coronary vasodilator response was measured by inducing constriction with 0.05 µmol U-44619 (9,11-methanoepoxyprostaglandin H<sub>2</sub>) followed by a vasodilating agent 3 minutes later: subgroup one - 15 nmol ACh; subgroup 2 - 15 nmol NTG and the increase in coronary perfusion pressure (CPP) level measured as an indication of vasodilation. When CPP levels returned to normal, the hearts were subjected to ischemia by reducing coronary infusion to 15% of control flow for 30 minutes, then reestablishing normal flow, i.e., reperfusion, for an additional 20 minutes.

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The vasodilator reponse then was remeasured by constriction and administration of vasodilating agent as described above.

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The results of these experiments are shown in FIG 2. Before the ischemic event, both Ach and NTG gave normal vasorelaxant results in all events. which received OP-1 24 hours prior to ischemia showed 5 an approximately 70% response to ACh while the hearts which received OP-1 2 hours prior to ischemia showed a 55% response to ACh. The group which received vehicle alone showed a 40% response to ACh. Finally, the control group which was not subjected to ischemia 10 showed an ACh response of approximately 95%. This shows that endothelium-dependent vasodilators exert a reduced vasodilator response following ischemia and reperfusion in the rat heart. Moreover, OP-1 significantly preserved endothelium-dependent dilation 15 when provided 24 hours prior to induction of myocardial ischemia. No defect in vasodilation occurred in response to the direct vasodilator (NTG); NTG-induced vasodilation activities were 95% of initial in hearts subject to ischemia and 100% of initial nonischemic 20 hearts.

### Example 5. Effect of Morphogen on Neutrophil Adherence

The role of neutrophil adherence in endothelium

25 dysfunction and the cardioprotective effects of
morphogens in modulating this activity can be assessed
using a standard polymorphonuclear neutrophil (PMN)
adherence assay such as described in Lefer et al.,
(1992) J. Mol. Cell. Cardiol. 24: 385-393, disclosed

30 hereinabove by reference. Briefly, segments of
superior mesenteric artery were isolated from rats
which had either been treated with morphogen (OP-1, 20

µg) or 0.9% NaCl, 24 h prior to isolation of the
artery. The segments were cleaned, cut into transverse

35 rings of 1-2mm in length, and these were subsequently

cut open and incubated in K-H solution at 37°C, pH 7.4.
Neutrophils were prepared and fluorescently labelled
using standard procedures (e.g., leukocytes were
isolated from rats essentially following the procedure
of Pertroft et. al. (1968) Exp Cell Res 50: 355-368,
washed in phosphate buffered saline (PBS), purified by
gradient centrifugation; and labelled by the method of
Yuan et. al. (1990) Microvasc Res 40: 218-229..

Labelled neutrophils then were added to open ring baths and activated with 100nM leukotriene  $B_4$  (LTB $_4$ ). Rings were incubated for 20 minutes and the number of neutrophils adhering to the endothelial surface then determined visually by fluorescent microscopy.

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As shown in Figure 3, unstimulated PMNs (i.e., PMNs alone) added to the baths did not significantly adhere to the vascular endothelium. In rings taken from rats injected with 0.9% NaCl, activation of neutrophils with LTB<sub>4</sub> (100 nM) greatly increased the number of PMNs adherent to the endothelium (P<0.001). OP-1 (20 µg administered 24 h prior) significantly inhibited adherence of PMNs activated by LTB<sub>4</sub> (P<0.01 from control).

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Example 6. In Vivo Models for Ischemic-Reperfusion
Protection in Lung, Nerve and Renal
Tissue.

Other tissues seriously affected by ischemicreperfusion injury include neural tissue, renal tissue
and lung tissue. The effect of morphogens on
alleviating the ischemic-reperfusion injury in these
tissues may be assessed using methodologies and models
known to those skilled in the art, and disclosed below.

Similarly, a methodology also is provided for assessing the tissue-protective effects of a morphogen on damaged lung tissue following hyperoxia injury.

For example, the rabbit embolic stroke model 5 provides a useful method for assessing the effect of morphogens on tissue injury following cerebral ischemia-reperfusion. The protocol disclosed below is essentially that of Phillips et al. (1989) Annals of 10 Neurology 25:281-285, the disclosure of which is herein incorporated by reference. Briefly, white New England rabbits (2-3kg) are anesthesized and placed on a respirator. The intracranial circulation then is selectively catheterized by the Seldinger technique. 15 Baseline cerebral angiography then is performed, employing a digital substration unit. The distal internal carotid artery or its branches then is selectively embolized with 0.035 ml of 18-hour-aged autologous thrombus. Arterial occlusion is documented 20 by repeat angiography immediately after embolization. After a time sufficient to induce cerebral infarcts (|15 minutes or 90 minutes), reperfusion is induced by administering a bolus of a reperfusion agent such as the TPA analogue Fb-FB-CF (e.g., 0.8 mg/kg over 2 25 minutes).

The effect of morphogen on cerebral infarcts can be assessed by administering varying concentrations of morphogens, e.g., OP1, at different times preceding or following embolization and/or reperfusion. The rabbits are sacrificed 3-14 days post embolization and their brains prepared for neuropathological examination by fixing by immersion in 10% neutral buffered formalin

for at least 2 weeks. The brains then are sectioned in a coronal plane at 2-3 mm intervals, numbered and submitted for standard histological processing in paraffin, and the degree of neutral tissue necrosis determined visually.

The renal-protective effects of morphogens on renal ischemia-reperfusion injury readily can be assessed using the mouse model disclosed by Oueliette, et al.

10 (1990), J. Clin. Invest. 85:766-771, the disclosure of which is hereby incorporated by reference. Briefly, renal ischemia is induced surgically in 35-45 days old out-bred Swiss male mice by performing a standard right nephrectomy, and occluding the artery to the left

15 kidney with a microaneurism clamp for 10-30 minutes. Morphogen then may be provided parentally, at various times prior to or following occulsion and/or reperfusion. The effects of morphogen then may be assessed by biological evaluation and histological evaluation using standard techniques well known in the art.

The tissue protective effects of morphogen on tissue exposed to lethally high oxygen concentrations may be assessed by the following procedure. Adult rats (275-300 gms) first are provided with morphogen (e.g., hOP1) or vehicle only, and then are exposed to 96-98% oxygen essentially as described by Rinaldo et al (1983) Am. Rev. Respir. Dis. 130:1065, to induce hyperoxia.

Animals are housed in plastic cages (38 cm x 48 xm x 21 cm). A cage containing 4-5 animals is placed in a 75 liter water-sealed plexiglass chamber. An atmosphere of 96-98% oxygen then is maintained by delivery of 02 gas (liquid O2). Gas flow through the chamber is adjusted to maintain at least 10 air changes/hr.,

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temperature at  $22 \pm 1^{\circ}$ C, minimal levels of condensation within the cage, and carbon dioxide concentration of < 0.5% as measured with a mass spetrophotometric medical gas analyzer.

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At the end of 72 hours all survivors are observed at room air for 1.5 hours and at longer time periods to assess degree of respiratory distress and cyanosis induced by the initial insult and subsequent immune cell-mediated damage. The number of survivors at the end of the challenge is recorded and the treated groups compared with the untreated control group by chi-square test of proportions. Several of the surviving animals for each group are randomly chosen for histological processing of lung tissue.

Lung tissue for histological processing is fixed by infusion of 10% buffered formalin through a tracheal cannula at a constant pressure of 20 cm H<sub>2</sub>O. After fixation for 24-48 hours, sections from each lobe are cut and subsequently stained with hematoxylin and eosin. Coded slides then are examined, preferably in a double-blind fashion for evidence of pathological changes such as edema, interstitial cellularity, and inflammatory response.

# Example 7. Morphogen Inhibition of Cellular and Humoral Inflammatory Response

Morphogens described herein inhibit multinucleation of mononuclear phagocytic cells under conditions where these cells normally would be activated, e.g., in response to a tissue injury or the presence of a foreign substance. For example, in the absence of morphogen, an implanted substrate material (e.g.,

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implanted subcutaneously) composed of, for example, mineralized bone, a ceramic such as titanium oxide or any other substrate that provokes multinucleated giant cell formation, rapidly becomes surrounded by 5 multinucleated giant cells, e.g., activated phagocytes stimulated to respond and destroy the foreign object. In the presence of morphogen however, the recruited cells remain in their mononuclear precursor form and the matrix material is undisturbed. Figure 4 10 illustrates this effect of morphogens, in a schematic representation of histology results of a titanium oxide substrate implanted subcutaneously. In the figure, "mg" means mononuclear giant cells and "ob" means osteoblasts. The substrate represented in Fig. 4B was implanted together with morphogen (OP-1) and newly 15 formed osteoblasts are evident surrounding the substrate. By contrast, the substrate represented in Fig. 4A was implanted without morphogen and extensive multinucleated giant cell formation is evident 20 surrounding the substrate. Accordingly, the morphogens' effect in inhibiting excessive bone mass loss in a mammal also may include inhibiting activation of these cells.

In addition, the morphogens described herein also suppress antibody production stimulated in response to a foreign antigen in a mammal. Specifically, when bovine bone collagen matrix alone was implanted in a bony site in a rat, a standard antibody response to the collagen is stimulated in the rat as determined by standard anti-bovine collagen ELISA experiments performed on blood samples taken at four week intervals following implantation (e.g., between 12 and 20 weeks.) Serum anti-collagen antibody titers, measured by ELISA essentially following the procedure described by

Nagler-Anderson et al, (1986) PNAS 83:7443-7446, the disclosure of which is incorporated herein by reference, increased consistently throughout the experiment. However, when the matrix was implanted together with a morphogen (e.g., OP-1, dispersed in the matrix and adsorbed thereto, essentially as described in U.S. Pat. No. 4,968,590) anti-bovine collagen antibody production was suppressed significantly. This ability of morphogen to suppress the humoral response is further evidence of morphogen utility in alleviating tissue damage associated with autoimmune diseases, including autoantibody diseases, such as rheumatoid arthritis.

15 Example 8. Morphogen protection of Gastrointestinal

Tract Mucosa from Ulceration and

Inflammation

inflammatory disease which involves ulcerations of the mouth mucosa as a consequence of, e.g., radiation therapy or chemotherapy. While not typically a chronic disease, the tissue destructive effects of oral mucositis mirror those of chronic inflammatory diseases such as IBD. The example below demonstrates morphogen efficacy in protecting the oral mucosa from oral mucositis in a hamster model, including both inhibiting inflammatory ulceration and enhancing regeneration of ulcerated tissue. Details of the protocol can be found in Sonis, et al., (1990) Oral Surg. Oral Med. Oral Pathol 69: 437-443, the disclosure of which is incorporated herein by reference. Based on these data,

the morphogens described herein should be efficacious in treating chronic inflammatory diseases including IBD, arthritis, psoriasis and psoriatic arthritis, multiple sclerosis, and the like.

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Golden syrian hamsters (6-8 wks old, Charles River Laboratories, Wilmington, MA) were divided into 3 test groups: Group 1, a placebo (e.g., saline) control, and a morphogen low dose group (100 ng) and a morphogen 10 high dose group (1 μg), Groups 2 and 3, respectively. Morphogen dosages were provided in 30% ethanol. Each group contained 12 animals.

Beginning on day 0 and continuing through day 5,

15 Groups 2 and 3 received twice daily morphogen
applications. On day 3, all groups began the
mucositis-induction procedure. 5-fluorouracil (60
mg/kg) was injected intraperitoneally on days 3 and 5.

On day 7, the right buccal pouch mucosa was

20 superficially irritated with a calibrated 18 gauge
needle. In untreated animals, severe ulcerative
mucositis was induced in at least 80% of the animals by
day 10.

25 For each administration of the vehicle control
(placebo) or morphogen, administration was performed by
first gently drying the cheek pouch mucosa, then
providing an even application over the mucosal surface
of the vehicle or morphogen material. A

30 hydroxypropylcellulose-based coating was used to
maintain contact of the morphogen with the mucosa.
This coating provided at least 4 hours of contact time.

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On day 12, two animals in each group were sacrificed for histological studies. The right buccal pouch mucosa and underlying connective tissue were dissected and fixed in 10% formalin using standard dissection and histology procedures. The specimens were mounted in paraffin and prepared for histologic examination. Sections then were stained with hematoxylin and eosin and were examined blindly by three oral pathologists with expertise in hamster histology and scored blind against a standard mucositis panel. The extent of atrophy, cellular infiltration, connective tissue breakdown, degree of ulceration and epithelialization were assessed.

15 The mean mucositis score for each group was determined daily for each experimental group for a period of 21 days by photography and visual examination of the right buccal cheek pouch. Differences between groups were determined using a standard 't' test, e.g., the Students' 't' test. In addition, data was evaluated between groups by comparing the numbers of animals with severe mucositis using Chi Square statistical analysis. The significance of differences in mean daily weights also was determined.

The experimental results are presented in Fig. 5, which graphs the effect of morphogen (high dose, squares; low dose, diamonds) and placebo (circles) on mean mucositis scores. Both low and high morphogen doses inhibit lesion formation significantly in a dose-dependent manner. In addition, histology results

consistently showed significantly reduced amounts of

tissue atrophy, cellular debris, and immune effector cells, including macrophages and activated neutrophils, in the morphogen-treated animals, as compared with the untreated, control animals.

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### Example 9. Morphogen Effect on Fibrogenesis and Scar Tissue Formation

The morphogens described herein induce tissue 10 morphogenesis of damaged or lost tissue. The ability of these proteins to regenerate new tissue enhances the anti-inflammatory effect of these proteins. below are a series of in vitro experiments demonstrating the ability of morphogens to induce migration and accumulation of mesenchymal cells. addition, the experiments demonstrate that morphogens, TGF-β, do not stimulate fibrogenesis or scar tissue formation. Specifically, morphogens do not stimulate production of collagen, hyaluronic acid (HA) 20 or metalloproteinases in primary fibroblasts, all of which are required for fibrogenesis or scar tissue formation. By contrast, TGF-β, a known inducer of fibrosis, but not of tissue morphogenesis, does stimulate production of these fibrosis markers.

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Chemotaxis and migration of mesenchymal progenitor cells were measured in modified Boyden chambers essentially as described by Fava, R.A. et al (1991) J. Exp. Med. 173: 1121-1132, the disclosure of which is incorporated herein by reference, using polycarbonate filters of 2, 3 and 8 micron ports to measure migration of progenitor neutrophils, monocytes and fibroblasts. Chemotaxis was measured over a range of morphogen concentrations, e.g.,  $10^{-20}$ M to  $10^{-12}$ M OP-1. For progenitor neutrophils and monocytes,  $10^{-18}$ - $10^{-17}$ M OP-1

consistently induced maximal migration, and 10<sup>-14</sup> to 10<sup>-13</sup> M OP-1 maximally induced migration of progenitor fibroblasts. In all cases the chemotactic activity could be inhibited with anti-OP-1 antibody. Similar migration activities also were measured and observed with TGF-β.

The effect of morphogen on fibrogenesis was determined by evaluating fibroblast production of hyaluronic acid (HA), collagen, collagenese and tissue inhibitor of metalloproteinases (TIMP).

Human fibroblasts were established from explants of infant foreskins and maintained in monolayer culture 15 using standard culturing procedures. (See, for example, (1976) J. Exp. Med. 144: 1188-1203.) Briefly, fibroblasts were grown in maintenance medium consisting of Eagle's MEM, supplemented with nonessential amino acids, ascorbic acid (50  $\mu$ g/ml), NaHCO<sub>3</sub> and HEPES 20 buffers (pH 7.2), penicillin (100 U/ml), streptomycin (100  $\mu$ g/ml), amphotericin B (1  $\mu$ g/ml) and 9% heat inactivated FCS. Fibroblasts used as target cells to measure chemotaxis were maintained in 150 mm diameter glass petri dishes. Fibroblasts used in assays to measure synthesis of collagen, hyaluronic acid, collagenase and tissue inhibitors of metalloproteinases (TIMP) were grown in 100 mm diameter plastic tissue culture petri dishes.

of hyaluronic acid, collagens, collagenase and TIMP were determined by standard assays (See, for example, Posttethwaite et al. (1989) <u>J. Clin. Invest</u>. 83: 629-636, Posttethwaithe (1988) <u>J./ Cell Biol</u>. 106: 311-318 and Clark et al (1985) <u>Arch. Bio-chem Biophys</u>. 241: 36-

44, the disclosures of which are incorporated by reference.) For these assays, fibroblasts were transferred to 24-well tissue culture plates at a density of 8 x 10<sup>4</sup> cells per well. Fibroblasts were 5 grown confluency in maintenance medium containing 9% FCS for 72 h and then grown in serum-free maintenance medium for 24 h. Medium was then removed from each well and various concentrations of OP-1 (recombinantly produced mature or soluble form) or TGF-β-1 (R&D 10 Systems, Minneapolis) in 50  $\mu$ l PBS were added to triplicate wells containing the confluent fibroblast monolayers. For experiments that measured production of collagenase and TIMP, maintenance medium (450  $\mu$ 1) containing 5% FCS was added to each well, and culture 15 supernatants were harvested from each well 48 h later and stored at -70°C until assayed. For experiments that assessed HA production, maintenance medium (450 μl) containing 2.5% FCS was added to each well, and cultures grown for 48 h. For experiments that measured 20 fibroblast production of collagens, serum-free maintenance medium (450  $\mu$ l) without non-essential amino acids was added to each well and cultures grown for 72 h. Fibroblast production of HA was measured by labeling newly synthesized glycosaminoglycans (GAG) 25 with [3H]-acetate the last 24 h of culture and quantitating released radioactivity after incubation with hyaluronidase from Streptomyces hyalurolyticus (ICN Biochemicals, Cleveland, OH) which specifically degrades hyaluronic acid. Production of total collagen 30 by fibroblasts was measured using a collagenasesensitive protein assay that reflects [3H]-proline incorporation the last 24 h of culture into newly synthesized collagens. Collagenase and TIMP protein levels in fibroblast cultures supernatants was measured 35 by specific ELISAs.

As shown in Fig. 6, OP1 does not stimulate significant collagen or HA production, as compared with TGF-β. In the figure, panel A shows OP-1 efect on collagen production, panel B shows TGF-β effect on collagen production, and panels C and D show OP-1 (panel C) and TGF-β (panel D) effect on HA production. The morphogen results were the same whether the soluble or mature form of OP1 was used. By contrast, the latent form of TGF-β (e.g., pro domain-associated form of TGF-β) was not active.

# Example 10. Morphogen Inhibition of Epithelial Cell Proliferation

This example demonstrates the ability of morphogens 15 to inhibit epithelial cell proliferation in vitro, as determined by 3H-thymidine uptake using culture cells from a mink lung epithelial cell line (ATCC No. CCL 64), and standard mammalian cell culturing procedures. 20 Briefly, cells were grown to confluency in Eagle's minimum essential medium (EMEM) supplemented with 10% fetal bovine serum (FBS), 200 units/ml penicillin, and 200  $\mu$ g/ml streptomycin, and used to seed a 48-well cell culture plate at a cell density of 200,000 cells per 25 well. When this culture became confluent, the media was replaced with 0.5 ml of EMEM containing 1% FBS and penicillin/streptomycin and the culture incubated for 24 hours at 37 C. Morphogen test samples in EMEM containing 5% FBS then were added to the wells, and the 30 cells incubated for another 18 hours. After incubation, 1.0  $\mu$ Ci of  $^3$ H-thymidine in 10  $\mu$ l was added to each well, and the cells incubated for four hours at The media then was removed and the cells washed once with ice-cold phosphate-buffer saline and DNA 35 precipitated by adding 0.5 ml of 10% TCA to each well

and incubating at room temperature of 15 minutes. The cells then were washed three times with ice-cold distilled water, lysed with 0.5 ml 0.4 M NaOH, and the lysate from each well then transferred to a scintillation vial and the radioactivity recorded using a scintillation counter (Smith-Kline Beckman).

The results are presented in Table III, below. The anti-proliferative effect of the various morphogens 10 tested was expressed as the counts of 3H-thymidine (x 1000) integrated into DNA, and were compared with untreated cells (negative control) and TGF-\$ (1 ng), a local-acting factor also known to inhibit epithelial cell proliferation. COP-5 and COP-7 are biosynthetic 15 constructs that previously have been shown to have osteogenic activity, capable of inducing the complete cascade resulting in endochondral bone formation in a standard rat bone assay (see U.S. Pat. No. 5,011,691.) The morphogens significantly inhibit epithelial cell 20 proliferation. Similar experiments, performed with the morphogens COP-16, bOP (bone-purified osteogenic protein, a dimeric protein comprising CBMP2 and OP-1), and recombinant OP-1, also inhibit cell proliferation. bOP and COP-16 also induce endochondral bone formation 25 (see US Pat. No. 4,968,590 and 5,011,691.)

#### TABLE III

		Thymidine uptake (x 1000)
30	control	50.048, 53.692
	COP-7-1 (10 ng)	11.874
	COP-7-2 (3 ng)	11.136
	COP-5-1 (66 ng)	16.094
	COP-5-2 (164 ng)	14.43
35	TGF-β (1 ng)	1.86, 1.478

## Example 11. Morphogen Treatment of a Systemic Inflammatory Disease

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The following example provides a rat adjuvantinduced arthritis model for demonstrating morphogen
efficacy in treating arthritis and other systemic
inflammatory diseases. Rat adjuvant-induced arthritis
induces a systemic inflammatory disease with bone and
cartilage changes similar to those observed in
rhematoid arthritis, but in an accelerated time span
(see, for example, Pearson (1964) Arth. Rheum. 7:80).
A detailed description of the protocol is provided in
Walz, et al., (1971) J. Pharmac. Exp. Ther. 178: 223231, the disclosure of which is incorporated herein by
reference.

Briefly, Sprague-Dawley female rats (e.g., Charles River Laboratories, Wilmington, MA) are randomized into 3 groups: control; morphogen, low dose (e.g., 1-  $10 \mu g/kg$  weight per day) and morphogen, high dose (e.g.,  $10-20 \mu g/kg$  weight per day), referred to as Groups 1, 2, and 3, respectively.

25

Adjuvant arthritis is induced in all three groups by injection of 0.05 ml of a suspension of 1.5% dead Mycobacterium butyricum in mineral oil into the subplantar surface of the right hand paw. On Day 18 after adjuvant injection, the limb volumes of both hind limb are determined. In the absence of morphogen treatment, a systemic arthritic condition is induced in adjuvant-injected rats by this time, as determined by significant swelling of the uninjected hind limbs (< 35 2.3 ml, volume measured by mercury displacement).

Subsequent determinations of paw edema and x-ray scores are made on the uninjected hind limb. Rats in Group 2 and 3 also are dosed orally daily, beginning on Day 1, with morphogen. Limb volumes are recorded on Days 29 and 50 after adjuvant injection and edema determined by volume difference compared to Day 18. The uninjected hind limb on each rat is x-rayed on Day 50 and the joint damage assayed on an arbitrary scale of 1 to 10 (1=no damage, 10=maximum damage). Data on differences between control and treated groups (Day 29 edema, Day 50 edema and Day 50 x-ray scores) are analyzed by using a standard "t-test. Morphogen-treated rats show consistently reduced joint damage (e.g., decreased in edema and in x-ray scores) as compared with untreated control rats.

As another, alternative example, Groups 2 and 3 are dosed daily with morphogen beginning on Day 18 and continuing through Day 50 to demonstrate the efficacy of morphogens in arthritic animals.

#### Example 12. Morphogen Inhibition of Localized Edema

The following example demonstrates morphogen
25 efficacy in inhibiting a localized inflammatory
response in a standard rat edema model. Experimental
rats (e.g., Long-Evans from Charles River Laboratories,
Wilmington, MA) are divided into three groups: Group
1, a negative control, which receives vehicle alone;
30 Group 2, a positive control, to which is administered a
well-known characterized anti-inflammatory agent
(e.g., indomethacin), and Group 3, to which morphogen
is provided.

Groups 2 and 3 may be further subdivided to test low, medium and high doses (e.g., Group 2: 1.0 mg/kg, 3.0 mg/kg and 9.0 mg/kg indomethacin; Group 3: 0.1-5μg; 5-20μg, and 20-50μg of morphogen). Sixty minutes after indomethacin or morphogen is provided to the rats of Group 2 or 3 (e.g., as by injection into the tail vein, or by oral gavage) inflammation is induced in all rats by a sub-plantar injection of a 1% carrageenin solution (50μl) into the right hind paw. Three hours after carrageenin administration paw thickness is measured as an indication of edema (e.g., swelling) and induced inflammatory response to the injected carrageenin solution.

- 15 Significant swelling is evident in untreated rats by three hours after carrageenin injection.

  Inflammation also is measured by histology by standard means, following euthanasia e.g.: the right hind paw from each animal is removed at the ankle joint and weighed and foot pad tissue is fixed in 10% neutral buffered formalin, and slides prepared for visual examination by staining the prepared tissue with hematoxylin and eosin.
- The morphogen-treated rats show substantially reduced edema induction following carrageenin injection as compared with the untreated rats.

### Example 13. Morphogen Treatment of Allergic Encephalomyelitis

The following example demonstrates morphogen

5 efficacy in treating experimental allergic
encephalomyelitis (EAE) in a rat. EAE is a
well-characterized animal model for multiple sclerosis,
an autoimmune disease. A detailed description of the
protocol is disclosed in Kuruvilla, et al., (1991) PNAS

10 88:2918-2921, the disclosure of which is incorporated
herein by reference.

Briefly, EAE is induced in rats (e.g., Long-Evans, Charles River Laboratories, Wilmington, MA) by

15 injection of a CNS tissue (e.g., spinal cord)
homogenate in complete Freund's adjuvant (CFA) on days
-44, -30 and 0 (last day of immunization), by
subcutaneous injection to three sites on the animal's
back. Morphogen is administered daily by

20 interperitoneal injection beginning on day -31.
Preferably, a series of morphogen dose ranges is
evaluated (e.g., low, medium and high) as for
Example 12, above.) Control rats receive morphogen
vehicle only (e.g. 0.9% NaCl or buffered saline). Rats

25 are examined daily for signs of disease and graded on
an increasing severity scale of 0-4.

In the absence of morphogen treatment, significant neurological dysfunction (e.g., hind and fore limb weakness, progressing to total hind limb paralysis) is evident by day +7 to +10. Hematology, serum chemistry profiles and histology are performed to evaluate the

degree of tissue necropsy using standard procedures.

Morphogen treatment significantly inhibits the
neurological dysfunction normally evident in an EAE
animal. In addition, the histopathological markers
typically associated with EAE are absent in the
morphogen-treated animals.

## Example 14. Morphogen Treatment of Collagen-Induced Arthritis

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The following example demonstrates the efficacy of morphogens in inhibiting the inflammatory response in a collagen-induced arthritis (CIA) in a rat. CIA is a well-characterized animal model for rheumatoid 15 arthritis, an autoimmune disease. The protocol disclosed is essentially that disclosed in Kuruvilla et al., (1991) PNAS 88:2918-2921, incorporated by reference hereinabove. Briefly, CIA is induced in experimental rats (e.g., Long-Evans, Charles River 20 Laboratories, Wilmington), by multiple intradermal injection of bovine Type II collagen (e.g.,  $100\mu g$ ) in CFA (0.2 ml) on Day 1. Animals are divided into two groups: Group 1, control animals, which receive vehicle alone, and Group 2: morphogen-treated animals, which, preferably, are subdivided into low, medium and high dose ranges, as described for Example 13, above. Morphogen is administered daily (e.g., by tail vein injection) beginning at different times following collagen injection, e.g., beginning on day 7, 14, 28, 35 and 42. Animals are evaluated visually and paw thickness and body weight is monitored throughout the experiment. Animals are sacrificed on day 60 and the proximal and distal limb joints, and ear, tail and spinal cord prepared for histological evaluation as described for Examples 12 and 13, above. In a

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variation of the experiment, morphogen may be administered for prescribed periods, e.g., five day periods, beginning at different times following collagen injection (e.g., on days 0-4, 7-11, 14-18, 28-32.)

In the absence of morphogen treatment, an arthritic condition typically is induced by 30 days post collagen injection. In morphogen-treated animals, CIA is suppressed and the histopathological changes typically evidenced in control CIA-induced animals are absent: e.g., accumulations of activated mononuclear inflammatory cells and fibrous connective tissue. In addition, consistent with the results in Example 7, above, serum anti-collagen antibody titers are suppressed significantly in the morphogen-treated animals.

Example 15. Screening Assay for Candidate Compounds which Alter Endogenous Morphogen Levels

Candidate compound(s) which may be administered to affect the level of a given morphogen may be found using the following screening assay, in which the level of morphogen production by a cell type which produces measurable levels of the morphogen is determined with and without incubating the cell in culture with the compound, in order to assess the effects of the compound on the cell. This can be accomplished by detection of the morphogen either at the protein or RNA level. A more detailed description also may be found in USSN 752,861, incorporated hereinabove by reference.

### 15.1 Growth of Cells in Culture

Cell cultures of kidney, adrenals, urinary bladder, brain, or other organs, may be prepared as described 5 widely in the literature. For example, kidneys may be explanted from neonatal or new born or young or adult rodents (mouse or rat) and used in organ culture as whole or sliced (1-4 mm) tissues. Primary tissue cultures and established cell lines, also derived from 10 kidney, adrenals, urinary, bladder, brain, mammary, or other tissues may be established in multiwell plates (6 well or 24 well) according to conventional cell culture techniques, and are cultured in the absence or presence of serum for a period of time (1-7 days). Cells may be 15 cultured, for example, in Dulbecco's Modified Eagle medium (Gibco, Long Island, NY) containing serum (e.g., fetal calf serum at 1%-10%, Gibco) or in serum-deprived medium, as desired, or in defined medium (e.g., containing insulin, transferrin, glucose, albumin, or 20 other growth factors).

Samples for testing the level of morphogen production includes culture supernatants or cell lysates, collected periodically and evaluated for OP-1 production by immunoblot analysis (Sambrook et al., eds., 1989, Molecular Cloning, Cold Spring Harbor Press, Cold Spring Harbor, NY), or a portion of the cell culture itself, collected periodically and used to prepare polyA+ RNA for RNA analysis. To monitor de novo OP-1 synthesis, some cultures are labeled according to conventional procedures with an 55 S-methionine/35 S-cysteine mixture for 6-24 hours and then evaluated to OP-1 synthesis by conventional immunoprecipitation methods.

### 15.2 Determination of Level of Morphogenic Protein

In order to quantitate the production of a morphogenic protein by a cell type, an immunoassay may be performed to detect the morphogen using a polyclonal or monoclonal antibody specific for that protein. For example, OP-1 may be detected using a polyclonal antibody specific for OP-1 in an ELISA, as follows.

1  $\mu$ g/100  $\mu$ l of affinity-purified polyclonal rabbit 10 IgG specific for OP-1 is added to each well of a 96-well plate and incubated at 37°C for an hour. wells are washed four times with 0.167M sodium borate buffer with 0.15 M NaCl (BSB), pH 8.2, containing 0.1% 15 Tween 20. To minimize non-specific binding, the wells are blocked by filling completely with 1% bovine serum albumin (BSA) in BSB and incubating for 1 hour at 37°C. The wells are then washed four times with BSB containing 0.1% Tween 20. A 100  $\mu$ l aliquot of an 20 appropriate dilution of each of the test samples of cell culture supernatant is added to each well in triplicate and incubated at 37°C for 30 min. After incubation, 100  $\mu$ l biotinylated rabbit anti-OP-1 serum (stock solution is about 1 mg/ml and diluted 1:400 in 25 BSB containing 1% BSA before use) is added to each well and incubated at 37°C for 30 min. The wells are then washed four times with BSB containing 0.1% Tween 20. 100  $\mu$ l strepavidin-alkaline (Southern Biotechnology Associates, Inc. Birmingham, Alabama, diluted 1:2000 in 30 BSB containing 0.1% Tween 20 before use) is added to each well and incubated at 37°C for 30 min. are washed four times with 0.5M Tris buffered Saline (TBS), pH 7.2.  $50\mu$ l substrate (ELISA Amplification System Kit, Life Technologies, Inc., Bethesda, MD) is 35 added to each well incubated at room temperature for 15 min. Then, 50 μl amplifier (from the same amplification system kit) is added and incubated for another 15 min at room temperature. The reaction is stopped by the addition of 50 μl 0.3 M sulphuric acid.
5 The OD at 490 nm of the solution in each well is recorded. To quantitate OP-1 in culture media, a OP-1 standard curve is performed in parallel with the test samples.

Polyclonal antibody may be prepared as follows. 10 Each rabbit is given a primary immunization of 100 ug/500  $\mu$ l E. coli produced OP-1 monomer (amino acids 328-431 in SEQ ID NO:5) in 0.1% SDS mixed with 500  $\mu$ l Complete Freund's Adjuvant. The antigen is injected 15 subcutaneously at multiple sites on the back and flanks of the animal. The rabbit is boosted after a month in the same manner using incomplete Freund's Adjuvant. Test bleeds are taken from the ear vein seven days later. Two additional boosts and test bleeds are 20 performed at monthly intervals until antibody against OP-1 is detected in the serum using an ELISA assay. Then, the rabbit is boosted monthly with 100  $\mu$ g of antigen and bled (15 ml per bleed) at days seven and ten after boosting.

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Monoclonal antibody specific for a given morphogen may be prepared as follows. A mouse is given two injections of <u>E. coli</u> produced OP-1 monomer. The first injection contains  $100\mu g$  of OP-1 in complete Freund's adjuvant and is given subcutaneously. The second injection contains  $50~\mu g$  of OP-1 in incomplete adjuvant and is given intraperitoneally. The mouse then receives a total of 230  $\mu g$  of OP-1 (amino acids 307-431 in SEQ ID NO:5) in four intraperitoneal injections at various times over an eight month period. One week

prior to fusion, both mice are boosted intraperitoneally with 100 µg of OP-1 (307-431) and 30 µg of the N-terminal peptide (Ser<sub>293</sub>-Asn<sub>309</sub>-Cys) conjugated through the added cysteine to bovine serum albumin with SMCC crosslinking agent. This boost was repeated five days (IP), four days (IP), three days (IP) and one day (IV) prior to fusion. The mouse spleen cells are then fused to myeloma (e.g., 653) cells at a ratio of 1:1 using PEG 1500 (Boeringer Mannheim), and the cell fusion is plated and screened for OP-1-specific antibodies using OP-1 (307-431) as antigen. The cell fusion and monoclonal screening then are according to standard procedures well described in standard texts widely available in the art.

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The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The present embodiments are therefore to be considered in all respects as illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced therein.

### SEQUENCE LISTING

_	(1)	GENERAL	INFOR	MATION:
5 10		(i)APPLIC	ANT:	KUBERASAHPATH, THANGAVEL PANG, ROY H.L. OPPERMANN, HERMANN RUEGER, DAVID C. COHEN, CHARLES H.
				OZKAYNAK, ENGIN SHART, JOHN
15		(ii)	TITL	E OF INVENTION: HORPHOGEN-INDUCED HODULATION OF AHMATORY RESPONSE
		(iii)	NUMB	ER OF SEQUENCES: 33
20		(iv)	(A) (B) (C) (D)	ESPONDENCE ADDRESS: ADDRESSEE: CREATIVE BIOHOLECULES STREET: 35 SOUTH STREET CITY: HOPKINTON STATE: MASSACHUSETTS COUNTRY: U.S.A.
25			(F)	ZIP:
30		(▽)	(A) (B)	JTER READABLE FORM: MEDIUM TYPE: Floppy disk COMPUTER: IBM PC compatible OPERATING SYSTEM: PC-DOS/MS-DOS SOFTWARE: Patent In Release #1.0, Version #1.25
35		(vii)	(A)	R APPLICATION DATA: APPLICATION NUMBER: US 667,274 FILING DATE: 11-MAR-1991
40		(vii)	(A)	R APPLICATION DATA: APPLICATION NUMBER: US 753,059 FILING DATE: 30-AUG-1991
40		(vii)	(A)	R APPLICATION DATA: APPLICATION NUMBER: US 752,764 FILING DATE: 30-AUG-1991
45		(2)	•	INFORMATION FOR SEQ ID NO:1:
4.7		i)	•	SEQUENCE CHARACTERISTICS: (A) LENGTH: 97 amino acids (B) TYPE: amino acids (C) TOPOLOGY: linear
50		i )	Li)	MOLECULE TYPE: protein

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                     (A) LENGTH: 139 amino acids
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              (ix)
                                mOP-1 (mature form)
                     (A) NAME:
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		Val	His	Phe	Ile	Asn 95	Pro	Asp	Thr	Val
15		Pro 100	Lys	Pro	Cys	Cys	Ala 105	Pro		Gln
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20		Asp	Asp	Ser 120	Ser	Asn	Val	Ile	Leu 125	Lys
		Lys	Tyr	Arg	Asn 130	Met	Val	Val	Arg	Ala 135
		Cys	Gly	Cys	His					
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30		(ii) (ix) (xi)	MOL FEA (A)	ECULE TURE: NAME	: hO	P-2 (RIPTI	matur	e for	m) DNO:	7:
35		Ala 1	Val	Arg	Pro	Leu 5	Arg	Arg	Arg	Gln
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		Asp	Val	His 30	Gly	Ser	His	Gly	Arg 35	Gln
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	Leu	Ser 110	Ala	Thr	Ser	Val	Leu 115	Tyr	Tyr	ŝ
10	Asp	Ser	Ser 120	Asn	Asn	Val	Ile	Leu 125	Arg	÷
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2 E		100	Cys	Arg								
25	(2)	INFO	(2	EQUE!	FOR NCE ( ENGTI (PE:	CHARI	ACTEI	RIST:	CS:	ids		
30		(ii) (ix)	(( ) M( ) FI	C) TO DLECU EATUR	OPOLO	OGY: TYPE:	lin pr	near rote:				
35		(xi	) ŠI	EQUE	NCE I	DESČI	RIPTI	ON:		O ID		
-		1	Lys			5					10	
			Val		15					20		
40			Gly	25					30			
		•	Ser 35					40				
15		45	Asn				50					55
			Met			60					65	
		Cys	Ala	Pro	Thr	Lys	Val	Asn	АТА	11e	ser	vaı

40

Ala Ala Ala Pro Gly Ala Ala Asp Leu Pro Cys Cys Val Pro Ala

Arg Leu Ser Pro Ile Ser Val Leu Phe Phe Asp Asn Ser Asp Asn

Val Val Leu Arg Gln Tyr Glu Asp Met Val Val Asp Glu Cys Gly 45 95 90

> Cys Arg 105

	(2) INFORMATION FOR SEQ ID NO:15:	
5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 5 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
10	(ii) MOLECULE TYPE: peptide	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
1 E	Cys Xaa Xaa Xaa 1 5	
15	(2) INFORMATION FOR SEQ ID NO:16:	
20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 1822 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
25	(ii) MOLECULE TYPE: cDNA	
	(vi) ORIGINAL SOURCE:  (A) ORGANISM: HOMO SAPIENS  (F) TISSUE TYPE: HIPPOCAMPUS	
30	<pre>(ix) FEATURE:     (A) NAME/KEY: CDS     (B) LOCATION: 491341     (D) OTHER INFORMATION:/standard_name= "hOP1"</pre>	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	
	GGTGCGGGCC CGGAGCCCGG AGCCCGGGTA GCGCGTAGAG CCGGCGCG ATG CAC GTG  Het His Val  1	57
10	CGC TCA CTG CGA GCT GCG GCG CCG CAC AGC TTC GTG GCG CTC TGG GCA Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala Leu Trp Ala 5 10 15	05
15	CCC CTG TTC CTG CTG CGC TCC GCC CTG GCC GAC TTC AGC CTG GAC AAC Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser Leu Asp Asn 20 25 30 35	53
50	GAG GTG CAC TCG AGC TTC ATC CAC CGG CGC CTC CGC AGC CAG GAG CGG Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser Gln Glu Arg	01

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	AAG Lys 260	Leu	GCG Ala	GGC Gly	CTG Leu	ATT Ile 265	GGG Gly	CGG Arg	CAC His	GGG Gly	CCC Pro 270	CAG Gln	AAC Asn	AAG Lys	CAG Gln	CCC Pro 275	873
5	TTC Phe	ATG Het	GTG Val	GCT Ala	TTC Phe 280	TTC Phe	AAG Lys	GCC Ala	ACG Thr	GAG Glu 285	GTC Val	CAC His	TTC Phe	CGC Arg	AGC Ser 290	ATC Ile	921
10	CGG Arg	TCC Ser	ACG Thr	GGG Gly 295	AGC Ser	AAA Lys	CAG Gln	CGC Arg	AGC Ser 300	CAG Gln	AAC Asn	CGC Arg	TCC Ser	AAG Lys 305	ACG Thr	CCC Pro	969
15	AAG Lys	AAC Asn	CAG Gln 310	GAA Glu	GCC Ala	CTG Leu	CGG Arg	ATG Met 315	GCC Ala	AAC Asn	GTG Val	GCA Ala	GAG Glu 320	AAC Asn	AGC Ser	AGC Ser	1017
	AGC Ser	GAC Asp 325	CAG Gln	AGG Arg	CAG Gln	GCC Ala	TGT Cys 330	AAG Lys	AAG Lys	CAC His	GAG Glu	CTG Leu 335	TAT Tyr	GTC Val	AGC Ser	TTC Phe	1065
20	CGA Arg 340	GAC Asp	CTG Leu	GGC Gly	TGG Trp	CAG Gln 345	GAC Asp	TGG Trp	ATC Ile	ATC Ile	GCG Ala 350	CCT Pro	GAA Glu	GGC Gly	TAC Tyr	GCC Ala 355	1113
25	GCC Ala	TAC Tyr	TAC Tyr	TGT Cys	GAG Glu 360	GGG Gly	GAG Glu	TGT Cys	GCC Ala	TTC Phe 365	CCT Pro	CTG Leu	AAC Asn	TCC Ser	TAC Tyr 370	ATG Met	1161
30	AAC Asn	GCC Ala	ACC Thr	AAC Asn 375	CAC His	GCC Ala	ATC Ile	GTG Val	CAG Gln 380	ACG Thr	CTG Leu	GTC Val	CAC His	TTC Phe 385	ATC Ile	AAC Asn	1209
35	CCG Pro	GAA Glu	ACG Thr 390	GTG Val	CCC Pro	AAG Lys	CCC Pro	TGC Cys 395	TGT Cys	GCG Ala	CCC Pro	ACG Thr	CAG Gln 400	CTC Leu	AAT Asn	GCC Ala	1257
	ATC Ile	TCC Ser 405	GTC Val	CTC Leu	TAC Tyr	TTC Phe	GAT Asp 410	GAC Asp	AGC Ser	TCC Ser	AAC Asn	GTC Val 415	ATC Ile	CTG Leu	AAG Lys	AAA Lys	1305
40	TAC Tyr 420	AGA Arg	AAC Asn	ATG Ket	GTG Val	GTC Val 425	CGG Arg	GCC Ala	TGT Cys	GGC Gly	TGC Cys 430	CAC His	TAGO	CTCCI	CC		1351
45	GAGA	ATTC	AG A	CCCI	TTG	G GC	CAAG	TTTT	TCI	GGAT	CCT	CCAT	TGCI	rcg c	CTT	GCCAG	1411
	GAAC	CAGO	AG A	CCAA	CTGC	C TI	TTGI	GAGA	CCI	TCCC	CTC	CCTA	TCCC	CA A	CTTI	'AAAGG	1471
	TGTG	AGAG	I AT	TAGG	AAAC	A TG	AGCA	GCAT	' ATC	GCTI	TTG	ATCA	GTTI	TT C	AGTG	GCAGC	1531
50	ATCC	AATG	AA C	AAGA	TCCI	'A CA	AGCI	GTGC	AGG	CAAA	LACC	TAGO	AGGA	AA A	AAAA	ACAAC	1591

										٠.								1/51
																CGGACI		1651
	CGT	TTCC	AGA (	GGTA	ATTA:	IG A	GCGC	CTAC	C AG	CCAG	GCCA	CCC	AGCC	GTG	GGAG	GAAGGG	ř	1711
5	GGC	GTGG	CAA (	GGGG'	TGGG(	CA CA	ATTG(	GTGT(	C TG:	rgcg/	AAAG	GAA	TTAA	GAC	CCGG	AAGTTC	,	1771
	CTG	TAAT	AAA :	TGTC.	ACAA:	TA AI	AACG/	AATG	A AT	SAAA	AAAA	AAA	AAAA	AAA	A			1822
10	(2)	IN	FORM															
		٠	<b>(i</b> )	(4 (1	A) 1 B) :	LENG:	CHARA TH: 4 : am: LOGY:	431 a ino a	amino acid	ICS:	ids					í		
15			(ii)	•	•		TYPE:			ı			-					
20			(ix)	(1	•	THER	INF(				Produ			-PP"		v.		
			(xi)	•	•									Phe	Val	Ala		
	Met 1	His	Val	Arg	Ser 5	ren	ALG	VIG	NTG	10	110				15			
25	Leu	Trp	Ala	Pro 20	Leu	Phe	Leu	Leu	Arg 25	Ser	Ala	Leu	Ala	Asp 30	Phe	Ser		
30	Leu	Asp	Asn 35	Glu	Val	His	Ser	Ser 40	Phe	Ile	His	Arg	Arg 45	Leu	Arg	Ser		
	Gln	Glu 50	Arg	Arg	Glu	Met	Gln 55	Arg	Glu	Ile	Leu	Ser 60	Ile	Leu	Gly	Leu		÷
35	Pro 65	His	Arg	Pro	Arg	Pro 70	His	Leu	Gln	Gly	Lys 75	His	Asn	Ser	Ala	Pro 80		
	Het	Phe	Met	Leu	Asp 85	Leu	Tyr	Asn	Ala	Met 90	Ala	Val	Glu	Glu	Gly 95	Gly		
40	Gly	Pro	Gly	Gly LOO	Gln	Gly	Phe	Ser	Tyr 105	Pro	Tyr	Lys	Ala	Val 110	Phe	Ser		
45			115					120					123		Leu			
		130					135					140			Asp			
50	Glu 145	Phe	Phe	His	Pro	Arg 150	Tyr	His	His	Arg	Glu 155	Phe	Arg	Phe	Asp	Leu 160		

	Ser	Lys	Ile	Pro	Glu 165	Gly	Glu	Ala	Val	Thr 170	Ala	Ala	Glu	Phe	Arg 175	Ile
5	Tyr	Lys		Tyr 180	Ile	Arg	Glu	Arg	Phe 185	Asp	Asn	Glu	Thr	Phe 190	Arg	Ile
	Ser	Val	Tyr 195	Gln	Val	Leu	Gln	Glu 200	His	Leu	Gly	Arg	Glu 205	Ser	Asp	Leu
10	Phe	Leu 210	Leu	Asp	Ser	Arg	Thr 215	Leu	Trp	Ala	Ser	Glu 220	Glu	Gly	Trp	Leu
1 5	Val 225	Phe	Asp	Ile	Thr	Ala 230	Thr	Ser	Asn	His	Trp 235	Val	Val	Asn	Pro	Arg 240
15	His	Asn	Leu	Gly	Leu 245	Gln	Leu	Ser	Val	Glu 250	Thr	Leu	Asp	Gly	Gln 255	Ser
20	Ile	Asn	Pro	Lys 260	Leu	Ala	Gly	Leu	Ile 265	Gly	Arg	His	Gly	Pro 270	Gln	Asn
	Lys	Gln	Pro 275	Phe	Het	Val	Ala	Phe 280	Phe	Lys	Ala	Thr	Glu 285	Val	His	Phe
25	Arg	Ser 290	Ile	Arg	Ser	Thr	Gly 295	Ser	Lys	Gln	Arg	Ser 300	Gln	Asn	Arg	Ser
30	Lys 305	Thr	Pro	Lys	Asn	Gln 310	Glu	Ala	Leu	Arg	Met 315	Ala	Asn	Val	Ala	Glu 320
30	Asn	Ser	Ser	Ser	Asp 325	Gln	Arg	Gln	Ala	Cys 330	Lys	Lys	His	Glu	<b>Leu</b> 335	Tyr
35	Val	Ser		Arg	Asp	Leu	Gly	Trp	Gln 345	Asp	Trp	Ile	Ile	Ala 350	Pro	Glu
	Gly	Tyr	Ala 355	Ala	Tyr	Tyr	Cys	Glu 360	Gly	Glu	Cys	Ala	Phe 365	Pro	Leu	Asn
40	Ser	Tyr 370	Het	Asn	Ala	Thr	Asn 375	His	Ala	Ile	Val	Gln 380	Thr	Leu	Val	His
15	Phe 385	Ile	Asn	Pro	Glu	Thr 390	Val	Pro	Lys	Pro	Cys 395	Cys	Ala	Pro	Thr	Gln 400
. J	Leu	Asn	Ala	Ile	Ser 405	Val	Leu	Tyr	Phe	Asp 410	Asp	Ser	Ser	Asn	Val 415	Ile
<b>-</b> 0	Leu	Lys	Lys ,	Tyr	Arg	Asn	Het	Val	Val	Arg	Ala	Cys	Gly	Cys 430	His	

	(2) INFORMAT	ION FOR SEQ ID NO:18:	
5	(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 1873 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
10	(ii) (vi)	HOLECULE TYPE: cDNA ORIGINAL SOURCE:	
	(12)	(A) ORGANISH: HURIDAE (F) TISSUE TYPE: EMBRYO	1
15	(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1041393 (D) OTHER INFORMATION: /not	e= "MOP1 (CDNA)"
20	•	SEQUENCE DESCRIPTION: SEQ ID	
		ACCTCGGG TCGTGGACCG CTGCCCTGC	
25	CGGCGCGGGC CC	GGTGCCCC GGATCGCGCG TAGAGCCGG	C GCG ATG CAC GTG CGC 115  Met His Val Arg  1
30	TCG CTG CGC G Ser Leu Arg A 5	CT GCG GCG CCA CAC AGC TTC GT la Ala Ala Pro His Ser Phe Va 10	I Ala Leu IIP Ala Pro
	CTG TTC TTG C Leu Phe Leu L	TG CGC TCC GCC CTG GCC GAT TT Eu Arg Ser Ala Leu Ala Asp Ph 25 30	C AGC CTG GAC AAC GAG e Ser Leu Asp Asn Glu 35
35	Val His Ser Se	CC TTC ATC CAC CGG CGC CTC CG er Phe Ile His Arg Arg Leu Ar 0 45	C AGC CAG GAG CGG CGG 259 g Ser Gln Glu Arg Arg 50
40	GAG ATG CAG Co	G GAG ATC CTG TCC ATC TTA GG g Glu Ile Leu Ser Ile Leu Gl 60	G TTG CCC CAT CGC CCG 307 y Leu Pro His Arg Pro 65
45	CGC CCG CAC C Arg Pro His Lo	C CAG GGA AAG CAT AAT TCG GC ou Gln Gly Lys His Asn Ser Al 75	G CCC ATG TTC ATG TTG 355 a Pro Met Phe Met Leu 80
50	GAC CTG TAC AS Asp Leu Tyr As 85	C GCC ATG GCG GTG GAG GAG AG n Ala Met Ala Val Glu Glu Se 90	F GIV Pro ASP GIV GIN

	GGC Gly	TTC Phe	TCC Ser	TAC Tyr	CCC Pro 105	TAC Tyr	AAG Lys	GCC Ala	GTC Val	TTC Phe 110	AGT Ser	ACC Thr	CAG Gln	GGC Gly	CCC Pro 115	CCT Pro		451
5	TTA Leu	GCC Ala	AGC Ser	CTG Leu 120	CAG Gln	GAC Asp	AGC Ser	CAT His	TTC Phe 125	CTC Leu	ACT Thr	GAC Asp	GCC Ala	GAC Asp 130	ATG Met	GTC Val		499
10	ATG Met	AGC Ser	TTC Phe 135	GTC Val	AAC Asn	CTA Leu	GTG Val	GAA Glu 140	CAT His	GAC Asp	AAA Lys	GAA Glu	TTC Phe 145	TTC Phe	CAC His	CCT Pro		547
	CGA Arg	TAC Tyr 150	CAC	CAT His	CGG Arg	GAG Glu	TTC Phe 155	CGG	TTT Phe	GAT Asp	CTT Leu	TCC Ser 160	AAG Lys	ATC Ile	CCC Pro	GAG Glu		595
15	GGC Gly 165	GAA Glu	CGG Arg	GTG Val	ACC Thr	GCA Ala 170	GCC Ala	GAA Glu	TTC Phe	AGG Arg	ATC Ile 175	TAT Tyr	AAG Lys	GAC Asp	TAC Tyr	ATC Ile 180	·	643
20	CGG Arg	GAG Glu	CGA Arg	TTT Phe	GAC Asp 185	AAC Asn	GAG Glu	ACC Thr	TTC Phe	CAG Gln 190	ATC Ile	ACA Thr	GTC Val	TAT Tyr	CAG Gln 195	GTG Val		<b>691</b>
25	CTC Leu	CAG Gln	GAG Glu	CAC His 200	TCA Ser	GGC Gly	AGG Arg	GAG Glu	TCG Ser 205	GAC Asp	CTC Leu	TTC Phe	TTG Leu	CTG Leu 210	GAC Asp	AGC Ser		739
30	CGC Arg	ACC Thr	ATC Ile 215	TGG Trp	GCT Ala	TCT Ser	GAG Glu	GAG Glu 220	GGC Gly	TGG Trp	TTG Leu	GTG Val	TTT Phe 225	GAT Asp	ATC Ile	ACA Thr		787
	GCC Ala	ACC Thr 230	AGC Ser	AAC Asn	CAC His	TGG Trp	GTG Val 235	GTC Val	AAC Asn	CCT Pro	CGG Arg	CAC His 240	AAC Asn	CTG Leu	GGC Gly	TTA Leu		835
35	CAG Gln 245	CTC Leu	TCT Ser	GTG Val	GAG Glu	ACC Thr 250	CTG Leu	GAT Asp	GGG Gly	CAG Gln	AGC Ser 255	ATC Ile	AAC Asn	CCC Pro	AAG Lys	TTG Leu 260		883
40	GCA Ala	GGC Gly	CTG Leu	ATT Ile	GGA Gly 265	CGG Arg	CAT His	GGA Gly	CCC Pro	CAG Gln 270	AAC Asn	AAG Lys	CAA Gln	CCC Pro	TTC Phe 275	ATG Net		931
45	GTG Val	GCC Ala	TTC Phe	TTC Phe 280	AAG Lys	GCC Ala	ACG Thr	GAA Glu	GTC Val 285	CAT His	CTC Leu	CGT Arg	AGT Ser	ATC Ile 290	CGG	TCC Ser		979
50	ACG Thr	GGG Gly	GGC Gly 295	AAG Lys	CAG Gln	CGC Arg	AGC Ser	CAG Gln 300	AAT Asn	CGC Arg	TCC Ser	AAG Lys	ACG Thr 305	CCA Pro	AAG Lys	AAC Asn	1	027

	CAA Gln	GAG Glu 310	Ala	CTG Leu	AGG Arg	ATG Het	GCC Ala 315	AGT Ser	GTG Val	GCA Ala	GAA Glu	AAC Asn 320	AGC Ser	AGC Ser	AGT Ser	GAC Asp	r	1075
5	CAG Gln 325	AGG Arg	CAG Gln	GCC Ala	TGC Cys	AAG Lys 330	AAA Lys	CAT His	GAG Glu	CTG Leu	TAC Tyr 335	AST	AGC Ser	TTC Phe	CGA Arg	GAC Asp 340		1123
10	CTT Leu	GGC Gly	TGG Trp	CAG Gln	GAC Asp 345	TGG Trp	ATC Ile	ATT Ile	GCA Ala	CCT Pro 350	GAA Glu	GGC Gly	TAT Tyr	GCT Ala	GCC Ala 355	TAC		1171
15	TAC Tyr	TGT Cys	Glu	GGA Gly 360	GAG Glu	TGC Cys	GCC Ala	TTC Phe	CCT Pro 365	CTG Leu	AAC Asn	TCC Ser	TAC Tyr	ATG Met 370	AAC Asn	GCC Ala	1	1219
	ACC Thr	AAC Asn	CAC His 375	GCC Ala	ATC Ile	GTC Val	CAG Gln	ACA Thr 380	CTG Leu	GTT Val	CAC His	TTC Phe	ATC Ile 385	W211	CCA Pro	GAC Asp		1267
20	ACA Thr	GTA Val 390	CCC Pro	AAG Lys	CCC Pro	TGC Cys	TGT Cys 395	GCG Ala	CCC Pro	ACC Thr	CAG Gln	CTC Leu 400	AAC Asn	GCC Ala	ATC Ile	TCT Ser		1315
25	GTC Val 405	CTC Leu	TAC Tyr	TTC Phe	GAC Asp	GAC Asp 410	AGC Ser	TCT Ser	AAT Asn	GTC Val	ATC Ile 415	ren	AAG Lys	AAG Lys	TAC Tyr	AGA Arg 420	,	1363
30	AAC Asn	ATG Het	GTG Val	GTC Val	CGG Arg 425	GCC Ala	TGT Cys	GGC Gly	TGC Cys	CAC His 430	TAGO	CTCTI	CC I	GAGA	CCCI	CG	-	1413
	ACCI	TTGC	GG G	GCCA	CACC	T TI	CCAA	ATCI	TCG	ATGI	CTC	ACCA	TCTA	AG I	CTCI	CAC'	<b>r</b> Ģ	1473
35	CCCA	CCTI	GG (	GAG	SAGAA	C AC	ACCA	ACCI	CTC	CTGA	.GCC	TTC	CTCA	CC 1	CCCA	ACC	GG	1533
	AAGO	ATGI	AA G	GGTT	CCAG	A AA	CCTG	AGCG	TGC	AGCA	GCT	GATG	AGCG	CC C	TITC	CTT	CT	1593
	GGCA	CGTG	AC C	GACA	AGAI	C CI	ACCA	GCTA	CCA	CAGO	AAA:	CGCC	TAAG	AG C	CAGGA	AAA	AT <sup>*</sup>	1653
40	GTCI	GCCA	.GG A	AAGI	GTCC	A GI	GTCC	ACAT	GGC	CCCI	GGC	GCTC	TGAG	TC I	TTGA	.GGA	GT	1713
	AATO	:GCAA	GC C	TCGI	TCAG	C TG	CAGO	AGAA	GGA	AGGG	CTT	AGCC	AGGG	TG 0	GCGC	TGG	CG	1773
45	TCTC	TGTI	GA A	\GGGA	AACC	A AG	CAGA	AGCC	ACT	GTAA	TGA	TATG	TCAC	AA I	AAAA	CCC	AT	1833
				AAAA														1873

(2) INFORMATION	FOR	SEQ	ID	NO:19:
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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 430 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- 10 (ix) FEATURE:
  (D) OTHER INFORMATION: /product= "mOP1-PP"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:
- 15 Met His Val Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala 1 5 10 15
  - Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser 20 25 30
- Leu Asp Asn Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser 35 40 45
- Gln Glu Arg Arg Glu Het Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu 25 50 55 60
  - Pro His Arg Pro Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro 65 70 75 80
- 30 Het Phe Het Leu Asp Leu Tyr Asn Ala Het Ala Val Glu Glu Ser Gly 85 90 95
  - Pro Asp Gly Gln Gly Phe Ser Tyr Pro Tyr Lys Ala Val Phe Ser Thr
- Gln Gly Pro Pro Leu Ala Ser Leu Gln Asp Ser His Phe Leu Thr Asp 115 120 125
- Ala Asp Met Val Met Ser Phe Val Asn Leu Val Glu His Asp Lys Glu 40 130 135 140
  - Phe Phe His Pro Arg Tyr His His Arg Glu Phe Arg Phe Asp Leu Ser 145 150 155 160
- 45 Lys Ile Pro Glu Gly Glu Arg Val Thr Ala Ala Glu Phe Arg Ile Tyr 165 170 175
  - Lys Asp Tyr Ile Arg Glu Arg Phe Asp Asn Glu Thr Phe Gln Ile Thr 180 185 190
- Val Tyr Gln Val Leu Gln Glu His Ser Gly Arg Glu Ser Asp Leu Phe
  195 200 205

	Leu	Leu 210	Asp	Ser	Arg	Thr	11e 215	Trp	ATA	Ser	GIU	220	<b>G1</b>	P	200	
5	225					230					233	Val				
	Asn	Leu	Gly	Leu	Gln 245	Leu	Ser	Val	Glu	Thr 250	Leu	Asp	Gly	Gln	Ser 255	Ile
10	Asn	Pro	Lys	Leu 260	Ala	Gly	Leu	Ile	Gly 265	Arg	His	Gly	Pro	Gln 270	Asn	Lys
	Gln	Pro	Phe 275	Het	Val	Ala	Phe	Phe 280	Lys	Ala	Thr	Glu	Val 285	His	Leu	Arg
15	Ser	Ile 290	Arg	Ser	Thr	Gly	Gly 295	Lys	Gln	Arg	Ser	Gln 300	Asn	Arg	Ser	Lys
20	Thr 305	Pro	Lys	Asn	Gln	Glu 310	Ala	Leu	Arg	Het	Ala 315	Ser	Val	Ala	Glu	Asn 320
	Ser	Ser	Ser	Asp	Gln 325	Arg	Gln	Ala	Cys	Lys 330	Lys	His	Glu	Leu	Tyr 335	Val
25	Ser	Phe	Arg	Asp 340	Leu	Gly	Trp	Gln	Asp 345	Trp	Ile	Ile	Ala	Pro 350	Glu	Gly
	Tyr	Ala	Ala 355	Tyr	Tyr	Cys	Glu	Gly 360	Glu	Cys	Ala	Phe	Pro 365	Leu	Asn	Ser
30	Tyr	Met 370	Asn	Ala	Thr	Asn	His 375	Ala	Ile	Val	Gln	Thr 380	Leu	Val	His	Phe
35	Ile 385	Asn	Pro	Asp	Thr	Val 390	Pro	Lys	Pro	Cys	Cys 395	Ala	Pro	Thr	Gln 100	Leu
	Asn	Ala	Ile	Ser	Val 405	Leu	Tyr	Phe	Asp	Asp 410	Ser	Ser	Asn	Val	Ile 415	Leu
40	Lys	Lys	Tyr	Arg	Asn	Het	Val	Val	Arg 425	Ala	Cys	Gly	Cys	His 430		
	(2)	INI	ORHA	TIOI	1 FOI	R SEC	] ID	NO: 2	20:							•
45		(i)	(A) (B)	LI TY	E CHA ENGTH PE: TRANI OPOLO	nuc] DEDNI	/23   Leic ESS:	acio sin	paı: İ	rs	•					
50		(ii	-		LE T				•							-

(vi	ORIGINAL	SOURCE:
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- (A) ORGANISH: Homo sapiens (F) TISSUE TYPE: HIPPOCAMPUS

## 5 (ix) FEATURE:

- (A) NAME/KEY: CDS
  (B) LOCATION: 490..1696
  (D) OTHER INFORMATION: /note= "hOP2 (cDNA)"

10	(xi)SEQUENCE DESCRIPTION: SEQ ID NO:20:	
	GGCGCCGGCA GAGCAGGAGT GGCTGGAGGA GCTGTGGTTG GAGCAGGAGG TGGCACGGCA	60
	GGGCTGGAGG GCTCCCTATG AGTGGCGGAG ACGGCCCCAGG AGGCGCTGGA GCAACAGCTC	120
15	CCACACCGCA CCAAGCGGTG GCTGCAGGAG CTCGCCCATC GCCCCTGCGC TGCTCGGACC	180
	GCGGCCACAG CCGGACTGGC GGGTACGGCG GCGACAGAGG CATTGGCCGA GAGTCCCAGT	240
20	CCGCAGAGTA GCCCCGGCCT CGAGGCGGTG GCGTCCCGGT CCTCTCCGTC CAGGAGCCAG	300
	GACAGGTGTC GCGCGGGGG GCTCCAGGGA CCGCGCCTGA GGCCGGCTGC CCGCCCGTCC	360
	CGCCCCGCCC CGCCCCCC CGCCCCCA GCCCAGCCTC CTTGCCGTCG GGGCGTCCCC	420
25	AGGCCCTGGG TCGGCCGCGG AGCCGATGCG CGCCCGCTGA GCGCCCCAGC TGAGCGCCCC	480
	CGGCCTGCC ATG ACC GCG CTC CCC GGC CCG CTC TGG CTC CTG GGC CTG	528
30	Met Thr Ala Leu Pro Gly Pro Leu Trp Leu Leu Gly Leu 1 5 10	
	GCG CTA TGC GCG CTG GGC GGG GGC CCC GGC CTG CGA CCC CCG CCC Ala Leu Cys Ala Leu Gly Gly Gly Pro Gly Leu Arg Pro Pro Pro 15 20 25	576
35	GGC TGT CCC CAG CGA CGT CTG GGC GCG CGC GAG CGC CGG GAC GTG CAG Gly Cys Pro Gln Arg Arg Leu Gly Ala Arg Glu Arg Arg Asp Val Gln 30 35 40 40	624
40	CGC GAG ATC CTG GCG GTG CTC GGG CTG CCT GGG CGG C	672
45	GCG CCA CCC GCC GCC TCC CGG CTG CCC GCG TCC GCG CCG C	720
50	CTG GAC CTG TAC CAC GCC ATG GCC GGC GAC GAC GAC GAC GAC GGC GCG Leu Asp Leu Tyr His Ala Met Ala Gly Asp Asp Glu Asp Gly Ala 80 85 90	768

	CCC Pro	GCG Ala 95	GAG Glu	CGG Arg	CGC	CTG Leu	GGC Gly 100	CGC Arg	GCC Ala	GAC Asp	CTG Leu	GTC Val 105	ATG Het	AGC Ser	TTC Phe	GTT Val	816
5	110	Het	Val	Glu	Arg	115	Arg	Ala	ren	GIY	120	GIII		•		125	864
10	AAG Lys	GAG Glu	TTC Phe	CGC Arg	TTT Phe 130	GAC Asp	CTG Leu	ACC Thr	CAG Gln	ATC Ile 135	CCG Pro	GCT Ala	GGG Gly	GAG Glu	GCG Ala 140	GTC Val	912
15	Thr	Ala	Ala	Glu 145	Phe	Arg	IIe	lyr	150	vai	rio	Ser		155			960
	Asn	Arg	Thr 160	Leu	His	Val	Ser	165	rne	GIII	Val	101	170	020			1008
20	AAC Asn	AGG Arg 175	GAG Glu	TCT Ser	GAC Asp	TTG Leu	TTC Phe 180	TTT Phe	TTG Leu	GAT Asp	CTT Leu	CAG Gln 185	ACG Thr	CTC Leu	CGA Arg	GCT Ala	1056
25	190	Asp	Glu	Gly	Trp	Leu 195	Val	ren	ASP	VAI	200	VIG	NTO	501		205	1104
30	Trp	Leu	Leu	Lys	Arg 210	His	Lys	Asp	ren	215	ren	urg	Deu	-,-	GTG Val 220		1152
35	ACT Thr	GAG Glu	GAC Asp	GGG Gly 225	CAC His	AGC Ser	GTG Val	GAT Asp	CCT Pro 230	GIA	CTG Leu	GCC Ala	GGC Gly	CTG Leu 235		GGT	1200
	Gln	Arg	Ala 240	Pro	Arg	Ser	Gin	245	PTO	rne	Val	141	250				1248
40	GCC Ala	AGT Ser 255	CCG Pro	AGT Ser	CCC Pro	ATC Ile	CGC Arg 260	ACC Thr	CCT Pro	CGG Arg	GCA Ala	GTG Val 265	AGG Arg	CCA Pro	CTG Leu	AGG Arg	1296
45	AGG Arg 270	AGG Arg	CAG Gln	CCG Pro	AAG Lys	AAA Lys 275	AGC Ser	AAC Asn	GAG Glu	CTG Leu	CCG Pro 280	CAG Gln	GCC Ala	AAC Asn	CGA	CTC Leu 285	1344
50	CCA Pro	GGG Gly	ATC Ile	TTT Phe	GAT Asp 290	GAC Asp	GTC Val	CAC His	GGC Gly	TCC Ser 295	CAC His	GGC Gly	CGG Arg	CAG Gln	GTC Val 300	TGC Cys	1392

	CGT Arg	CGG Arg	CAC His	GAG Glu 305	CTC Leu	TAC Tyr	GTC Val	AGC Ser	TTC Phe 310	CAG Gln	GAC Asp	CTC Leu	GGC Gly	TGG Trp 315	CTG Leu	GAC Asp	1440
5	TGG Trp	GTC Val	ATC Ile 320	GCT Ala	CCC Pro	CAA Gln	GGC Gly	TAC Tyr 325	TCG Ser	GCC Ala	TAT Tyr	TAC Tyr	TGT Cys 330	GAG Glu	GGG Gly	GAG Glu	1488
10	TGC Cys	TCC Ser 335	TTC Phe	CCA Pro	CTG Leu	GAC Asp	TCC Ser 340	TGC Cys	ATG Met	AAT Asn	GCC Ala	ACC Thr 345	AAC Asn	CAC His	GCC Ala	ATC Ile	1536
15	CTG Leu 350	CAG Gln	TCC Ser	CTG Leu	GTG Val	CAC His 355	CTG Leu	ATG Met	AAG Lys	CCA Pro	AAC Asn 360	GCA Ala	GTC Val	CCC Pro	AAG Lys	GCG Ala 365	1584
20	TGC Cys	TGT Cys	GCA Ala	CCC Pro	ACC Thr 370	AAG Lys	CTG Leu	AGC Ser	GCC Ala	ACC Thr 375	TCT Ser	GTG Val	CTC Leu	TAC Tyr	TAT Tyr 380	GAC Asp	1632
20	AGC Ser	AGC Ser	AAC Asn	AAC Asn 385	GTC Val	ATC Ile	CTG Leu	CGC Arg	AAA Lys 390	CAC His	CGC Arg	AAC Asn	ATG Met	GTG Val 395	GTC Val	AAG Lys	1680
25			GGC Gly 400			T GA	GTCA		GCC	CCAGO	CCT	ACTO	GCAG				1723
30	(2)		FORMA SEQU	ENCE						_							
35		(ii	(A) (B) (D)	TY	PE:	I: 40 amin )GY: 'PE:	o ac line	id ar	acid	ls							
40			(A) (SEQ	OTHE	R IN	FORM SCRI							P"				
	Met 1	•	Ala										Leu	Ala	Leu 15	Cys	
45	Ala	Leu	Gly	Gly 20	Gly	Gly	Pro	Gly	Leu 25	Arg	Pro	Pro	Pro	Gly 30	Cys	Pro	
50	Gln	Arg	Arg 35	Leu	Gly	Ala	Arg	Glu 40	Arg	Arg	Asp	Val	Gln 45	Arg	Glu	Ile	

	Leu	Ala 50	Val	Leu	Gly	Leu	Pro 55	Gly	Arg	Pro	Arg	Pro 60	Arg	Ala	Pro	Pro
5	Ala 65		Ser	Arg	Leu	Pro 70	Ala	Ser	Ala	Pro	Leu 75	Phe	Met	Leu	Asp	Leu 80
	Tyr	His	Ala	Het	Ala 85	Gly	Asp	Asp	Asp	Glu 90	Asp	Gly	Ala	Pro	Ala 95	Glu
10	Arg	Arg	Leu	Gly 100	Arg	Ala	Asp	Leu	Val 105	Het	Ser	Phe	Val	Asn 110	Met	Val
	Glu	Arg	Asp 115	Arg	Åla	Leu	Gly	His 120	Gln	Glu	Pro	His	Trp 125	Lys	Glu	Phe
15	Arg	Phe 130	Asp	Leu	Thr	Gln	Ile 135	Pro	Ala	Gly	Glu	Ala 140	Val	Thr	Ala	Ala
20	Glu 145	Phe	Arg	Ile	Tyr	Lys 150	Val	Pro	Ser	Ile	His 155	Leu	Leu	Asn	Arg	Thr 160
	Leu	His	Val	Ser	Met 165	Phe	Gln	Val	Val	Gln 170	Glu	Gln	Ser	Asn	Arg 175	Glu
25	Ser	Asp	Leu	Phe 180	Phe	Leu	Asp	Leu	Gln 185	Thr	Leu	Arg	Ala	Gly 190	Asp	Glu
	Gly	Trp	Leu 195	Val	Leu	Asp	Val	Thr 200	Ala	Ala	Ser	Asp	Cys 205	Trp	Leu	Leu
30	Lys	Arg 210	His	Lys	Asp	Leu	Gly 215	Leu	Arg	Leu	Tyr	Val 220	Glu	Thr	Glu	Asp
35	Gly 225	His	Ser	Val	Asp	Pro 230	Gly	Leu	Ala	Gly	Leu 235	Leu	Gly	Gln	Arg	Ala 240
	Pro	Arg	Ser	Gln	Gln 245	Pro	Phe	Val	Val	Thr 250	Phe	Phe	Arg	Ala	Ser 255	Pro
40	Ser	Pro	Ile	Arg 260	Thr	Pro	Arg	Ala	Val 265	Arg	Pro	Leu	Arg	Arg 270	Arg	Gln
	Pro	Lys	Lys 275	Ser	Asn	Glu	Leu	Pro 280	Gln	Ala	Asn	Arg	Leu 285	Pro	Gly	Ile
45	Phe	Asp 290	Asp	Val	His	Gly	Ser 295	His	Gly	Arg	Gln	Val 300	Cys	Arg	Arg	His
50	Glu 305	Leu	Tyr	Val	Ser	Phe 310	Gln	Asp	Leu	Gly	Trp 315	Leu	Asp	Trp	Val	Ile 320

	Ala	Pro	Gln	Gly	Tyr 325	Ser	Ala	Tyr	Tyr	Cys 330	Glu	Gly	Glu	Cys	Ser 335	Phe	
5	Pro	Leu	Asp	Ser 340		Het	Asn	Ala	Thr 345	Asn	His	Ala	Ile	Leu 350	Gln	Ser	
	Leu	Val	His 355	Leu	Het	Lys	Pro	Asn 360	Ala	Val	Pro	Lys	Ala 365	Cys	Cys	Ala	
10	Pro	Thr 370	Lys	Leu	Ser	Ala	Thr 375	Ser	Val	Leu	Tyr	Tyr 380	Asp	Ser	Ser	Asn	
15	Asn 385	Val	Ile	Leu	Arg	Lys 390	His	Arg	Asn	Het	Val 395	Val	Lys	Ala	Cys	Gly 400	
13	Cys	His													•		
	(2)	INI	FORMA	OITA	v FOI	R SEC	Q ID	NO:2	22:								
20			(i)	( <i>I</i>	A) I	ENGT	CHARA	1926 :leic	base aci	e pai id	irs						
25			( <b>ii</b> )	(I	ו (	COPOI	IDEDN LOGY : TYPE :	lir	ear	igte							
30			(vi)		RIGIN	IAL S	OURC IISM: IE TY	E:	RIDAE								
35			(ix)	(A	3) L	AHE/	KEY: ION:	93.	.128		ote=	: "mC	)P2 c	DNA"		:	
			(xi)	SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	10:22	:				
10			GCCA														50
	CCGA	CCAG	CT A	.CCAG	TGGA	T GC	GCGC	CGGC	TGA	AAGT	CCG		TG G et A				104
15	CCC Pro	GGG Gly	CCA Pro	CTC Leu	TGG Trp	CTA Leu 10	TTG Leu	GGC Gly	CTT Leu	GCT Ala	CTG Leu 15	TGC Cys	GCG Ala	CTG Leu	GGA Gly	GGC Gly 20	152
50	GGC (	CAC His	GGT Gly	CCG Pro	CGT Arg	CCC Pro	CCG Pro	CAC His	ACC Thr	TGT Cys	CCC Pro	CAG Gln	CGT Arg	CGC Arg	CTG Leu	GGA Gly	200

	GCG Ala	CGC Arg	GAG Glu	CGC Arg 40	CGC Arg	GAC Asp	ATG Het	CAG Gln	CGT Arg 45	GAA Glu	ATC Ile	CTG Leu	GCG Ala	GTG Val 50	CTC	GGG Gly	248
5	CTA Leu	CCG Pro	GGA Gly 55	CGG Arg	CCC Pro	CGA Arg	CCC Pro	CGT Arg 60	GCA Ala	CAA Gln	CCC Pro	GCG Ala	GCT Ala 65	GCC Ala	CGG Arg	CAG Gln	296
10	CCA Pro	GCG Ala 70	TCC Ser	GCG Ala	CCC Pro	CTC Leu	TTC Phe 75	ATG Met	TTG Leu	GAC Asp	CTA Leu	TAC Tyr 80	CAC His	GCC Ala	ATG Het	ACC Thr	344
15	GAT Asp 85	GAC Asp	GAC Asp	GAC Asp	GGC Gly	GGG Gly 90	CCA Pro	CCA Pro	CAG Gln	GCT Ala	CAC His 95	TTA Leu	GGC Gly	CGT Arg	GCC Ala	GAC Asp 100	392
	CTG Leu	GTC Val	ATG Met	AGC Ser	TTC Phe 105	GTC Val	AAC Asn	ATG Het	GTG Val	GAA Glu 110	CGC Arg	GAC Asp	CGT Arg	ACC Thr	CTG Leu 115	GGC Gly	440
20	TAC Tyr	CAG Gln	GAG Glu	CCA Pro 120	CAC His	TGG Trp	AAG Lys	GAA Glu	TTC Phe 125	CAC His	TTT Phe	GAC Asp	CTA Leu	ACC Thr 130	CAG Gln	ATC Ile	488
25	CCT Pro	GCT Ala	GGG Gly 135	GAG Glu	GCT Ala	GTC Val	ACA Thr	GCT Ala 140	GCT Ala	GAG Glu	TTC Phe	CGG Arg	ATC Ile 145	TAC Tyr	AAA Lys	GAA Glu	536
30	CCC Pro	AGC Ser 150	ACC Thr	CAC His	CCG Pro	CTC Leu	AAC Asn 155	ACA Thr	ACC Thr	CTC Leu	CAC His	ATC Ile 160	AGC Ser	ATG Het	TTC Phe	GAA Glu	584
35	GTG Val 165	GTC Val	CAA Gln	GAG Glu	CAC His	TCC Ser 170	AAC Asn	AGG Arg	GAG Glu	TCT Ser	GAC Asp 175	TTG Leu	TTC Phe	TTT Phe	TTG Leu	GAT Asp 180	632
	CTT Leu	CAG Gln	ACG Thr	CTC Leu	CGA Arg 185	TCT Ser	GGG Gly	GAC Asp	GAG Glu	GGC Gly 190	TGG Trp	CTG Leu	GTG Val	CTG Leu	GAC Asp 195	ATC Ile	680
40	ACA Thr	GCA Ala	GCC Ala	AGT Ser 200	GAC Asp	CGA Arg	TGG Trp	CTG Leu	CTG Leu 205	AAC Asn	CAT His	CAC His	AAG Lys	GAC Asp 210	CTG Leu	GGA Gly	728
45	CTC Leu	Arg	CTC Leu 215	TAT Tyr	GTG Val	GAA Glu	ACC Thr	GCG Ala 220	GAT Asp	GGG Gly	CAC His	AGC Ser	ATG Het 225	GAT Asp	CCT Pro	GGC Gly	776
50	CTG Leu	GCT Ala 230	GGT Gly	CTG Leu	CTT Leu	Gly	CGA Arg 235	CAA Gln	GCA Ala	CCA Pro	CGC Arg	TCC Ser 240	AGA Arg	CAG Gln	CCT Pro	TTC Phe	824

	ATG Het 245	GTA Val	ACC Thr	TTC Phe	TTC Phe	AGG Arg 250	GCC Ala	AGC Ser	CAG Gln	AGT Ser	CCT Pro 255	GTG Val	CGG Arg	GCC Ala	CCT Pro	CGG Arg 260	872
5	GCA Ala	GCG Ala	AGA Arg	CCA Pro	CTG Leu 265	AAG Lys	AGG Arg	AGG Arg	CAG Gln	CCA Pro 270	AAG Lys	AAA Lys	ACG Thr	AAC Asn	GAG Glu 275	CTT Leu	920
10	CCG Pro	CAC His	CCC Pro	AAC Asn 280	AAA Lys	CTC Leu	CCA Pro	GGG Gly	ATC Ile 285	TTT Phe	GAT Asp	GAT Asp	GGC Gly	CAC His 290	GGT Gly	TCC Ser	968
15	CGC Arg	GGC Gly	AGA Arg 295	GAG Glu	GTT Val	TGC Cys	CGC Arg	AGG Arg 300	CAT His	GAG Glu	CTC Leu	TAC Tyr	GTC Val 305	AGC Ser	TTC Phe	CGT Arg	1016
20	GAC Asp	CTT Leu 310	GGC Gly	TGG Trp	CTG Leu	GAC Asp	TGG Trp 315	GTC Val	ATC Ile	GCC Ala	CCC Pro	CAG Gln 320	GGC Gly	TAC Tyr	TCT Ser	GCC Ala	1064
20	TAT Tyr 325	TAC Tyr	TGT Cys	GAG Glu	GGG Gly	GAG Glu 330	TGT Cys	GCT Ala	TTC Phe	CCA Pro	CTG Leu 335	GAC Asp	TCC Ser	TGT Cys	ATG Net	AAC Asn 340	1112
25	GCC Ala	ACC Thr	AAC Asn	CAT His	GCC Ala 345	ATC Ile	TTG Leu	CAG Gln	TCT Ser	CTG Leu 350	GTG Val	CAC His	CTG Leu	ATG Met	AAG Lys 355	CCA Pro	1160
30	GAT Asp	GTT Val	GTC Val	CCC Pro 360	AAG Lys	GCA Ala	TGC Cys	TGT Cys	GCA Ala 365	CCC Pro	ACC Thr	AAA Lys	CTG Leu	AGT Ser 370	GCC Ala	ACC Thr	1208
35	TCT Ser	GTG Val	CTG Leu 375	TAC Tyr	TAT Tyr	GAC Asp	AGC Ser	AGC Ser 380	AAC Asn	AAT Asn	GTC Val	ATC Ile	CTG Leu 385	CGT Arg	AAA Lys	CAC His	1256
	CGT Arg	AAC Asn 390	ATG Het	GTG Val	GTC Val	AAG Lys	GCC Ala 395	TGT Cys	GGC Gly	TGC Cys	CAC His	TGAC	GCCC	CCG (	CCAC	GCATCO	1309
40	TGCI	TCTA	CT A	CCTI	CACCA	T CI	rggco	GGGC	ccc	CTCTC	CAG	AGGC	AGA	AC (	CCTT	TATGI	1369
	TATO	ATAC	CT C	CAGAC	CAGGO	G CA	ATG	GAGO	CCC	TTCA	CTT	ccc	TGG	CCA (	CTTCC	CTGCTA	1429
45	TAAA	TCT	GT C	TTTC	CCAG	T TO	CTCI	GTC	TTO	CATGO	GGT	TTC	GGGG	TA 1	CACO	CCGCC	1489
	CTCI	CCAT	CC I	CCTA	rccc	CA AC	CATA	GACI	GAA	TGCA	CAC	AGCA	TCC	CAG A	AGCTA	LTGCTA	1549
<b></b>	ACTG	AGAG	GT C	TGGG	GTCA	LG CA	CTGA	AGGC	CCA	CATO	AGG	AAGA	CTGA	ATC (	CTTGC	CCATO	1609
50	CTCA	GCCC	AC A	ATGO	CAAA	T TC	TGGA	TGGI	CTA	AGAA	.GGC	CGTG	GAAT	TC 1	) AAA	TAGAT	1669

	THE PROPERTY OF THE PROPERTY O	1729
	GATCTGGGCT CTCTGCACCA TTCATTGTGG CAGTTGGGAC ATTTTTAGGT ATAACAGACA	1789
	CATACACTTA GATCAATGCA TCGCTGTACT CCTTGAAATC AGAGCTAGCT TGTTAGAAAA	
5	AGAATCAGAG CCAGGTATAG CGGTGCATGT CATTAATCCC AGCGCTAAAG AGACAGAGAC	1849
	AGGAGAATCT CTGTGAGTTC AAGGCCACAT AGAAAGAGCC TGTCTCGGGA GCAGGAAAAA	1909
	AAAAAAAAC GGAATTC	1926
10	(2) INFORMATION FOR SEQ ID NO:23:	
15	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 399 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear	
20	<pre>(ii) MOLECULE TYPE: protein (ix) FEATURE:     (D) OTHER INFORMATION: /product= "mOP2-PP"</pre>	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
25	Het Ala Het Arg Pro Gly Pro Leu Trp Leu Leu Gly Leu Ala Leu Cys 1 10 15	
	Ala Leu Gly Gly Gly His Gly Pro Arg Pro Pro His Thr Cys Pro Gln 20 25 30	
30	Arg Arg Leu Gly Ala Arg Glu Arg Arg Asp Met Gln Arg Glu Ile Leu Ala 35 40	
35	Val Leu Gly Leu Pro Gly Arg Pro Arg Pro Arg Ala Gln Pro Ala Ala 50 55 60 65	
	Ala Arg Gln Pro Ala Ser Ala Pro Leu Phe Met Leu Asp Leu Tyr His Ala 70 75 80	
40	Het Thr Asp Asp Asp Gly Gly Pro Pro Gln Ala His Leu Gly Arg 85 90 95	
	Ala Asp Leu Val Met Ser Phe Val Asn Het Val Glu Arg Asp Arg Thr 100 105 110	
45	Leu Gly Tyr Gln Glu Pro His Trp Lys Glu Phe His Phe Asp Leu Thr 115 120 130	
50	Gln Ile Pro Ala Gly Glu Ala Val Thr Ala Ala Glu Phe Arg Ile Tyr 135 140 145	

	Lys	Glu	Pro	Ser 150	Thr	His	Pro	Leu	Asn 155	Thr	Thr	Leu	His	Ile 160	Ser	Het
5	Phe	Glu	Val 165	Val	Gln	Glu	His	Ser 170		Arg	Glu	Ser	Asp 175	Leu	Phe	Phe
	Leu	Asp 180	Leu	Gln	Thr	Leu	Arg 185	Ser	Gly	Asp	Glu	Gly 190	Trp	Leu	Val	Leu
10	Asp 195	Ile	Thr	Ala	Ala	Ser 200	Asp	Arg	Trp	Leu	Leu 205	Asn	His	His	Lys	Asp 210
	Leu	Gly	Leu	Arg	Leu 215	Tyr	Val	Glu	Thr	Ala 220	Asp	Gly	His	Ser	<b>Met</b> 225	Asp
15	Pro	Gly	Leu	Ala 230	Gly	Leu	Leu	Gly	Arg 235	Gln	Ala	Pro	Arg	Ser 240	Arg	Gln
20	Pro	Phe	Met 245	Val	Thr	Phe	Phe	Arg 250	Ala	Ser	Gln	Ser	Pro 255	Val	Arg	Ala
	Pro	Arg 260	Ala	Ala	Arg	Pro	Leu 265	Lys	Arg	Arg	Gln	Pro 270	Lys	Lys	Thr	Asn
25	Glu 275	Leu	Pro	His	Pro	Asn 280	Lys	Leu	Pro	Gly	Ile 285	Phe	Asp	Asp	Gly	His 290
20	Gly	Ser	Arg	Gly	Arg 295	Glu	Val	Cys	Arg	Arg 300	His	Glu	Leu	Tyr	Val 305	Ser
30	Phe	Arg	Asp	Leu 310	Gly	Trp	Leu	Asp	Trp 315	Val	Ile	Ala	Pro	Gln 320	Gly	Tyr
35	Ser	Ala	Tyr 325	Tyr	Cys	Glu	Gly	Glu 330	Cys	Ala	Phe	Pro	Leu 335	Asp	Ser	Cys
	Het	Asn 340	Ala	Thr	Asn	His	Ala 345	Ile	Leu	Gln	Ser	Leu 350	Val	His	Leu	<b>Met</b>
40	Lys 355	Pro	Asp	Val	Val	Pro 360	Lys	Ala	Cys	Cys	Ala 365	Pro	Thr	Lys	Leu	Ser 370
45	Ala	Thr	Ser	Val	Leu 375	Tyr	Tyr	Asp	Ser	Ser 380	Asn	Asn	Val	Ile	Leu 385	Arg
45	Lys	His	Arg	Asn 390	Het	Val	Val	Lys	Ala 395	Cys	Gly	Cys	His			

	(2) INFORMATION FOR SEQ ID NO:24:		
5	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 1368 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	·	
10	(ii) HOLECULE TYPE: cDNA		
10	(ix) FEATURE:  (A) NAME/KEY: CDS  (B) LOCATION: 11368  (D) OTHER INFORMATION:/STANDARD NAME="60A"		
15	(x) PUBLICATION INFORMATION:  (A) AUTHORS: WHARTON, KRISTI A.; THOMSEN,  GELBERT, WILLIAM M.  (R) TITLE: DROSOPHILA 60A GENE	GERALD H.;	
20	TOTALLE DECC MATTER ACAD SULLIDA	1 TO 1368	
25	· ·		
30	ATG TCG GGA CTG CGA AAC ACC TCG GAG GCC GTT GCA GTG CTC G Het Ser Gly Leu Arg Asn Thr Ser Glu Ala Val Ala Val Leu A 1 5 10	CC TCC la Ser 15	48
	CTG GGA CTC GGA ATG GTT CTG CTC ATG TTC GTG GCG ACC ACG C Leu Gly Leu Gly Met Val Leu Leu Met Phe Val Ala Thr Thr P 20 25 30	CG CCG To Pro	96
35	GCC GTT GAG GCC ACC CAG TCG GGG ATT TAC ATA GAC AAC GGC A Ala Val Glu Ala Thr Gln Ser Gly Ile Tyr Ile Asp Asn Gly I 35 40 45	AG GAC ys Asp	144
40	CAG ACG ATC ATG CAC AGA GTG CTG AGC GAG GAC GAC AAG CTG GGIn Thr Ile Met His Arg Val Leu Ser Glu Asp Asp Lys Leu A 50 60	AC GTC sp Val	192
45	65 70 /3	80	240
50	CTG AGC AGC CAC CAG TTG TCG CTG AGG AAG TCG GCT CCC AAG T Leu Ser Ser His Gln Leu Ser Leu Arg Lys Ser Ala Pro Lys P 85 90	TC CTG he Leu 95	288

	CTG Leu	GAC Asp	GTC Val	TAC Tyr 100	His	CGC Arg	ATC Ile	ACG Thr	GCG Ala 105	GAG Glu	GAG Glu	GGT Gly	CTC Leu	AGC Ser 110	GAT Asp	CAG Gln		336
5	GAT Asp	GAG Glu	GAC Asp 115	GAC Asp	GAC Asp	TAC Tyr	GAA Glu	CGC Arg 120	GGC Gly	CAT His	CGG Arg	TCC Ser	AGG Arg 125	AGG Arg	AGC Ser	GCC Ala		384
10	GAC Asp	CTC Leu 130	GAG Glu	GAG Glu	GAT Asp	GAG Glu	GGC Gly 135	GAG Glu	CAG Gln	CAG Gln	AAG Lys	AAC Asn 140	TTC Phe	ATC Ile	ACC Thr	GAC Asp		432
15	CTG Leu 145	Asp	AAG Lys	CGG Arg	GCC Ala	ATC Ile 150	GAC Asp	GAG Glu	AGC Ser	GAC Asp	ATC Ile 155	ATC Ile	ATG Net	ACC Thr	TTC Phe	CTG Leu 160		480
20	AAC Asn	AAG Lys	CGC Arg	CAC His	CAC His 165	AAT Asn	GTG Val	GAC Asp	GAA Glu	CTG Leu 170	CGT Arg	CAC His	GAG Glu	CAC His	GGC Gly 175	CGT Arg	v	528
20	CGC Arg	CTG Leu	TGG Trp	TTC Phe 180	GAC Asp	GTC Val	TCC Ser	AAC Asn	GTG Val 185	CCC Pro	AAC Asn	GAC Asp	AAC Asn	TAC Tyr 190	CTG Leu	GTG Val		576
25												GAG Glu				CTG Leu		624
30	ACC Thr	GCC Ala 210	AAC Asn	AGG Arg	GAG Glu	TTC Phe	ACC Thr 215	ATC Ile	ACG Thr	GTA Val	TAC Tyr	GCC Ala 220	ATT Ile	GGC Gly	ACC Thr	GGC Gly		672
35	ACG Thr 225	CTG Leu	GGC Gly	CAG Gln	CAC His	ACC Thr 230	ATG Met	GAG Glu	CCG Pro	CTG Leu	TCC Ser 235	TCG Ser	GTG Val	AAC Asn	ACC Thr	ACC Thr: 240		720
40	GGG Gly	GAC Asp	TAC Tyr	GTG Val	GGC Gly 245	TGG Trp	TTG Leu	GAG Glu	CTC Leu	AAC Asn 250	GTG Val	ACC Thr	GAG Glu	GGC Gly	CTG Leu 255	CAC His		768
40	GAG Glu	TGG Trp	CTG Leu	GTC Val 260	AAG Lys	TCG Ser	AAG Lys	GAC Asp	AAT Asn 265	CAT His	GGC Gly	ATC Ile	TAC Tyr	ATT Ile 270	GGA Gly	GCA Ala		816
45	CAC His	GCT Ala	GTC Val 275	AAC Asn	CGA Arg	CCC Pro	GAC Asp	CGC Arg 280	GAG Glu	GTG Val	AAG Lys	CTG Leu	GAC Asp 285	Asp	ATT Ile	GGA Gly		864
50	CTG Leu	ATC Ile 290	CAC His	CGC Arg	AAG Lys	GTG Val	GAC Asp 295	GAC Asp	GAG Glu	TTC Phe	CAG Gln	CCC Pro 300	TTC Phe	ATG Het	ATC Ile	GGC Gly		912

	TTC Phe 305	TTC Phe	CGC	GGA Gly	CCG Pro	GAG Glu 310	CTG Leu	ATC Ile	AAG Lys	GCG Ala	ACG Thr 315	GCC Ala	CAC His	AGC Ser	AGC Ser	CAC His 320	960	0
5	CAC	AGG Arg	AGC Ser	AAG Lys	CGA Arg 325	AGC Ser	GCC Ala	AGC Ser	CAT His	CCA Pro 330	CGC Arg	AAG Lys	CGC Arg	AAG Lys	AAG Lys 335	TCG Ser	100	В
10	GTG Val	TCG Ser	CCC Pro	AAC Asn 340	AAC Asn	GTG Val	CCG Pro	CTG Leu	CTG Leu 345	GIU	CCG Pro	ATG Met	GAG Glu	AGC Ser 350	ACG Thr	CGC Arg	1050	5
15	AGC Ser	TGC Cys	CAG Gln 355	ATG Net	CAG Gln	ACC Thr	CTG Leu	TAC Tyr 360	ATA Ile	GAC Asp	TTC Phe	AAG Lys	GAT Asp 365	CTG Leu	GGC Gly	TGG Trp	110	
	CAT His	GAC Asp 370	TGG Trp	ATC Ile	ATC Ile	GCA Ala	CCA Pro 375	GAG Glu	GGC Gly	TAT	GGC Gly	GCC Ala 380	TTC Phe	TAC Tyr	TGC Cys	AGC Ser	115	2
20	GGC Gly 385	GAG Glu	TGC Cys	AAT Asn	TTC Phe	CCG Pro 390	CTC Leu	AAT Asn	GCG Ala	CAC His	ATG Met 395	AAC Asn	GCC Ala	ACG Thr	AAC Asn	CAT His 400	120	0
25	GCG Ala	ATC Ile	GTC Val	CAG Gln	ACC Thr 405	CTG Leu	GTC Val	CAC His	CTG Leu	CTG Leu 410	GAG Glu	CCC Pro	AAG Lys	AAG Lys	GTG Val 415	CCC Pro	124	В
30	AAG Lys	CCC Pro	TGC Cys	TGC Cys 420	GCT Ala	CCG Pro	ACC Thr	AGG Arg	CTG Leu 425	GGA Gly	GCA Ala	CTA Leu	CCC	GTT Val 430	CTG Leu	TAC Tyr	129	6
35	CAC His	CTG Leu	AAC Asn 435	GAC Asp	GAG Glu	AAT Asn	GTG Val	AAC Asn 440	CTG Leu	AAA Lys	AAG Lys	TAT Tyr	AGA Arg 445	AAC Asn	ATG Het	ATT Ile	134	4
	GTG Val	AAA Lys 450	TCC Ser	TGC Cys	GGG Gly	TGC Cys	CAT His 455	TGA									136	8
40	(2)	IN	FORM	ATIO	N FO	R SE	Q ID	NO:	25:									
45			(i	, (,	EQUE A) L B) T D) T	ENGT: YPE:	H: 4 ami:	55 a no a	mino cid	ICS: aci	ds							
			(ii	•	OLEC											-		
50			(xi	) S	EQUE	NCE	DESC	RIPT	ION:	SEQ	ID	NO:2	5:					

	Het 1	Ser	Gly	Leu	Arg 5	Asn	Thr	Ser	Glu	Ala 10	Val	Ala	Val	ren	15	Ser
5	Leu	Gly	Leu	Gly 20	Net	Val	Leu	Leu	Het 25	Phe	Val	Ala	Thr	Thr 30	Pro	Pro
	Ala	Val	Glu 35	Ala	Thr	Gln	Ser	Gly 40	Ile	Tyr	Ile	Asp	Asn 45	Gly	Lys	Asp
10	Gln	Thr 50	Ile	Het	His	Arg	Val 55	Leu	Ser	Glu	Asp	Asp 60	Lys	Leu	Asp	Val
	Ser 65	Tyr	Glu	Ile	Leu	Glu 70	Phe	Leu	Gly	Ile	Ala 75	Glu	Arg	Pro	Thr	His 80
15	Leu	Ser	Ser	His	Gln 85	Leu	Ser	Leu	Arg	Lys 90	Ser	Ala	Pro	Lys	Phe 95	Leu
20	Leu	Asp	Val	Tyr 100	His	Arg	Ile	Thr	Ala 105	Glu	Glu	Gly	Leu	Ser 110	Asp	Gln
	Asp	Glu	Asp 115	Asp	Asp	Tyr	Glu	Arg 120	Gly	His	Arg	Ser	Arg 125	Arg	Ser	Ala
25	Asp	Leu 130	Glu	Glu	Asp	Glu	Gly 135	Glu	Gln	Gln	Lys	Asn 140	Phe	Ile	Thr	Asp
	Leu 145	Asp	Lys	Arg	Ala	Ile 150	Asp	Glu	Ser	Asp	Ile 155	Ile	Het	Thr	Phe	Leu 160
30	Asn	Lys	Arg	His	His 165	Asn	Val	Asp	Glu	Leu 170	Arg	His	Glu	His	Gly 175	Arg
35	Arg	Leu	Trp	Phe 180	Asp	Val	Ser	Asn	Val 185	Pro	Asn	Asp	Asn	Tyr 190	Leu	Val
	Het	Ala	Glu 195	Leu	Arg	Ile	Tyr	Gln 200	Asn	Ala	Asn	Glu	Gly 205	Lys	Trp	Leu
40	Thr	Ala 210	Asn	Arg	Glu	Phe	Thr 215	Ile	Thr	Val	Tyr	Ala 220	Ile	Gly	Thr	Gly
	Thr 225	Leu	Gly	Gln	His	Thr 230	Het	Glu	Pro	Leu	Ser 235	Ser	Val	Asn	Thr	Thr 240
45	Gly	Asp	Tyr	Val	Gly 245	Trp	Leu	Glu	Leu	Asn 250	Val	Thr	Glu	Gly	Leu 255	His
50	Glu	Trp	Leu	Val 260	Lys	Ser	Lys	Asp	Asn 265	His	Gly	Ile	Tyr	Ile 270	Gly	Ala

	His	Ala	Val 275	Asn	Arg	Pro	Asp	Arg 280	Glu	Val	Lys	Leu	Asp 285	Asp	Ile	Gly
5		290					293					200			Ile	
	305					310					313				Ser	
10					325					330					Lys 335	
				340					343	-				330	Thr	
15			355					360					303		Gly	
20	His	Asp 370	Trp	Ile	Ile	Ala	Pro 375	Glu	Gly	Tyr	Gly	Ala 380	Phe	Tyr	Cys	Ser
	Gly 385	Glu	Cys	Asn	Phe	Pro 390	Leu	Asn	Ala	His	Met 395	Asn	Ala	Thr	Asn	His 400
25					405					410					Val 415	
				420				•	427						Leu	
30	His	Leu	Asn 435	Asp	Glu	Asn	Val	Asn 440	Leu	Lys	Lys	Tyr	Arg 445	Asn	Het	Ile
35	Val	Lys 450	Ser	Cys	Gly	Cys	His 455									
	(2)	INF	ORMA!	rion	FOR	SEQ	ID 1	NO:2	6:							
40			(i)	, (1	A) LI B) T	ENGTI	H: an	ACTE mino no a lin	RIST: aci cid ear	ICS: ds						
			(ii	) <b>H</b> (	OLEC	ULE :	TYPE	: pr	otei	n						
45			(iii	) OI	RIGII A) O	NAL ( RGAN)	SOUR ISM:	CE: Hom	o Sa	pien	s		4			
50			(ix	, (	n's Ti	AHE/	TAN.	1 .	tein 102 TION		ote=	"BHP	3"			

	tx)	L) SEQU	JENCE	DESC	RIPT	CION	: SE	Q II	NO:	26:					
5	(i)SEC (A (E (C (I	S) TYPI	STH:	104 mino NESS:	amin acid	o a l .ngl		i							
10	(ii)MC	OLECULE	TYPE:	pr	otei	.n									
		3) LOCA	E/KEY: ATION: ER INE	: 1.	otei .104 TION	•	/not	:e="E	MP3"	,				!	
15	(xi)SE	EQUENCE	DESCE	RIPTI	ON:	SE	Q II	NO:	26:						
	Cys A	Ala Arg	Arg 1	Tyr L	eu I	ys	Val	Asp	Phe 10	Ala	Asp	Ile	Gly	Trp 15	Ser
20	Glu I	Trp Ile	Ile S 20	Ser P	ro L	ys	Ser	Phe 25	Asp	Ala	Tyr	Try	Cys 30	Ser	Gly
25	Ala (	Cys Gln 35	Phe I	Pro M	et P	ro	Lys 40	Ser	Leu	Lys	Pro	Ser 45	Asn	His	Ala
		lle Gln 50	Ser 1	Ile V		la 55	Arg	Ala	Val	Gly	Val 60	Val	Pro	Gly	Ile
30	Pro 0 65	Glu Pro	Cys (		al P O	Pro	Glu	Lys	Het	Ser 75	Ser	Leu	Ser	Ile	Leu 80
35	Phe F	Phe Asp		Asn L 85	ys A	Asn	Val	Val	Leu 90	Lys	Val	Tyr	Pro	Asn 95	Het
))	Thr V	/al Glu	Ser (	Cys A	.la C	Cys	Arg							ι	
10	(2) INFORMA	ATION FO	OR SEC	Q ID	NO:2	27:									
15	i)	(A) (B) (C)	JENCE LENGT TYPE: STRAN TOPOI	TH: 1 : ami NDEDN	02 a no a ESS:	min cid si	o ac ngle	ids							
	(ii	i) Moli	ECULE	TYPE	: pr	ote	in								
50	(vi	i) ORI( (A)	GINAL ORGAN	SOUR	CE:	10 S	APIE	ENS							

			(ix)	(1	EATUI A) NA B) L( C) O	ME/I	TON:	1	[02	: /no	ote=	"BMI	?5"			
5			(xi)	SI	EQUE	ICE I	ESCI	RIPT	ON:	SEQ	ID	NO:27	7:			
	Cys 1	Lys	Lys	His	Glu 5	Leu	Tyr	Val	Ser	Phe 10	Arg	Asp	Leu	Gly	Trp 15	Gln
10	Asp	Trp	Ile	Ile 20	Ala	Pro	Glu	Gly	Tyr 25	Ala	Ala	Phe	Tyr	Cys 30	Asp	Gly
15	Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Het	Asn	Ala	Thr 45	Asn	His	Ala
	Ile	Val 50	Gln	Thr	Leu	Val	His 55	Leu	Het	Phe	Pro	Asp 60	His	Val	Pro	Lys
20	Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Leu	Asn	Ala	Ile 75	Ser	Val	Leu	Tyr	Phe 80
	Asp	Asp	Ser	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Asn	Het	Val 95	Val
25	Arg	Ser	Cys	Gly 100	Cys	His										
30	(2)	INFO	ORMA:	\ S!	FOR EQUEI A) L	NCE (	CHAR	ACTE	RIST	ICS:	ds				-	
35				(1	B) T: C) S: D) T(	YPE: [RAN]	amiı DEDN	no a ESS:	cid sin		÷					
			(ii	) H	OLEC	ULE	TYPE	: pr	otei	n .						
40			(vi	) O:	RIGII A) O	NAL RGAN	SOUR ISH:	CE: HOM	O SA	PIEN	S					
45			(ix	( )	EATU A) N. B) L D) O	AHE/	TON:	1	102		ote=	"BM	P6"			
			(xi	•	EQUE											
	Cys	Arg	Lys	His	Glu	Leu	Tyr	Val	Ser	Phe 10	Gln	Asp	Leu	Gly	Trp 15	Glr

	Asp	Trp	Ile	Ile 20	Ala	Pro	Lys	Gly	Tyr 25	Ala	Ala	Asn	Tyr	Cys 30	Asp	GIÀ
5	Glu	Cys	Ser 35	Phe	Pro	Leu	Asn	Ala 40	His	Het	Asn	Ala	Thr 45	Asn	His	Ala
	Ile	Val 50	Gln	Thr	Leu	Val	His 55	Leu	Het	Asn	Pro	Glu 60	Tyr	Val	Pro	Lys
10	Pro 65	Cys	Cys	Ala	Pro	Thr 70	Lys	Leu	Asn	Ala	Ile 75	Ser	Val	Leu	Tyr	Phe 80
	Asp	Asp	Asn	Ser	Asn 85	Val	Ile	Leu	Lys	Lys 90	Tyr	Arg	Trp	Met	Val 95	<b>Val</b>
15	Arg	Ala	Cys	Gly 100	Cys	His						•		•		
20	(2)	INF				SEQ				TCC.						
			(i)	( / (1	A) LI 3) T	YPE:	H: 10 amin	02 ar no ar	mino cid	aci	is					
25			(ii)	•	-	JLE :				n						
30			(ix)	( / ( ]	B) L( D) O'	AME/I OCAT: THER	ION: INF( e= ")	1 ORMA' VHER	102 TION EIN :	: /1: XAA /	AT E	ACH I	POS '1	N IS	IND	EPENDENTLY
35					(	CORRI	ESPOI UMAN	NDIN OP1	G PO	S'N : OP2	IN TI	HE C	-TERI	MINA!	AT :	QUENCE OF HOUS 6,7 and 8 or
			(xi	) S	EQUE	NCE I	DESC	RIPT	ION:	SEQ	ID I	NO:2	9:			
40	Cys 1	Xaa	Xaa	His	Glu 5	Leu	Tyr	Val	Xaa	Phe 10	Xaa	Asp	Leu	Gly	Trp 15	Xaa
	Asp	Trp	Xaa	Ile 20	Ala	Pro	Xaa	Gly	Tyr 25	Xaa	Ala	Tyr	Туг	Cys 30	Glu	Gly
45	Glu	Cys	<b>Xaa</b> 35	Phe	Pro	Leu	Xaa	Ser 40	Xaa	Het	Asn	Ala	Thr 45	Asn	His	Ala
50	Ile	Xaa 50	Gln	Xaa	Leu	Val	His 55	Xaa	Xaa	Xaa	Pro	<b>Xaa</b> 60	Xaa	Val	Pro	Lys

20

```
Xaa Cys Cys Ala Pro Thr Xaa Leu Xaa Ala Xaa Ser Val Leu Tyr Xaa
                    70
Asp Xaa Ser Xaa Asn Val Xaa Leu Xaa Lys Xaa Arg Asn Het Val Val
```

Xaa Ala Cys Gly Cys His 100

## INFORMATION FOR SEQ ID NO:30: 10 (2)

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 97 amino acids
- (B) TYPE: amino acids
- TOPOLOGY: linear 15 (ii) MOLECULE TYPE: protein
  - (ix)FEATURE:

(A) NAME: Generic Sequence 5

(D) OTHER INFORMATION: wherein each Kaa is independently selected from a group of one or more specified amino acids as defined in the specification.

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Leu Xaa Xaa Yaa Phe 25 Xaa Xaa Xaa Gly Trp Xaa Xaa Trp Xaa 10

Xaa Xaa Pro Xaa Xaa Xaa Ala 20 30 15 Xaa Tyr Cys Xaa Gly Xaa Cys Xaa 30 25

Xaa Pro Xaa Xaa Xaa Xaa Xaa 35

Xaa Xaa Xaa Asn His Ala Xaa Xaa 35 45 40 Xaa Xaa Xaa Xaa Xaa Xaa Xaa

50 Xaa Xaa Xaa Xaa Xaa Xaa Cys

55 40 Cys Xaa Pro Xaa Xaa Xaa Xaa Xaa 65

Xaa Xaa Xaa Leu Xaa Xaa Xaa 70

Xaa Xaa Xaa Xaa Val Xaa Leu Xaa 45 80

Xaa Xaa Xaa Xaa Het Xaa Val Xaa

Xaa Cys Xaa Cys Xaa

•	(2) INFORM	LATION FOR SE	Q ID NO:3	1:	·	
5	(A (B (C (ii)MO	) TYPE: an	102 amino ino acids linear			
10	(A (D	) NAME: Ger O) OTHER INI selected	eric Seque CORMATION: from a gro n the spec	wherei oup of o	ne or more	is independently specified amino acids as
15	(xi) SE	QUENCE DESCR	RIPTION:	SEQ ID N	0:31:	T.
15	Cys Xaa	Xaa Xaa Xaa		Xaa Xaa	Phe 10	
	Xaa Xaa	Xaa Gly Tr	Xaa Xaa S	Trp Xaa		,
20	Xaa Xaa 20	Pro Xaa Xaa		Ala		
		Cys Xaa Gly	Xaa Cys	Xaa 35		
25	Xaa Pro	Xaa Xaa Xaa 40				
	Xaa Xaa	Xaa Asn His 45	Ala Xaa X	Xaa 50		
		Xaa Xaa Xaa 55	i			
30	60		65			·
	-	Pro Xaa Xaa 70		Xaa		
35	75	Xaa Leu Xaa	80	77		• •
		Xaa Xaa Val				
	90	Xaa Xaa Het	95	Aaa		
40	Xaa Cys	Xaa Cys Xaa 100	<b>L</b>			
	(2) INFO	RMATION FOR	SEQ ID NO	:32:		
45	(i) (A) (B) (C) (D)	SEQUENCE ( LENGTH: 12 TYPE: nucl STRANDEDNE TOPOLOGY:	38 base pa eic acid, SS: single	airs, 37 amino a	2 amino aci cid	ds

(ii) MOLECULE TYPE: cDNA

	(111 (A) (F)	ORG	IGINAL GANISH: SSUE TY	huma	an	I									
5	(iv) (A) (B) (D)	NAP LOC	ATURE: HE/KEY: CATION: HER INF	ORMA?	t= "G	:DF-1	n				-				
10			/no	te= '	"GDF-	-1 CI	NA"								
15	(x) (A) (B) (C) (D) (E) (F) (G)	AUT JOU VOI REI PAC	BLICATI THORS: TTLE: E URNAL: LUME: 8 LEVANT GES: 42 TE: Ma	Lee, xpres Proc 8 RESII 50-42	Se-J ssion Nat DUES: 254	Jin n of :'1 #	Grov cad.	SC1			iati:	lon F	acto	or 1	
20	(xi)	SEC	QUENCE	DESCI	RIPTI								20010	·me	60
	GGGGACACCG														
25	TCTGGTCATC	GCCT	GGGAGG	,	ATG ( Met I 1	CCA ( Pro I	CCG ( Pro I	ccc c	CAG ( Gln ( 5	CAA C	GT (	CCC I	rgc G Cys G	GC ly 10	113
30	CAC His	CAC (	CTC CTC Leu Leu	CTC Leu 15	CTC Leu	CTG Leu	GCC Ala	CTG Leu	CTG Leu 20	CTG Leu	CCC Pro	TCG Ser	CTG Leu	CCC Pro 25	158
	CTG Leu	ACC (	CGC GCC Arg Ala	CCC Pro 30	GTG Val	CCC Pro	CCA Pro	GGC Gly	CCA Pro 35	GCC Ala	GCC Ala	GCC Ala	CTG Leu	CTC Leu 40	203
35	CAG Gln	GCT (	CTA GG/ Leu Gly	CTG Leu 45	CGC Arg	GAT Asp	GAG Glu	CCC Pro	CAG Gln 50	GGT Gly	GCC Ala	CCC Pro	AGG Arg	CTC Leu 55	248
40	CGG Arg	CCG (	GTT CCC Val Pro	CCG Pro 60	GTC Val	ATG Het	TGG Trp	CGC Arg	CTG Leu 65	TTT Phe	CGA Arg	CGC Arg	CGG Arg	GAC Asp 70	293
45	CCC	CAG (	GAG AC( Glu Thi	AGG Arg 75	TCT Ser	GGC Gly	TCG Ser	CGG Arg	CGG Arg 80	ACG Thr	TCC Ser	CCA Pro	GGG Gly	GTC Val 85	338
50	ACC Thr	CTG (	CAA CCO Gln Pro	TGC Cyc 90	His	GTG Val	GAG Glu	GAG Glu	CTG Leu 95	GGG Gly	GTC Val	GCC Ala	GGA Gly	AAC Asn 100	383

	ATC Ile	GTG Val	CGC Arg	CAC His	ATC Ile 105	CCG Pro	GAC Asp	CGC Arg	GGT Gly	GCG Ala 110	CCC Pro	ACC Thr	CGG Arg	GCC Ala	TCG Ser 115	428
5	GAG Glu	CCT Pro	GTC Val	TCG Ser	GCC Ala 120	GCG Ala	GGG Gly	CAT His	TGC Cys	CCT Pro 125	GAG Glu	TGG Trp	ACA Thr	GTC Val	GTC Val 130	473
10	TTC Phe	GAC Asp	CTG Leu	TCG Ser	GCT Ala 135	GTG Val	GAA Glu	CCC Pro	GCT Ala	GAG Glu 140	CGC Arg	CCG Pro	AGC Ser	CGG Arg	GCC Ala 145	518
15	CGC Arg	CTG Leu	GAG Glu	CTG Leu	CGT Arg 150	TTC Phe	GCG Ala	GCG Ala	GCG Ala	GCG Ala 155	GCG Ala	GCA Ala	GCC Ala	CCG Pro	GAG Glu 160	563
	GGC Gly	GGC Gly	TGG Trp	GAG Glu	CTG Leu 165	AGC Ser	GTG Val	GCG Ala	CAA Gln	GCG Ala 170	GGC Gly	CAG Gln	GGC Gly	GCG Ala	GGC Gly 175	608
20	GCG Ala	GAC Asp	CCC Pro	GGG Gly	CCG Pro 180	GTG Val	CTG Leu	CTC Leu	CGC Arg	CAG Gln 185	TTG Leu	GTG Val	CCC Pro	GCC Ala	CTG Leu 190	653
25	GGG Gly	CCG Pro	CCA Pro	GTG Val	Arg	GCG Ala	GAG Glu	CTG Leu	CTG Leu	GGC Gly 200	GCC Ala	GCT Ala	TGG Trp	GCT Ala	CGC Arg	698
					195				000		ccc	CTC	ccc	СТА		743
30	AAC Asn	GCC Ala	TCA Ser	TGG	Pro 210	Arg	Ser	Leu	Arg	Leu 215	Ala	Leu	Ala	Leu	Arg 220	, 43
35	CCC Pro	CGG Arg	GCC Ala	CCT Pro	GCC Ala 225	GCC Ala	TGC Cys	GCG Ala	CGC Arg	CTG Leu 230	GCC Ala	GAG Glu	GCC Ala	TCG Ser	CTG Leu 235	788
40	CTG Leu	CTG Leu	GTG Val	ACC Thr	CTC Leu 240	GAC Asp	CCG Pro	CGC Arg	CTG Leu	TGC Cys 245	CAC His	CCC Pro	CTG Leu	GCC Ala	CGG Arg 250	833
	CCG Pro	CGG Arg	CGC Arg	GAC Asp	GCC Ala 255	GAA Glu	CCC Pro	GTG Val	TTG Leu	GGC Gly 260	GGC Gly	GGC Gly	CCC Pro	GGG Gly	GGC Gly 265	878
45	GCT Ala	TGT Cys	CGC Arg	GCG Ala	CGG Arg 270	CGG Arg	CTG Leu	TAC Tyr	GTG Val	AGC Ser 275	TTC Phe	CGC Arg	CAG Glu	GTG Val	GGC Gly 280	923
50	TGG Trp	CAC His	CGC Arg	TGG Trp	GTC Val 285	ATC Ile	GCG Arg	CCG Pro	CGC Arg	CCC Gly 290	TTC Phe	CTG Leu	GCC Ala	AAC Asn	TAC Tyr 295	968

		TGC ( Cys (	CAG Gln	GGT Gly	CAG Gln	TGC Cys 300	GCG Ala	CTG Leu	CCC Pro	GTC Val	GCG Ala 305	CTG Leu	TCG Ser	GGG Gly	TCC Ser	GGG Gly 310	1013
5		GGG (	CCG Pro	CCG Pro	GCG Ala	CTC Leu 315	AAC Asn	CAC His	GCT Ala	GTG Val	CTG Leu 320	CGC Arg	GCG Ala	CTC Leu	ATG Het	CAC His 325	1058
10		GCG (	GCC Ala	GCC Ala	CCG Pro	GGA Gly 330	GCC Ala	GCC Ala	GAC Asp	CTG Leu	CCC Pro 335	TGC Cys	TGC Cys	GTG Val	CCC Pro	GCG Ala 340	1103
15		CGC (	CTG Leu	TCG Ser	CCC Pro	ATC Ile 345	TCC Ser	GTG Val	CTC Leu	TTC Phe	TTT Phe 350	GAC Asp	AAC Asn	AGC Ser	GAC Asp	AAC Asn 355	1148
13		GTG (	GTG Val	CTG Leu	CGG Arg	CAG Gln 360	TAT Tyr	GAG Glu	GAC Asp	ATG Met	GTG Val 365	GTG Val	GAC Asp	GAG Glu	TGC Cys	GGC Gly 370	1193
20		Cys A		AAT	CCCG(	GGG (	CGGG(	CAGG	GA CO	CCGG(	GCCC	A AC	AATA	AATG	CCG	CGTGG	1238
25	(2)	INFO	RMA?	CION	FOR	SEQ	ID 1	NO:3	3:								
30		(i)	(1 (1	EQUEI A) B) C) D)	TYP	GTH: E: a: ANDE	372 mino DNES	RIST ami aci S: s inea	no a d ingl								
		(ii)	H	OLEC	ULE '	TYPE	: cD	NA							,		
35		(iii	) H	YPOT	HETI	CAL:	NO										
		(iv)	Al	NTI-	SENS	E: N	0										
40		(vi)	(4	RIGI A) F)	NAL ORG TIS	ANIS	H: h	uman : BR	AIN								
45		(ix)	(, ()	EATU A) B) (D)	NAM	E/KE ATIO ER I	N: NFOR	DS MATI ct=	ON: "GDF	/fun '-1"	ctio	n=					
50		(xi	.) S	EQUE	NCE	DESC	RIPT								•		
		-											Gln	Gly	Pro	Cys C	31 <b>y</b> 10

	His	His	Leu	Leu	Leu 15	Leu	Leu	Ala	Leu	Leu 20	Leu	Pro	Ser	Leu	Pro 25
5	Leu	Thr	Arg	Ala	Pro 30	Val	Pro	Pro	Gly	Pro 35	Ala	Ala	Ala	Leu	Leu 40
	Gln	Ala	Leu	Gly	Leu 45	Arg	Asp	Glu	Pro	Gln 50	Gly	Ala	Pro	Arg.	Leu 55
10	Arg	Pro	Val	Pro	Pro 60	Val	Met	Trp	Arg	Leu 65	Phe	Arg	Arg	Arg	Asp 70
	Pro	Gln	Glu	Thr	Arg 75	Ser	Gly	Ser	Arg	Arg 80	Thr	Ser	Pro	Gly	Val 85
15	Thr	Leu	Gln	Pro	Cyc 90	His	Val	Glu	Glu	Leu 95	Gly	Val	Ala	Gly	Asn 100
20	Ile	Val	Arg	His	Ile 105	Pro	Asp	Arg	Gly	Ala 110	Pro	Thr	Arg	Ala	Ser 115
	Glu	Pro	Val	Ser	Ala 120	Ala	Gly	His	Cys	Pro 125	Glu	Trp	Thr	Val	Val 130
25	Phe	Asp	Leu	Ser	Ala 135	Val	Glu	Pro	Ala	Glu 140	Arg	Pro	Ser	Arg	Ala 145
	Arg	Leu	Glu	Leu	Arg 150	Phe	Ala	Ala	Ala	Ala 155	Ala	Ala	Ala	Pro	Glu 160
30	Gly	Gly	Trp	Glu	Leu 165	Ser	Val	Ala	Gln	Ala 170	Gly	Gln	Gly	Ala	Gly 175
35	Ala	Asp	Pro	Gly	Pro 180	Val	Leu	Leu	Arg	Gln 185	Leu	Val	Pro	Ala	Leu 190
	Gly	Pro	Pro	Val	Arg 195	Ala	Glu	Leu	Leu	Gly 200	Ala	Ala	Trp	Ala	Arg 205
40	Asn	Ala	Ser	Trp	Pro 210	Arg	Ser	Leu	Arg	Leu 215	Ala	Leu	Ala	Leu	Arg 220
	Pro	Arg	Ala	Pro	Ala 225	Ala	Cys	Ala	Arg	Leu 230	Ala	Glu	Ala	Ser	Leu 235
45	Leu	Leu	Val	Thr	Leu 240	Asp	Pro	Arg	Leu	Cys 245	His	Pro	Leu	Ala	Arg 250
50	Pro	Arg	Arg	Asp	Ala 255	Glu	Pro	Val	Leu	Gly 260	Gly	Gly	Pro	Gly	Gly 265

	Ala	Cys	Arg	Ala	Arg 270	Arg	Leu	Tyr	Val	Ser 275	Phe	Arg	Glu	Val	G1y 280
5	Trp	His	Arg	Trp	Val 285	Ile	Arg	Pro	Arg	Gly 290	Phe	Leu	Ala	Asn	Tyr 295
	Cys	Gln	Gly	Gln	Cys 300	Ala	Leu	Pro	Val	Ala 305	Leu	Ser	Gly	Ser	Gly 310
10	Gly	Pro	Pro	Ala	Leu 315	Asn	His	Ala	Val	Leu 320	Arg	.Ala	Leu	Het	His 325
,	Ala	Ala	Ala	Pro	Gly 330	Ala	Ala	Asp	Leu	Pro 335	Cys	Cys	Val	Pro	Ala 340
15	Arg	Leu	Ser	Pro	Ile 345	Ser	Val	Leu	Phe	Phe 350	Asp	Asn	Ser	Asp	Asn 355
20	Val	Val	Leu	Arg	Gln 360	Tyr	Glu	Asp	Het	Val 365	Val	Asp	Glu	Cys	Gly 370
	Cys	Arg 372													

### What is claimed is:

1. A method for alleviating the tissue destructive effects associated with the inflammatory response to tissue injury in a mammal, the method comprising the step of:

providing to the injured tissue a therapeutically effective concentration of a morphogen sufficient to substantially inhibit or reduce the tissue damage resulting from said inflammatory response.

- 2. The method of claim 1 where said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering a therapeutically effective concentration of a morphogen to said mammal.
- 3. The method of claim 1 where said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.
- 4. The method of claim 1 wherein said step of providing a therapeutically effective concentration of a morphogen is conducted prior to reduction or interruption of blood flow to the tissue.

- 5. The method of claim 1 wherein said step of providing a therapeutically effective concentration of a morphogen is conducted after reduction or interruption of blood flow to the tissue and before reperfusion.
- 6. The method of claim 1 wherein said step of administering a therapeutically effective amount of a morphogen is conducted following ischemiareperfusion injury.
- 7. The method of claim 1 wherein said said step of administering a therapeutically effective amount of a morphogen is conducted following hyperoxia injury.
- 8. The method of claim 1 wherein said morphogen is provided to said tissue prior to said tissue injury.
- 9. The method of claim 1 wherein said step of providing a therapeutically effective concentration of a morphogen is conducted prior to ischemia-reperfusion injury.
- 10. The method of claim 1 wherein said tissue damage results from an abnormal immune response in said mammal.
- 11. The method of claim 1 wherein said tissue damage is associated with an inflammatory disease.
- 12. The method of claim 11 wherein said inflammatory disease is an autoimmune disease.

- 13. The method of claim 11 wherein said inflammatory disease comprises arthritis, psoriasis, dermatitis or diabetes.
- 14. The method of claim 13 wherein said arthritis is rhematoid, degenerative or psoriatic arthritis.
- 15. The method of claim 11 wherein said inflammatory disease comprises an airway inflammation in a mammal.
- 16. The method of claim 15 wherein said airway inflammation comprises chronic bronchitis, emphysema, idiopathic pulmonary fibrosis or asthma.
- 17. The method of claim 11 wherein inflammatory disease comprises a generalized acute inflammatory response.
- 18. The method of claim 17 wherein said inflammatory disease comprises adult respiratory distress syndrome.
- 19. The method of claim 1 wherein said tissue damage is to a transplanted organ or tissue.
- 20. A method for reducing tissue damage associated with ischemia-reperfusion injury in a human, the method comprising the step of:

providing to the injured tissue a therapeutic concentration of a morphogen sufficient to alleviate the damage associated with said injury.

- 21. A method for reducing the tissue damage associated with hyperoxia injury in a human, the method comprising the step of:
  - providing to the injured tissue a therapeutic concentration of a morphogen sufficient to alleviate the damage associated with said injury.
- 22. The method of claim 20 or 21 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering a therapeutically effective concentration of a morphogen to said mammal.
- 23. The method of claim 20 or 21 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.
- 24. The method of claim 1, 20 or 21 wherein said tissue is lung tissue, cardiac tissue, hepatic tissue or renal tissue.
- 25. The method of claim 6, 9 or 20 wherein said ischemic-reperfusion injury results from cardiac arrest, preliminary occlusion, arterial occlusion, coronary occlusion or occlusive stroke.

- 26. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vgl(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 27. The method of claim 26 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- 28. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 29. The method of claim 28 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 30. The method of claim 29 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 31. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).

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- 32. The method of claim 1, 20 or 21 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 33. A method for reducing the ischemic-reperfusion injury associated with the interruption of blood flow to an organ in a clinical procedure, the method comprising the step of providing a therapeutic concentration of a morphogen to said organ prior to the interruption of blood flow.
- 34. A method for reducing the tissue injury associated with the reduction or interruption of blood flow to an organ or tissue in a clinical procedure, the method comprising the step of providing a therapeutic concentration of a morphogen to said organ or tissue after the reduction or interruption of blood flow to said organ or tissue.
- 35. The method of claim 33 or 34 wherein said clinical procedure is a carotid enterectomy, a coronary artery bypass, a tissue grafting procedure, an organ transplant, or a fibrinolytic therapy.
- 36. The method of claim 1, 33 or 34 wherein said morphogen is administered parenterally.
- 37. The method of claim 1, 33 or 34 wherein said morphogen is administered prophylactically.

- 38. A pharmaceutical composition for use in alleviating the injury associated with tissue exposure to toxic oxygen concentrations comprising a therapeutically effective amount of a morphogen in admixture with a free oxygen radical inhibiting agent or an anticoagulent.
- 39. A pharmaceutical composition for topical administration comprising a therapeutically effective concentration of a morphogen in admixture with a dermatologically acceptable carrier.
- 40. A pharmaceutical composition for topical administration to a tissue comprising a therapeutically effective concentration of a morphogen dispersed in a biocompatible, non-irritating tissue surface adhesive.
- 41. The composition of claim 40 wherein said adhesieve comprises hydroxypropylcellulose.
- 42. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence sharing at least 70% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).
- The composition of claim 42 wherein said morphogen comprises an amino acid sequence sharing a last 80% homology with one of the sequences selected from the group consisting of: OP-1, OP-2, CBMP2, BMP3(fx), BMP5(fx), BMP6(fx), Vg1(fx), Vgr(fx), DPP(fx), GDF-1(fx) and 60A(fx).

- 44. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- .

- 45. The composition of claim 44 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 46. The method of claim 45 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 47. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).
- 48. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 49. A method of enhancing the viability of an organ or tissue to be transplanted in a mammal, the method of comprising the step of:

providing a therapeutically effective concentration of a morphogen to said tissue or organ to be transplanted.

- 50. The method of claim 49 wherein said therapeutically effective concentration is sufficient to substantially inhibit reperfusion injury to said tissue or organ.
- 51. The method of claim 49 wherein said morphogen is provided to said tissue or organ prior to reperfusion injury.
- 52. The method of claim 49 wherein said morphogen is provided to said tissue or organ prior to removal of said tissue or organ from the donor.
- 53. The method of claim 49 wherein said organ is placed in an organ preservation solution containing said morphogen or a morphogenstimulating agent after removal of said organ from the donor and prior to transplantation in the recipient.
- 54. The method of claim 49 wherein said organ is selected from the group consisting of lung, heart, kidney, liver or pancreas.
- 55. The method of claim 49 wherein said living tissue comprises skin, bone marrow or gastrointestinal mucosa tissue.

56. A method for protecting a living tissue or transplant organ from the tissue destructive effects associated with the inflammatory response in a mammal, the method comprising the step of:

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providing to said tissue or organ a therapeutically effective concentration of a morphogen.

57. A method of protecting a living tissue or transplanted organ from ischemia-reperfusion injury in a mammal, the method comprising the step of:

providing to said tissue or organ a therapeutically effective concentration of a morphogen, said concentration being sufficient to substantially inhibit or reduce the tissue damage associated with ischemia-reperfusion injury.

- 58. The method of claim 49, 56 or 57 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering a therapeutically effective concentration of a morphogen to said mammal.
- 59. The method of claim 49, 56 or 57 wherein said step of providing a therapeutically effective morphogen concentration to said injured tissue comprises the step of administering to said mammal an agent that stimulates in vivo a therapeutically effective concentration of an endogenous morphogen.

- 60. A composition useful as a living cell or living tissue preservation solution comprising:
  - a fluid formulation having as osmotic pressure substantially equivalent to the osmotic pressure of living mammalian cells in admixture with
  - a therapeutically effective concentration of a morphogen or morphogen-stimulating agent, said concentration being sufficient to protect living cell or tissue from the tissue destructive effects associated with the inflammatory response in a mammal when exposed to said cells or tissue.
- 61. The preservation solution of claim 60 wherein said therapeutically effective concentration is sufficient to substantially inhibit or reduce the tissue damage associated with ischemia-reperfusion injury.
- 62. The preservation solution of claim 60 wherein said formulation further comprises a sugar.
- 63. The preservation solution of claim 60 wherein said formulation further comprises an anticoagulant or a free oxygen radical inhibiting agent.
- 64. The invention of claim 49, 56, 57 or 60 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).

- 65. A composition useful in a treatment method to alleviate tissue damage associated with the inflammatory response in a mammal, the composition comprising a therapeutically effective concentration of a morphogen or morphogenstimulating agent.
- 66. The composition of claim 65 wherein said tissue damage is associated with ischemia-reperfusion injury or hyperoxia injury.
- 67. The composition of claim 65 wherein said tissue damage is to lung, cardiac, renalor hepatic tissue.
- 68. The composition of claim 65 wherein said tissue damage is to a transplanted organ or tissue.

### AMENDED CLAIMS

[received by the International Bureau on 10 February 1993 (10.02.93); original claims 46 and 49 amended; remaining claims unchanged (1 page)]

- 44. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence having greater than 60% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 45. The composition of claim 44 wherein said morphogen comprises an amino acid sequence having greater than 65% amino acid identity with the sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1).
- 46. The composition of claim 45 wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOP1), including allelic and species variants thereof.
- 47. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by Generic Sequences 1, 2, 3, 4, 5 or 6 (Seq. ID Nos. 1, 2, 3, 4, 30 or 31).
- 48. The composition of claim 38, 39 or 40 wherein said morphogen comprises an amino acid sequence defined by OPX (Seq. ID No. 29).
- 49. A method of enhancing the viability of an organ or tissue to be transplanted in a mammal, the method comprising the step of:

providing a therapeutically effective concentration of a morphogen to said tissue or organ to be transplanted.

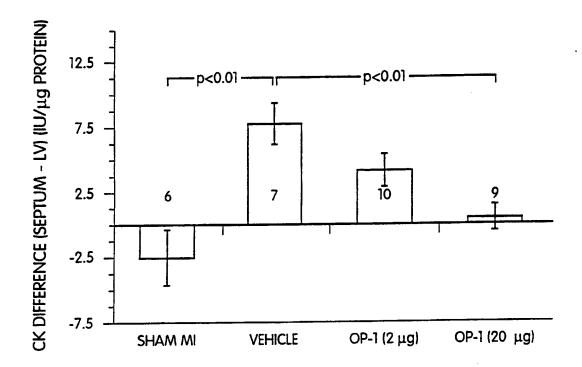


Fig. 1

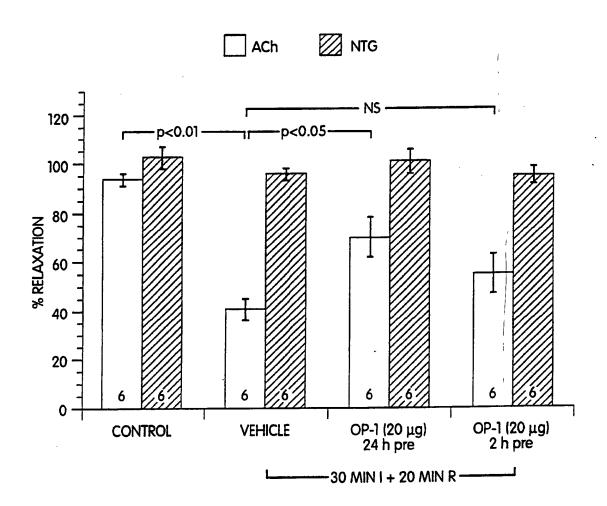


Fig 2

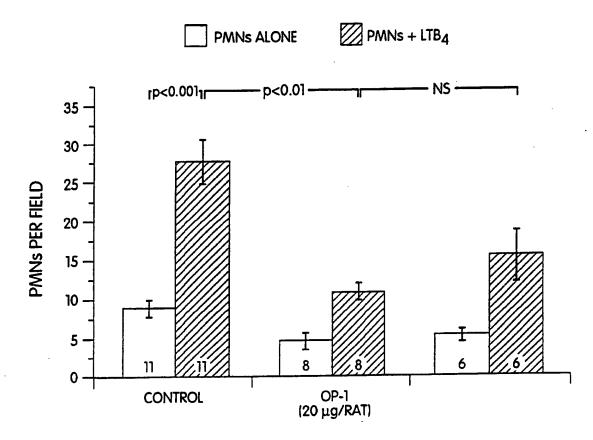
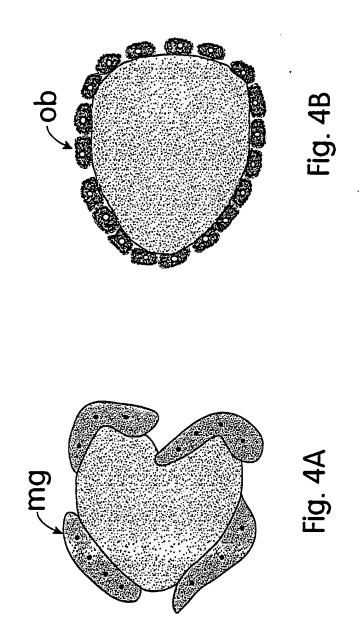
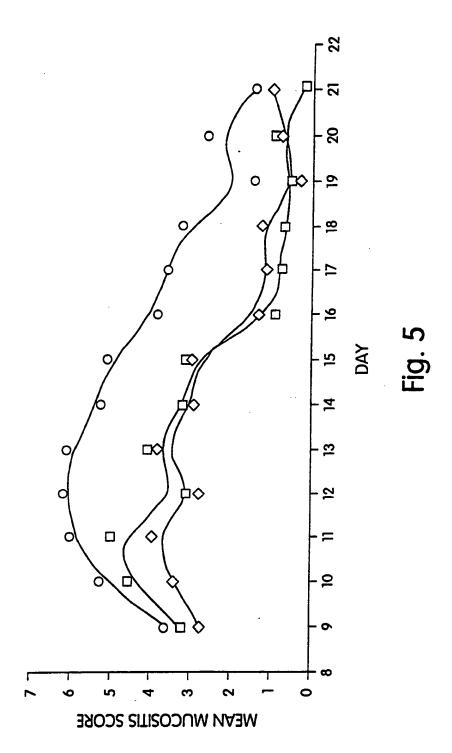


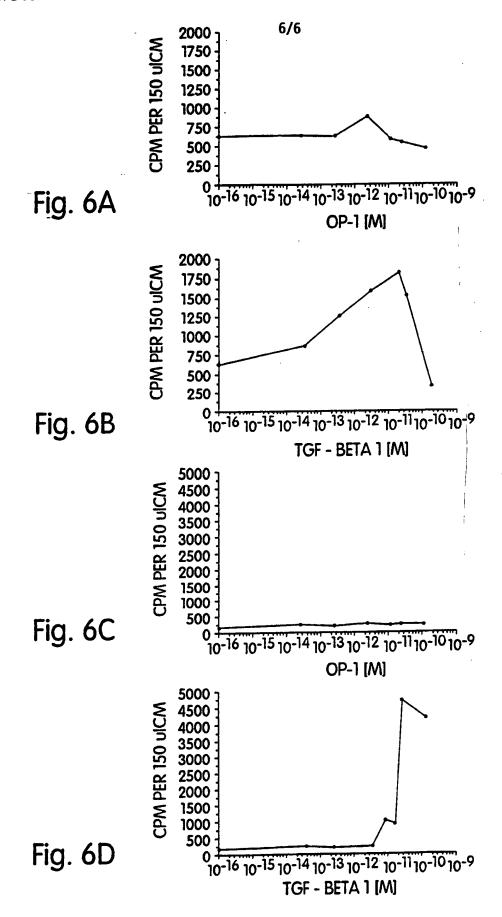
Fig. 3



SUBSTITUTE SHEET



SUBSTITUTE SHEET



## **SUBSTITUTE SHEET**

May year

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) According to International Patent Classification (IPC) or to both National Classification and IPC A01N1/02 Int.C1. 5 A61K37/02; II. FIELDS SEARCHED Minimum Documentation Searched? Classification Symbols Classification System C07K · A61K ; Int.Cl. 5 Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched® III. DOCUMENTS CONSIDERED TO BE RELEVANT? Relevant to Claim No.13. Citation of Document, 11 with indication, where appropriate, of the relevant passages 12 Category o 1-2,4-9, vol. 249, no. 4964, 6 July 1990, LANCASTER, PA US 20-22, 25, 33-37, pages 61 - 64 57-58. LEFER A.M. ET AL 'Mediation of 65-68 cardioprotection by Transforming growth factor-beta' cited in the application see the whole document PROCEEDINGS OF THE NATIONAL ACADEMY OF 1-2, 10-19. SCIENCES OF USA. 36-37, vol. 88, no. 7, April 1991, WASHINGTON US 56,58,65 pages 2918 - 2921 KURUVILLA, A.P. 'Protective effect of transforming growth factor betal on experimental autoimmune diseases in mice1 cited in the application see the whole document "T" later document published after the international filing date <sup>o</sup> Special categories of cited documents: 10 or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to earlier document but published on or after the international filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) involve an inventive step "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-"O" document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled other means document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family IV. CERTIFICATION Date of Mailing of this International Search Report Date of the Actual Completion of the International Search 19 NOVEMBER 1992 Signature of Authorized Officer
FERIANDEZ Y BRA F. International Searching Authority **EUROPEAN PATENT OFFICE** 

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III. DOCUME	INTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)	Del
Category °	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
x	EP,A,O 269 408 (GENENTECH INC.) 1 June 1988	1-2, 10-19, 36-37, 56,58,65
	see the whole document	
X	WO,A,9 000 900 (AMGEN INC.) 8 February 1990	1-2, 10-19, 36-37, 56,58,65
	see the whole document	
E	WO,A,9 215 323 (CREATIVE BIOMOLECULES) 17 September 1992	1-6, 8-12, 15-17,
	see page 6, line 1 - page 7, line 27	22-37, 56-59, 64-67
	see page 77 - page 119	
x	WO,A,9 105 802 (CREATIVE BIOMOLECULES) 2 May 1991	1-2,- 10-13, 26-32, 36-37,65
	see page 20, line 19 - line 31 see page 59 - page 80	30 3/,03
Х,Р	WO,A,9 207 073 (CREATIVE BIOMOLECULES) 30 April 1992	1-2, 10-13, 26-29, 31-32, 36-37,65
	see page 11, line 3 - line 15 see page 41 - page 60	
X	WO,A,8 909 788 (CREATIVE BIOMOLECULES) 19 October 1989 cited in the application	1-2, 10-13, 26-29, 31-32, 36-37,65
	see page 6 - page 15	·
<b>A</b>	EMBO JOURNAL vol. 9, no. 7, 1990, EYNSHAM, OXFORD GB pages 2085 - 2093 ÖZKAYNAK, E. ET AL 'OP-1 cDNA encodes an osteogenic protein in the TGF-beta family' see the whole document	1-68

# INTERNATIONAL SEARCH REPORT

It ational application No.

PCT/US 92/07358

Par -	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
Box I	
This int	ernational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
]	see annex
2. X	Claims Nos.:  because they relate to parts of the international application that do not comply with the prescribed requirements to such an examination and a carried out, specifically:
]	see annex
3.	Claims Nos.:  because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
	1 Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This I	nternational Searching Authority found multiple inventions in this international application, as follows:
1	
1	
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
	Or may managed and a second and
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. [	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
	,
_	ark on Protest  The additional search fees were accompanied by the applicant's protest.
Kem	No protest accompanied the payment of additional search fees.

### FURTHER INFORMATION CONTINUED FROM PCT/ISA/210

Remark: Although claims 1-37,49-52 (partially, when the method is carried out in vivo), 54-57 (partially, when the method is carried out in vivo), 58 - 59.64 (partially, according to the method of claims 49,56 or 57) are directed to a method of treatment of the human or animal body the search has been carried out and based on the alleged effects of the composition.

OBSCURITIES, INCONSISTENCIES, CONTRADICTIONS, LACK OF CONCISENESS; LACK OF READY COMPREHENSIBILITY)

(ART. 6 PCT)

### **REASON:**

- 1. Claim 46 has been understood as being dependant of claim 45. Therefore claim 46 should read: "The composition of claim 45, wherein said morphogen comprises an amino acid sequence defined by residues 43-139 of Seq. ID No. 5 (hOPI), including allelic and species variants thereof.
- 2. In view of the extremely large number of compounds used in the methods and compositions of claims 26-29, 31 (in as far as seq. ID 1 to 4 and 30-31), 42-45,47 (in as far as seq. ID 1 to 4 and 30-31), 64, the search division considers that it is not economically reasonable to draw up a search report for the methods using, or the compositions comprising all the compounds defined in the claims. The search has therefore been limited, on the basis of the examples and claims, to the methods using, or the compositions comprising the seq. ID no. 5 to 29, 32 and 33 (Art. 17 (2) (a)(ii) and (b) PCT.
- 3. The term "morphogen" is not concise.

Therefor, and for the same reaons as given in paragraph 2 above, it has been understood as being one of the proteins defines in seq. ID 5 to 29, 32 or 33.

(Art. 6 PCT and Art. 17/2)(a)(ii) and (b) PCT)

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#### ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO. US 9207358 SA 64364

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 19/11/92.

Patent document cited in search report	Publication date	Patent family member(s)		Publication date	
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		EP-A-	0411105	06-02-91	
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		WO-A-	9010018	07 <b>-</b> 09-90	
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